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(54) Title: USE OF NEONICODINOIDS ON TRANSGENIC PLANTS

(57) Abstract

There is now described a method of controlling pests with nitroimino– or nitroguanidino–compounds; more specifically a method of controlling pests in and on transgenic crops of useful plants, such as, for example, in crops of maize, cereals, soya beans, tomatoes, cotton, potatoes, rice and mustard, with a nitroimino– or nitroguanidino–compound, especially with thiamethoxam, characterized in that a pestical composition comprising a nitroimino– or nitroguanidino– compound in free form or in agrochemically useful salt form and at least one auxiliary is applied to the pests or their environment, in particular to the crop plant itself.

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USE OF NEONICODINOIDS ON TRANSGENIC PLANTS

The present invention relates to a method of controlling pests with a nitroimino- or nitroguanidino-compound, especially thiamethoxam; more specifically to a novel method of controlling pests in and on transgenic crops of useful plants with a nitroimino- or nitroguanidino-compound.

Certain pest control methods are proposed in the literature. However, these methods are not fully satisfactory in the field of pest control, which is why there is a demand for providing further methods for controlling and combating pests, in particular insects and representatives of the order Acarina, or for protecting plants, especially crop plants. This object is achieved according to the invention by providing the present method.

The present invention therefore relates to a method of controlling pests in crops of transgenic useful plants, such as, for example, in crops of maize, cereals, soya beans, tomatoes, cotton, potatoes, rice and mustard, characterized in that a pesticidal composition comprising a nitroimino- or nitroguanidino-compound, especially thiamethoxam, imidacloprid, Ti-435 or thiacloprid in free form or in agrochemically useful salt form and at least one auxiliary is applied to the pests or their environment, in particular to the crop plant itself; to the use of the composition in question and to propagation material of transgenic plants which has been treated with it.

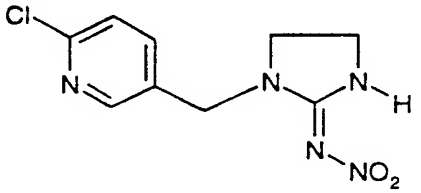
Surprisingly, it has now emerged that the use of a nitroimino- or nitroguanidino-compound compound for controlling pests on transgenic useful plants which contain - for instance - one or more genes expressing a pesticidally, particularly insecticidally, acaricidally, nematocidally or fugicidally active ingredient, or which are tolerant against herbicides or resistent against the attack of fungi, has a synergistic effect. It is highly surprising that the use of a nitroimino- or nitroguanidino-compound in combination with a transgenic plant exceeds the additive effect, to be expected in principle, on the pests to be controlled and thus extends the range of action of the nitroimino- or nitroguanidino-compound and of the active principle expressed by the transgenic plant in particular in two respects:

In particular, it has been found, surprisingly, that within the scope of invention the pesticidal activity of a nitroimino- or nitroguanidino-compound in combination with the effect expressed by the transgenic useful plant, is not only additive in comparison with the pesticidal activities of the nitroimino- or nitroguanidino-compound alone and of the

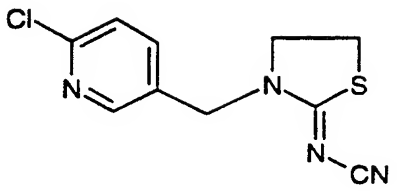
transgenic crop plant alone, as can generally be expected, but that a synergistic effect is present. The term "synergistic", however, is in no way to be understood in this connection as being restricted to the pesticidal activity, but the term also refers to other advantageous properties of the method according to the invention compared with the nitroimino- or nitroguanidino-compound and the transgenic useful plant alone. Examples of such advantageous properties which may be mentioned are: extension of the pesticidal spectrum of action to other pests, for example to resistant strains; reduction in the application rate of the nitroimino- or nitroguanidino-compound, or sufficient control of the pests with the aid of the compositions according to the invention even at an application rate of the nitroimino- or nitroguanidino-compound alone and the transgenic useful plant alone are entirely ineffective; enhanced crop safety; improved quality of produce such as higher content of nutrient or oil, better fiber quality, enhanced shelf life, reduced content of toxic products such as mycotoxins, reduced content of residues or unfavorable constituents of any kind or better digestability; improved tolerance to unfavorable temperatures, draughts or salt content of water; enhanced assimilation rates such as nutrient uptake, water uptake and photosynthesis; favorable crop properties such as altered leaf aerea, reduced vegetative growth, increased yields, favorable seed shape/seed thickness or germination properties, altered colonialisation by saprophytes or epiphytes, reduction of senescense, improved phytoalexin production, improved of accelerated ripening, flower set increase, reduced boll fall and shattering, better attraction to beneficials and predators, increased pollination, reduced attraction to birds; or other advantages known to those skilled in the art.

Nitroimino- and nitroguanidino-cpmpounds, such as thiamethoxam (5-(2-Chlorthiazol-5-ylmethyl)-3-methyl-4-nitroimino-perhydro-1,3,5-oxadiazin), are known from EP-A-0'580'553. Within the scope of invention thiamethoxam is preferred.

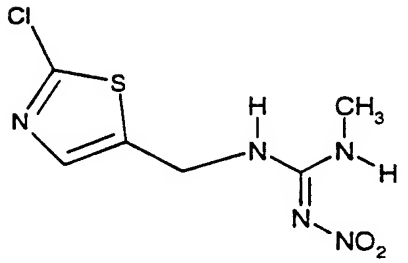
Also preferred within the scope of invention is imidacloprid of the formula



, known from The Pesticide Manual, 10thEd. (1991), The British Crop Protection Council, London, page 591;

also preferred is Thiacloprid of the formula  , known from

EP-A-235'725;

also preferred is the compound of the formula  , known as

Ti-435 (Clothiamidin) from EP-A-376'279

The agrochemically compatible salts of the nitroimino- or nitroguanidino-compounds are, for example, acid addition salts of inorganic and organic acids, in particular of hydrochloric acid, hydrobromic acid, sulfuric acid, nitric acid, perchloric acid, phosphoric acid, formic acid, acetic acid, trifluoroacetic acid, oxalic acid, malonic acid, toluenesulfonic acid or benzoic acid. Preferred within the scope of the present invention is a composition known per se which comprises, as active ingredient, thiamethoxam and imidacloprid, each in the free form, especially thiamethoxam.

The transgenic plants used according to the invention are plants, or propagation material thereof, which are transformed by means of recombinant DNA technology in such a way that they are - for instance - capable of synthesizing selectively acting toxins as are known, for example, from toxin-producinginvertebrates, especially of the phylum Arthropoda, as can be obtained from Bacillus thuringiensis strains; or as are known from plants, such as lectins; or in the alternative capable of expressing a herbicidal or fungicidal resistance. Examples of such toxins, or transgenic plants which are capable of synthesizing such toxins, have been disclosed, for example, in EP-A-0 374 753, WO 93/07278, WO 95/34656, EP-A-0 427 529 and EP-A-451 878 and are incorporated by reference in the present application.

The methods for generating such transgenic plants are widely known to those skilled in the art and described, for example, in the publications mentioned above.

The toxins which can be expressed by such transgenic plants include, for example, toxins, such as proteins which have insecticidal properties and which are expressed by transgenic plants, for example Bacillus cereus proteins or Bacillus popliae proteins; or Bacillus thuringiensis endotoxins (B.t.), such as CryIA(a), CryIA(b), CryIA(c), CryIIA, CryIIIA, CryIIIB2 orCytA; VIP1; VIP2; VIP3; or insecticidal proteins of bacteria colonising nematodes like Photorhabdus spp or Xenorhabdus spp such as Photorhabdus luminescens, Xenorhabdus nematophilus etc.; proteinase inhibitors, such as trypsin inhibitors, serine protease inhibitors, patatin, cystatin, papain inhibitors; ribosome-inactivating proteins (RIP), such as ricin, maize RIP, abrin, luffin, saporin or bryodin; plant lectins such as pea lectins, barley lectins or snowdrop lectins; or agglutinins; toxins produced by animals, such as scorpion toxins, spider venoms, wasp venoms and other insect-specific neurotoxins; steroid metabolism enzymes, such as 3-hydroxysteroid oxidase, ecdysteroid UDP-glycosyl transferase, cholesterol oxidases, ecdysone inhibitors, HMG-COAreductase, ion channel blockers such as sodium and calcium, juvenile hormone esterase, diuretic hormone receptors, stilbene synthase, bibenzyl synthase, chitinases and glucanases.

Examples of known transgenic plants which comprise one or more genes which encode insecticidal resistance and express one or more toxins are the following: KnockOut® (maize), YieldGard® (maize); NuCOTN 33B® (cotton), Bollgard® (cotton), NewLeaf® (potatoes), NatureGard® and Protecta®.

The following tables comprise further examples of targets and principles and crop phenotypes of transgenic crops which show tolerance against pests mainly insects, mites, nematodes, virus, bacteria and diseases or are tolerant to specific herbicides or classes of herbicides.

Table A1: Crop: Maize

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
Acetolactate synthase (ALS)	Sulfonylureas, Imidazolinones, Triazolopyrimidines, Pyrimidyloxybenzoates, Phtalides
AcetylCoA Carboxylase (ACCase)	Aryloxyphenoxyalkanecarboxylic acids, cyclohexanediones

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
Hydroxyphenylpyruvate dioxygenase (HPPD)	Isoxazoles such as Isoxaflutol or Isoxachlortol, Triones such as mesotrione or sulcotrione
Phosphinothricin acetyl transferase	Phosphinothricin
O-Methyl transferase	altered lignin levels
Glutamine synthetase	Glufosinate, Bialaphos
Adenylosuccinate Lyase (ADSL)	Inhibitors of IMP and AMP synthesis
Adenylosuccinate Synthase	Inhibitors of adenylosuccinate synthesis
Anthranilate Synthase	Inhibitors of tryptophan synthesis and catabolism
Nitrilase	3,5-dihalo-4-hydroxy-benzonitriles such as Bromoxynil and Ioxinyl
5-Enolpyruvyl-3phosphoshikimate Synthase (EPSPS)	Glyphosate or sulfosate
Glyphosate oxidoreductase	Glyphosate or sulfosate
Protoporphyrinogen oxidase (PROTOX)	Diphenylethers, cyclic imides, phenylpyrazoles, pyridin derivatives, phenopylate, oxadiazoles etc.
Cytochrome P450 eg. P450 SU1	Xenobiotics and herbicides such as Sulfonylureas
Dimboa biosynthesis (Bx1 gene)	Helminthosporium turcicum, Rhopalosiphum maydis, Diplodia maydis, Ostrinia nubilalis, lepidoptera sp.
CMIII (small basic maize seed peptide	plant pathogenes eg. fusarium, alternaria, sclerotina
Corn- SAFP (zeamatin)	plant pathogenes eg. fusarium, alternaria, sclerotina, rhizoctonia, chaetomium,phycomyces
Hm1 gene	Cochliobulus
Chitinases	plant pathogenes
Glucanases	plant pathogenes
Coat proteins	viruses such as maize dwarf mosaic

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
	virus, maize chlorotic dwarf virus
Bacillus thuringiensis toxins, VIP 3, Bacillus cereus toxins, Photorabdus and Xenorhabdus toxins	lepidoptera, coleoptera, diptera, nematodes, eg. ostrinia nubilalis, heliothis zea, armyworms eg. spodoptera frugiperda, corn rootworms, sesamia sp., black cutworm, asian corn borer,weevils
3- Hydroxysteroid oxidase	lepidoptera, coleoptera, diptera, nematodes, eg. ostrinia nubilalis, heliothis zea, armyworms eg. spodoptera frugiperda, corn rootworms, sesamia sp., black cutworm, asian corn borer, weevils
Peroxidase	lepidoptera, coleoptera, diptera, nematodes, eg. ostrinia nubilalis, heliothis zea, armyworms eg. spodoptera frugiperda, corn rootworms, sesamia sp., black cutworm, asian corn borer, weevils
Aminopeptidase inhibitors eg. Leucine aminopeptidase inhibitor (LAPI)	lepidoptera, coleoptera, diptera, nematodes, eg. ostrinia nubilalis, heliothis zea, armyworms eg. spodoptera frugiperda, corn rootworms, sesamia sp., black cutworm, asian corn borer, weevils
Limonene synthase	corn rootworms
Lectines	lepidoptera, coleoptera, diptera, nematodes, eg. ostrinia nubilalis, heliothis zea, armyworms eg. spodoptera frugiperda, corn rootworms, sesamia sp., black cutworm, asian corn borer, weevils

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
Protease Inhibitors eg. cystatin, patatin, virgiferin, CPTI	weevils, corn rootworm
ribosome inactivating protein	lepidoptera, coleoptera, diptera, nematodes, eg. ostrinia nubilalis, heliothis zea, armyworms eg. spodoptera frugiperda, corn rootworms, sesamia sp., black cutworm, asian corn borer, weevils
maize 5C9 polypeptide	lepidoptera, coleoptera, diptera, nematodes, eg. ostrinia nubilalis, heliothis zea, armyworms eg. spodoptera frugiperda, corn rootworms, sesamia sp., black cutworm, asian corn borer, weevils
HMG-CoA reductase	lepidoptera, coleoptera, diptera, nematodes, eg. ostrinia nubilalis, heliothis zea, armyworms eg. spodoptera frugiperda, corn rootworms, sesamia sp., black cutworm, asian corn borer, weevils

Table A2: Crop Wheat

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
Acetolactate synthase (ALS)	Sulfonylureas, Imidazolinones, Triazolopyrimidines, Pyrimidyloxybenzoates, Phtalides
AcetylCoA Carboxylase (ACCase)	Aryloxyphenoxyalkanecarboxylic acids, cyclohexanediones
Hydroxyphenylpyruvate dioxygenase (HPPD)	Isoxazoles such as Isoxaflutol or Isoxachlortol, Triones such as mesotrione or sulcotrione

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
Phosphinothricin acetyl transferase	Phosphinothricin
O-Methyl transferase	altered lignin levels
Glutamine synthetase	Glufosinate, Bialaphos
Adenylosuccinate Lyase (ADSL)	Inhibitors of IMP and AMP synthesis
Adenylosuccinate Synthase	Inhibitors of adenylosuccinate synthesis
Anthranilate Synthase	Inhibitors of tryptophan synthesis and catabolism
Nitrilase	3,5-dihalo-4-hydroxy-benzonitriles such as Bromoxynil and Ioxinyl
5-Enolpyruvyl-3phosphoshikimate Synthase (EPSPS)	Glyphosate or sulfosate
Glyphosate oxidoreductase	Glyphosate or sulfosate
Protoporphyrinogen oxidase (PROTOX)	Diphenylethers, cyclic imides, phenylpyrazoles, pyridin derivatives, phenopylate, oxadiazoles etc.
Cytochrome P450 eg. P450 SU1	Xenobiotics and herbicides such as Sulfonylureas
Antifungal polypeptide AlyAFP	plant pathogenes eg septoria and fusarioum
glucose oxidase	plant pathogenes eg. fusarium, septoria
pyrrolnitrin synthesis genes	plant pathogenes eg. fusarium, septoria
serine/threonine kinases	plant pathogenes eg. fusarium, septoria and other diseases
Hypersensitive response eliciting polypeptide	plant pathogenes eg. fusarium, septoria and other diseases
Systemic acquires resistance (SAR) genes	viral, bacterial, fungal, nematodal pathogens
Chitinases	plant pathogenes
Glucanases	plant pathogenes
double stranded ribonuclease	viruses such as BYDV and MSMV
Coat proteins	viruses such as BYDV and MSMV
Bacillus thuringiensis toxins, VIP 3,	lepidoptera, coleoptera, diptera,

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
Bacillus cereus toxins, Photorabdus and Xenorhabdus toxins	nematodes,
3- Hydroxysteroid oxidase	lepidoptera, coleoptera, diptera, nematodes,
Peroxidase	lepidoptera, coleoptera, diptera, nematodes,
Aminopeptidase inhibitors eg. Leucine aminopeptidase inhibitor	lepidoptera, coleoptera, diptera, nematodes,
Lectines	lepidoptera, coleoptera, diptera, nematodes, aphids
Protease Inhibitors eg. cystatin, patatin, virgiferin, CPTI	lepidoptera, coleoptera, diptera, nematodes, aphids
ribosome inactivating protein	lepidoptera, coleoptera, diptera, nematodes, aphids
HMG-CoA reductase	lepidoptera, coleoptera, diptera, nematodes, eg. ostrinia nubilalis, heliothis zea, armyworms eg. spodoptera frugiperda, corn rootworms, sesamia sp., black cutworm, asian corn borer, weevils

Table A3: Crop Barley

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
Acetolactate synthase (ALS)	Sulfonylureas, Imidazolinones, Triazolopyrimidines, Pyrimidyloxybenzoates, Phtalides
AcetylCoA Carboxylase (ACCase)	Aryloxyphenoxyalkanecarboxylic acids, cyclohexanediones
Hydroxyphenylpyruvate dioxygenase (HPPD)	Isoxazoles such as Isoxaflutol or Isoxachlortol, Triones such as mesotrione or sulcotrione

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
Phosphinothricin acetyl transferase	Phosphinothricin
O-Methyl transferase	altered lignin levels
Glutamine synthetase	Glufosinate, Bialaphos
Adenylosuccinate Lyase (ADSL)	Inhibitors of IMP and AMP synthesis
Adenylosuccinate Synthase	Inhibitors of adenylosuccinate synthesis
Anthranilate Synthase	Inhibitors of tryptophan synthesis and catabolism
Nitrilase	3,5-dihalo-4-hydroxy-benzonitriles such as Bromoxynil and Ioxinyl
5-Enolpyruvyl-3phosphoshikimate Synthase (EPSPS)	Glyphosate or sulfosate
Glyphosate oxidoreductase	Glyphosate or sulfosate
Protoporphyrinogen oxidase (PROTOX)	Diphenylethers, cyclic imides, phenylpyrazoles, pyridin derivatives, phenopylate, oxadiazoles etc.
Cytochrome P450 eg. P450 SU1	Xenobiotics and herbicides such as Sulfonylureas
Antifungal polypeptide AlyAFP	plant pathogenes eg septoria and fusarioum
glucose oxidase	plant pathogenes eg. fusarium, septoria
pyrrolnitrin synthesis genes	plant pathogenes eg. fusarium, septoria
serine/threonine kinases	plant pathogenes eg. fusarium, septoria and other diseases
Hypersensitive response eliciting polypeptide	plant pathogenes eg. fusarium, septoria and other diseases
Systemic acquires resistance (SAR) genes	viral, bacterial, fungal, nematodal pathogens
Chitinases	plant pathogenes
Glucanases	plant pathogenes
double stranded ribonuclease	viruses such as BYDV and MSMV
Coat proteins	viruses such as BYDV and MSMV
Bacillus thuringiensis toxins, VIP 3,	lepidoptera, coleoptera, diptera,

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
Bacillus cereus toxins, Photorabdus and Xenorhabdus toxins	nematodes,
3- Hydroxysteroid oxidase	lepidoptera, coleoptera, diptera, nematodes,
Peroxidase	lepidoptera, coleoptera, diptera, nematodes,
Aminopeptidase inhibitors eg. Leucine aminopeptidase inhibitor	lepidoptera, coleoptera, diptera, nematodes,
Lectines	lepidoptera, coleoptera, diptera, nematodes, aphids
Protease Inhibitors eg. cystatin, patatin, virgiferin, CPTI	lepidoptera, coleoptera, diptera, nematodes, aphids
ribosome inactivating protein	lepidoptera, coleoptera, diptera, nematodes, aphids
HMG-CoA reductase	lepidoptera, coleoptera, diptera, nematodes, aphids

Table A4: Crop Rice

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
Acetolactate synthase (ALS)	Sulfonylureas, Imidazolinones, Triazolopyrimidines, Pyrimidyloxybenzoates, Phtalides
AcetylCoA Carboxylase (ACCase)	Aryloxyphenoxyalkanecarboxylic acids, cyclohexanediones
Hydroxyphenylpyruvate dioxygenase (HPPD)	Isoxazoles such as Isoxaflutol or Isoxachlortol, Triones such as mesotrione or sulcotrione
Phosphinothricin acetyl transferase	Phosphinothricin
O-Methyl transferase	altered lignin levels
Glutamine synthetase	Glufosinate, Bialaphos
Adenylosuccinate Lyase (ADSL)	Inhibitors of IMP and AMP synthesis

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
Adenylosuccinate Synthase	Inhibitors of adenylosuccinate synthesis
Anthranilate Synthase	Inhibitors of tryptophan synthesis and catabolism
Nitrilase	3,5-dihalo-4-hydroxy-benzonitriles such as Bromoxynil and Ioxinyl
5-Enolpyruvyl-3phosphoshikimate Synthase (EPSPS)	Glyphosate or sulfosate
Glyphosate oxidoreductase	Glyphosate or sulfosate
Protoporphyrinogen oxidase (PROTOX)	Diphenylethers, cyclic imides, phenylpyrazoles, pyridin derivatives, phenopylate, oxadiazoles etc.
Cytochrome P450 eg. P450 SU1	Xenobiotics and herbicides such as Sulfonylureas
Antifungal polypeptide AlyAFP	plant pathogenes
glucose oxidase	plant pathogenes
pyrrolnitrin synthesis genes	plant pathogenes
serine/threonine kinases	plant pathogenes
Phenylalanine ammonia lyase (PAL)	plant pathogenes eg bacterial leaf blight and rice blast, inducible
phytoalexins	plant pathogenes eg bacterial leaf blight and rice blast
B-1,3-glucanase antisense	plant pathogenes eg bacterial leaf blight and rice blast
receptor kinase	plant pathogenes eg bacterial leaf blight and rice blast
Hypersensitive response eliciting polypeptide	plant pathogenes
Systemic acquires resistance (SAR) genes	viral, bacterial, fungal, nematodal pathogens
Chitinases	plant pathogenes eg bacterial leaf blight and rice blast
Glucanases	plant pathogenes

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
double stranded ribonuclease	viruses such as BYDV and MSMV
Coat proteins	viruses such as BYDV and MSMV
Bacillus thuringiensis toxins, VIP 3, Bacillus cereus toxins, Photorabdus and Xenorhabdus toxins	lepidoptera eg. stemborer, coleoptera eg rice water weevil, diptera, rice hoppers eg brown rice hopper
3- Hydroxysteroid oxidase	lepidoptera eg. stemborer, coleoptera eg rice water weevil, diptera, rice hoppers eg brown rice hopper
Peroxidase	lepidoptera eg. stemborer, coleoptera eg rice water weevil, diptera, rice hoppers eg brown rice hopper
Aminopeptidase inhibitors eg. Leucine aminopeptidase inhibitor	lepidoptera eg. stemborer, coleoptera eg rice water weevil, diptera, rice hoppers eg brown rice hopper
Lectines	lepidoptera eg. stemborer, coleoptera eg rice water weevil, diptera, rice hoppers eg brown rice hopper
Protease Inhibitors,	lepidoptera eg. stemborer, coleoptera eg rice water weevil, diptera, rice hoppers eg brown rice hopper
ribosome inactivating protein	lepidoptera eg. stemborer, coleoptera eg rice water weevil, diptera, rice hoppers eg brown rice hopper
HMG-CoA reductase	lepidoptera eg. stemborer, coleoptera eg rice water weevil, diptera, rice hoppers eg brown rice hopper

Table A5: Crop Soya

Effected target or expressed principle(s)	Crop phenotype / Tolerance to

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
Acetolactate synthase (ALS)	Sulfonylureas, Imidazolinones, Triazolopyrimidines, Pyrimidyloxybenzoates, Phtalides
AcetylCoA Carboxylase (ACCase)	Aryloxyphenoxyalkanecarboxylic acids, cyclohexanediones
Hydroxyphenylpyruvate dioxygenase (HPPD)	Isoxazoles such as Isoxaflutol or Isoxachlortol, Triones such as mesotrione or sulcotrione
Phosphinothricin acetyl transferase	Phosphinothricin
O-Methyl transferase	altered lignin levels
Glutamine synthetase	Glufosinate, Bialaphos
Adenylosuccinate Lyase (ADSL)	Inhibitors of IMP and AMP synthesis
Adenylosuccinate Synthase	Inhibitors of adenylosuccinate synthesis
Anthranilate Synthase	Inhibitors of tryptophan synthesis and catabolism
Nitrilase	3,5-dihalo-4-hydroxy-benzonitriles such as Bromoxynil and Ioxinyl
5-Enolpyruvyl-3phosphoshikimate Synthase (EPSPS)	Glyphosate or sulfosate
Glyphosate oxidoreductase	Glyphosate or sulfosate
Protoporphyrinogen oxidase (PROTOX)	Diphenylethers, cyclic imides, phenylpyrazoles, pyridin derivatives, phenopylate, oxadiazoles etc.
Cytochrome P450 eg. P450 SU1 or selection	Xenobiotics and herbicides such as Sulfonylureas
Antifungal polypeptide AlyAFP	bacterial and fungal pathogens such as fusarium, sclerotinia, stemrot
oxalate oxidase	bacterial and fungal pathogens such as fusarium, sclerotinia, stemrot
glucose oxidase	bacterial and fungal pathogens such as fusarium, sclerotinia, stemrot
pyrrolnitrin synthesis genes	bacterial and fungal pathogens such as

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
	fusarium, sclerotinia, stemrot
serine/threonine kinases	bacterial and fungal pathogens such as fusarium, sclerotinia, stemrot
Phenylalanine ammonia lyase (PAL)	bacterial and fungal pathogens such as fusarium, sclerotinia, stemrot
phytoalexins	plant pathogenes eg bacterial leaf blight and rice blast
B-1,3-glucanase antisense	plant pathogenes eg bacterial leaf blight and rice blast
receptor kinase	bacterial and fungal pathogens such as fusarium, sclerotinia, stemrot
Hypersensitive response eliciting polypeptide	plant pathogenes
Systemic acquires resistance (SAR) genes	viral, bacterial, fungal, nematodal pathogens
Chitinases	bacterial and fungal pathogens such as fusarium, sclerotinia, stemrot
Glucanases	bacterial and fungal pathogens such as fusarium, sclerotinia, stemrot
double stranded ribonuclease	viruses such as BPMV and SbMV
Coat proteins	viruses such as BYDV and MSMV
Bacillus thuringiensis toxins, VIP 3, Bacillus cereus toxins, Photorabdus and Xenorhabdus toxins	lepidoptera, coleoptera, aphids
3- Hydroxysteroid oxidase	lepidoptera, coleoptera, aphids
Peroxidase	lepidoptera, coleoptera, aphids
Aminopeptidase inhibitors eg. Leucine aminopeptidase inhibitor	lepidoptera, coleoptera, aphids
Lectines	lepidoptera, coleoptera, aphids
Protease Inhibitors eg virgiferin	lepidoptera, coleoptera, aphids

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
ribosome inactivating protein	lepidoptera, coleoptera, aphids
HMG-CoA reductase	lepidoptera, coleoptera, aphids
Barnase	nematodes eg root knot nematodes and cyst nematodes
Cyst nematode hatching stimulus	cyst nematodes
Antifeeding principles	nematodes eg root knot nematodes and cyst nematodes

Table A6: Crop Potatoes

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
Acetolactate synthase (ALS)	Sulfonylureas, Imidazolinones, Triazolopyrimidines, Pyrimidyloxybenzoates, Phtalides
AcetylCoA Carboxylase (ACCase)	Aryloxyphenoxyalkanecarboxylic acids, cyclohexanediones
Hydroxyphenylpyruvate dioxygenase (HPPD)	Isoxazoles such as Isoxaflutol or Isoxachlortol, Triones such as mesotrione or sulcotrione
Phosphinothricin acetyl transferase	Phosphinothricin
O-Methyl transferase	altered lignin levels
Glutamine synthetase	Glufosinate, Bialaphos
Adenylosuccinate Lyase (ADSL)	Inhibitors of IMP and AMP synthesis
Adenylosuccinate Synthase	Inhibitors of adenylosuccinate synthesis
Anthranilate Synthase	Inhibitors of tryptophan synthesis and catabolism
Nitrilase	3,5-dihalo-4-hydroxy-benzonitriles such as Bromoxynil and Ioxinyl
5-Enolpyruvyl-3phosphoshikimate Synthase (EPSPS)	Glyphosate or sulfosate

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
Glypnosate oxidoreductase	Glyphosate or sulfosate
Protoporphyrinogen oxidase (PROTOX)	Diphenylethers, cyclic imides, phenylpyrazoles, pyridin derivatives, phenopylate, oxadiazoles etc.
Cytochrome P450 eg. P450 SU1 or selection	Xenobiotics and herbicides such as Sulfonylureas
Polyphenol oxidase or Polyphenol oxidase antisense	blackspot bruise
Metallothionein	bacterial and fungal pathogens such as phytophtora
Ribonuclease	Phytophtora, Verticillium, Rhizoctonia
Antifungal polypeptide AlyAFP	bacterial and fungal pathogens such as phytophtora
oxalate oxidase	bacterial and fungal pathogens such as Phytophtora, Verticillium, Rhizoctonia
glucose oxidase	bacterial and fungal pathogens such as Phytophtora, Verticillium, Rhizoctonia
pyrrolnitrin synthesis genes	bacterial and fungal pathogens such as Phytophtora, Verticillium, Rhizoctonia
serine/threonine kinases	bacterial and fungal pathogens such as Phytophtora, Verticillium, Rhizoctonia
Cecropin B	bacteria such as corynebacterium sepedonicum, Erwinia carotovora
Phenylalanine ammonia lyase (PAL)	bacterial and fungal pathogens such as Phytophtora, Verticillium, Rhizoctonia
phytoalexins	bacterial and fungal pathogens such as Phytophtora, Verticillium, Rhizoctonia
B-1,3-glucanase antisense	bacterial and fungal pathogens such as Phytophtora, Verticillium, Rhizoctonia
receptor kinase	bacterial and fungal pathogens such as Phytophtora, Verticillium, Rhizoctonia

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
Hypersensitive response eliciting polypeptide	bacterial and fungal pathogens such as Phytophtora, Verticillium, Rhizoctonia
Systemic acquires resistance (SAR) genes	viral, bacterial, fungal, nematodal pathogens
Chitinases	bacterial and fungal pathogens such as Phytophtora, Verticillium, Rhizoctonia
Barnase	bacterial and fungal pathogens such as Phytophtora, Verticillium, Rhizoctonia
Disease resistance response gene 49	bacterial and fungal pathogens such as Phytophtora, Verticillium, Rhizoctonia
trans aldolase antisense	blackspots
Glucanases	bacterial and fungal pathogens such as Phytophtora, Verticillium, Rhizoctonia
double stranded ribonuclease	viruses such as PLRV, PVY and TRV
Coat proteins	viruses such as PLRV, PVY and TRV
17kDa or 60 kDa protein	viruses such as PLRV, PVY and TRV
Nuclear inclusion proteins eg. a or b	viruses such as PLRV, PVY and TRV
Pseudoubiquitin	viruses such as PLRV, PVY and TRV
Replicase	viruses such as PLRV, PVY and TRV
Bacillus thuringiensis toxins, VIP 3, Bacillus cereus toxins, Photorabdus and Xenorhabdus toxins	coleoptera eg colorado potato beetle, aphids
3- Hydroxysteroid oxidase	coleoptera eg colorado potato beetle, aphids
Peroxidase	coleoptera eg colorado potato beetle, aphids
Aminopeptidase inhibitors eg. Leucine aminopeptidase inhibitor	coleoptera eg colorado potato beetle, aphids
stilbene synthase	coleoptera eg colorado potato beetle, aphids

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
Lectines	coleoptera eg colorado potato beetle, aphids
Protease Inhibitors eg cystatin, patatin	coleoptera eg colorado potato beetle, aphids
ribosome inactivating protein	coleoptera eg colorado potato beetle, aphids
HMG-CoA reductase	coleoptera eg colorado potato beetle, aphids
Cyst nematode hatching stimulus	cyst nematodes
Barnase	nematodes eg root knot nematodes and cyst nematodes
Antifeeding principles	nematodes eg root knot nematodes and cyst nematodes

Table A7: Crop Tomatoes

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
Acetolactate synthase (ALS)	Sulfonylureas, Imidazolinones, Triazolopyrimidines, Pyrimidyloxybenzoates, Phtalides
AcetylCoA Carboxylase (ACCase)	Aryloxyphenoxyalkanecarboxylic acids, cyclohexanediones
Hydroxyphenylpyruvate dioxygenase (HPPD)	Isoxazoles such as Isoxaflutol or Isoxachlortol, Triones such as mesotrione or sulcotrione
Phosphinothricin acetyl transferase	Phosphinothricin
O-Methyl transferase	altered lignin levels
Glutamine synthetase	Glufosinate, Bialaphos
Adenylosuccinate Lyase (ADSL)	Inhibitors of IMP and AMP synthesis
Adenylosuccinate Synthase	Inhibitors of adenylosuccinate synthesis
Anthranilate Synthase	Inhibitors of tryptophan synthesis and

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
	catabolism
Nitrilase	3,5-dihalo-4-hydroxy-benzonitriles such as Bromoxynil and Ioxinyl
5-Enolpyruvyl-3phosphoshikimate Synthase (EPSPS)	Glyphosate or sulfosate
Glyphosate oxidoreductase	Glyphosate or sulfosate
Protoporphyrinogen oxidase (PROTOX)	Diphenylethers, cyclic imides, phenylpyrazoles, pyridin derivatives, phenopylate, oxadiazoles etc.
Cytochrome P450 eg. P450 SU1 or selection	Xenobiotics and herbicides such as Sulfonylureas
Polyphenol oxidase or Polyphenol oxidase antisense	blackspot bruise
Metallothionein	bacterial and fungal pathogens such as phytophtora
Ribonuclease	Phytophtora, Verticillium, Rhizoctonia
Antifungal polypeptide AlyAFP	bacterial and fungal pathogens such as bacterial speck, fusarium, soft rot, powdery mildew, crown rot, leaf mould etc.
oxalate oxidase	bacterial and fungal pathogens such as bacterial speck, fusarium, soft rot, powdery mildew, crown rot, leaf mould etc.
glucose oxidase	bacterial and fungal pathogens such as bacterial speck, fusarium, soft rot, powdery mildew, crown rot, leaf mould etc.
pyrrolnitrin synthesis genes	bacterial and fungal pathogens such as bacterial speck, fusarium, soft rot, powdery mildew, crown rot, leaf mould

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
	etc.
serine/threonine kinases	bacterial and fungal pathogens such as bacterial speck, fusarium, soft rot, powdery mildew, crown rot, leaf mould etc.
Cecropin B	bacterial and fungal pathogens such as bacterial speck, fusarium, soft rot, powdery mildew, crown rot, leaf mould etc.
Phenylalanine ammonia lyase (PAL)	bacterial and fungal pathogens such as bacterial speck, fusarium, soft rot, powdery mildew, crown rot, leaf mould etc.
Cf genes eg. Cf 9 Cf5 Cf4 Cf2	leaf mould
Osmotin	alternaria solani
Alpha Hordothionin	bacteria
Systemin	bacterial and fungal pathogens such as bacterial speck, fusarium, soft rot, powdery mildew, crown rot, leaf mould etc.
Polygalacturonase inhibitors	bacterial and fungal pathogens such as bacterial speck, fusarium, soft rot, powdery mildew, crown rot, leaf mould etc.
Prf regulatory gene	bacterial and fungal pathogens such as bacterial speck, fusarium, soft rot, powdery mildew, crown rot, leaf mould etc.
I2 Fusarium resistance locus	fusarium
phytoalexins	bacterial and fungal pathogens such as bacterial speck, fusarium, soft rot, powdery mildew, crown rot, leaf mould

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
	etc.
B-1,3-glucanase antisense	bacterial and fungal pathogens such as bacterial speck, fusarium, soft rot, powdery mildew, crown rot, leaf mould etc.
receptor kinase	bacterial and fungal pathogens such as bacterial speck, fusarium, soft rot, powdery mildew, crown rot, leaf mould etc.
Hypersensitive response eliciting polypeptide	bacterial and fungal pathogens such as bacterial speck, fusarium, soft rot, powdery mildew, crown rot, leaf mould etc.
Systemic acquires resistance (SAR) genes	viral, bacterial, fungal, nematodal pathogens
Chitinases	bacterial and fungal pathogens such as bacterial speck, fusarium, soft rot, powdery mildew, crown rot, leaf mould etc.
Barnase	bacterial and fungal pathogens such as bacterial speck, fusarium, soft rot, powdery mildew, crown rot, leaf mould etc.
Glucanases	bacterial and fungal pathogens such as bacterial speck, fusarium, soft rot, powdery mildew, crown rot, leaf mould etc.
double stranded ribonuclease	viruses such as PLRV, PVY and ToMoV
Coat proteins	viruses such as PLRV, PVY and ToMoV
17kDa or 60 kDa protein	viruses such as PLRV, PVY and ToMoV
Nuclear inclusion proteins eg. a or b or	viruses such as PLRV, PVY and ToMoV

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
Nucleoprotein	TRV
Pseudoubiquitin	viruses such as PLRV, PVY and ToMoV
Replicase	viruses such as PLRV, PVY and ToMoV
Bacillus thuringiensis toxins, VIP 3, Bacillus cereus toxins, Photorabdus and Xenorhabdus toxins	lepidoptera eg heliothis, whiteflies aphids
3- Hydroxysteroid oxidase	lepidoptera eg heliothis, whiteflies aphids
Peroxidase	lepidoptera eg heliothis, whiteflies aphids
Aminopeptidase inhibitors eg. Leucine aminopeptidase inhibitor	lepidoptera eg heliothis, whiteflies aphids
Lectines	lepidoptera eg heliothis, whiteflies aphids
Protease Inhibitors eg cystatin, patatin	lepidoptera eg heliothis, whiteflies aphids
ribosome inactivating protein	lepidoptera eg heliothis, whiteflies aphids
stilbene synthase	lepidoptera eg heliothis, whiteflies aphids
HMG-CoA reductase	lepidoptera eg heliothis, whiteflies aphids
Cyst nematode hatching stimulus	cyst nematodes
Barnase	nematodes eg root knot nematodes and cyst nematodes
Antifeeding principles	nematodes eg root knot nematodes and cyst nematodes

Table A8: Crop Peppers

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
Acetolactate synthase (ALS)	Sulfonylureas, Imidazolinones,

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
	Triazolopyrimidines, Pyrimidyloxybenzoates, Phtalides
AcetylCoA Carboxylase (ACCase)	Aryloxyphenoxyalkanecarboxylic acids, cyclohexanediones
Hydroxyphenylpyruvate dioxygenase (HPPD)	Isoxazoles such as Isoxaflutol or Isoxachlortol, Triones such as mesotrione or sulcotrione
Phosphinothricin acetyl transferase	Phosphinothricin
O-Methyl transferase	altered lignin levels
Glutamine synthetase	Glufosinate, Bialaphos
Adenylosuccinate Lyase (ADSL)	Inhibitors of IMP and AMP synthesis
Adenylosuccinate Synthase	Inhibitors of adenylosuccinate synthesis
Anthranilate Synthase	Inhibitors of tryptophan synthesis and catabolism
Nitrilase	3,5-dihalo-4-hydroxy-benzonitriles such as Bromoxynil and Ioxinyl
5-Enolpyruvyl-3phosphoshikimate Synthase (EPSPS)	Glyphosate or sulfosate
Glyphosate oxidoreductase	Glyphosate or sulfosate
Protoporphyrinogen oxidase (PROTOX)	Diphenylethers, cyclic imides, phenylpyrazoles, pyridin derivatives, phenopylate, oxadiazoles etc.
Cytochrome P450 eg. P450 SU1 or selection	Xenobiotics and herbicides such as Sulfonylureas
Polyphenol oxidase or Polyphenol oxidase antisense	bacterial and fungal pathogens
Metallothionein	bacterial and fungal pathogens
Ribonuclease	bacterial and fungal pathogens
Antifungal polypeptide AlyAFP	bacterial and fungal pathogens
oxalate oxidase	bacterial and fungal pathogens
glucose oxidase	bacterial and fungal pathogens
pyrrolnitrin synthesis genes	bacterial and fungal pathogens

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
serine/threonine kinases	bacterial and fungal pathogens
Cecropin B	bacterial and fungal pathogens rot, leaf mould etc.
Phenylalanine ammonia lyase (PAL)	bacterial and fungal pathogens
Cf genes eg. Cf 9 Cf5 Cf4 Cf2	bacterial and fungai pathogens
Osmotin	bacterial and fungal pathogens
Alpha Hordothionin	bacterial and fungal pathogens
Systemin	bacterial and fungal pathogens
Polygalacturonase inhibitors	bacterial and fungal pathogens
Prf regulatory gene	bacterial and fungal pathogens
I2 Fusarium resistance locus	fusarium
phytoalexins	bacterial and fungal pathogens
B-1,3-glucanase antisense	bacterial and fungal pathogens
receptor kinase	bacterial and fungal pathogens
Hypersensitive response eliciting polypeptide	bacterial and fungal pathogens
Systemic acquires resistance (SAR) genes	viral, bacterial, fungal, nematodal pathogens
Chitinases	bacterial and fungal pathogens
Barnase	bacterial and fungal pathogens
Glucanases	bacterial and fungal pathogens
double stranded ribonuclease	viruses such as CMV, TEV
Coat proteins	viruses such as CMV, TEV
17kDa or 60 kDa protein	viruses such as CMV, TEV
Nuclear inclusion proteins eg. a or b or Nucleoprotein	viruses such as CMV, TEV
Pseudoubiquitin	viruses such as CMV, TEV
Replicase	viruses such as CMV, TEV
Bacillus thuringiensis toxins, VIP 3, Bacillus cereus toxins, Photorabdus and Xenorhabdus toxins	lepidoptera, whiteflies aphids

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
3- Hydroxysteroid oxidase	lepidoptera, whiteflies aphids
Peroxidase	lepidoptera, whiteflies aphids
Aminopeptidase inhibitors eg. Leucine aminopeptidase inhibitor	lepidoptera, whiteflies aphids
Lectines	lepidoptera, whiteflies aphids
Protease Inhibitors eg cystatin, patatin	lepidoptera, whiteflies aphids
ribosome inactivating protein	lepidoptera, whiteflies aphids
stilbene synthase	lepidoptera, whiteflies aphids
HMG-CoA reductase	lepidoptera, whiteflies aphids
Cyst nematode hatching stimulus	cyst nematodes
Barnase	nematodes eg root knot nematodes and cyst nematodes
Antifeeding principles	nematodes eg root knot nematodes and cyst nematodes

Table A9: Crop Grapes

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
Acetolactate synthase (ALS)	Sulfonylureas, Imidazolinones, Triazolopyrimidines, Pyrimidyloxybenzoates, Phtalides
AcetylCoA Carboxylase (ACCase)	Aryloxyphenoxyalkanecarboxylic acids, cyclohexanediones
Hydroxyphenylpyruvate dioxygenase (HPPD)	Isoxazoles such as Isoxaflutol or Isoxachlortol, Triones such as mesotrione or sulcotrione
Phosphinothricin acetyl transferase	Phosphinothricin
O-Methyl transferase	altered lignin levels
Glutamine synthetase	Glufosinate, Bialaphos
Adenylosuccinate Lyase (ADSL)	Inhibitors of IMP and AMP synthesis
Adenylosuccinate Synthase	Inhibitors of adenylosuccinate synthesis
Anthranilate Synthase	Inhibitors of tryptophan synthesis and

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
	catabolism
Nitrilase	3,5-dihalo-4-hydroxy-benzonitriles such as Bromoxynil and Ioxinyl
5-Enolpyruvyl-3phosphoshikimate Synthase (EPSPS)	Glyphosate or sulfosate
Glyphosate oxidoreductase	Glyphosate or sulfosate
Protoporphyrinogen oxidase (PROTOX)	Diphenylethers, cyclic imides, phenylpyrazoles, pyridin derivatives, phenopylate, oxadiazoles etc.
Cytochrome P450 eg. P450 SU1 or selection	Xenobiotics and herbicides such as Sulfonylureas
Polyphenol oxidase or Polyphenol oxidase antisense	bacterial and fungal pathogens like Botrytis and powdery mildew
Metallothionein	bacterial and fungal pathogens like Botrytis and powdery mildew
Ribonuclease	bacterial and fungal pathogens like Botrytis and powdery mildew
Antifungal polypeptide AlyAFP	bacterial and fungal pathogens like Botrytis and powdery mildew
oxalate oxidase	bacterial and fungal pathogens like Botrytis and powdery mildew
glucose oxidase	bacterial and fungal pathogens like Botrytis and powdery mildew
pyrrolnitrin synthesis genes	bacterial and fungal pathogens like Botrytis and powdery mildew
serine/threonine kinases	bacterial and fungal pathogens like Botrytis and powdery mildew
Cecropin B	bacterial and fungal pathogens like Botrytis and powdery mildew
Phenylalanine ammonia lyase (PAL)	bacterial and fungal pathogens like Botrytis and powdery mildew
Cf genes eg. Cf 9 Cf5 Cf4 Cf2	bacterial and fungal pathogens like

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
	Botrytis and powdery mildew
Osmotin	bacterial and fungal pathogens like
	Botrytis and powdery mildew
Alpha Hordothionin	bacterial and fungal pathogens like
	Botrytis and powdery mildew
Systemin	bacterial and fungal pathogens like
	Botrytis and powdery mildew
Polygalacturonase inhibitors	bacterial and fungal pathogens like
	Botrytis and powdery mildew
Prf regulatory gene	bacterial and fungal pathogens like
	Botrytis and powdery mildew
phytoalexins	bacterial and fungal pathogens like
	Botrytis and powdery mildew
B-1,3-glucanase antisense	bacterial and fungal pathogens like
	Botrytis and powdery mildew
receptor kinase	bacterial and fungal pathogens like
	Botrytis and powdery mildew
Hypersensitive response eliciting	bacterial and fungal pathogens like
polypeptide	Botrytis and powdery mildew
Systemic acquires resistance (SAR)	viral, bacterial, fungal, nematodal
genes	pathogens
Chitinases	bacterial and fungal pathogens like
	Botrytis and powdery mildew
Barnase	bacterial and fungal pathogens like
	Botrytis and powdery mildew
Glucanases	bacterial and fungal pathogens like
	Botrytis and powdery mildew
double stranded ribonuclease	viruses
Coat proteins	viruses
17kDa or 60 kDa protein	viruses
Nuclear inclusion proteins eg. a or b or	viruses

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
Nucleoprotein	
Pseudoubiquitin	viruses
Replicase	viruses
Bacillus thuringiensis toxins, VIP 3, Bacillus cereus toxins, Photorabdus and Xenorhabdus toxins	lepidoptera, aphids
3- Hydroxysteroid oxidase	lepidoptera, aphids
Peroxidase	lepidoptera, aphids
Aminopeptidase inhibitors eg. Leucine aminopeptidase inhibitor	lepidoptera, aphids
Lectines	lepidoptera, aphids
Protease Inhibitors eg cystatin, patatin	lepidoptera, aphids
ribosome inactivating protein	lepidoptera, aphids
stilbene synthase	lepidoptera, aphids, diseases
HMG-CoA reductase	lepidoptera, aphids
Cyst nematode hatching stimulus	cyst nematodes
Barnase	nematodes eg root knot nematodes and cyst nematodes or general diseases
CBI	root knot nematodes
Antifeeding principles	nematodes eg root knot nematodes or root cyst nematodes

Table A10: crop Oil Seed rape

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
Acetolactate synthase (ALS)	Sulfonylureas, Imidazolinones, Triazolopyrimidines, Pyrimidyloxybenzoates, Phtalides
AcetylCoA Carboxylase (ACCase)	Aryloxyphenoxyalkanecarboxylic acids, cyclohexanediones
Hydroxyphenylpyruvate dioxygenase (HPPD)	Isoxazoles such as Isoxaflutol or Isoxachlortol, Triones such as

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
	mesotrione or sulcotrione
Phosphinothricin acetyl transferase	Phosphinothricin
O-Methyl transferase	altered lignin levels
Glutamine synthetase	Glufosinate, Bialaphos
Adenylosuccinate Lyase (ADSL)	Inhibitors of IMP and AMP synthesis
Adenylosuccinate Synthase	Inhibitors of adenylosuccinate synthesis
Anthranilate Synthase	Inhibitors of tryptophan synthesis and catabolism
Nitrilase	3,5-dihalo-4-hydroxy-benzonitriles such as Bromoxynil and Ioxinyl
5-Enolpyruvyl-3phosphoshikimate Synthase (EPSPS)	Glyphosate or sulfosate
Glyphosate oxidoreductase	Glyphosate or sulfosate
Protoporphyrinogen oxidase (PROTOX)	Diphenylethers, cyclic imides, phenylpyrazoles, pyridin derivatives, phenopylate, oxadiazoles etc.
Cytochrome P450 eg. P450 SU1 or selection	Xenobiotics and herbicides such as Sulfonylureas
Polyphenol oxidase or Polyphenol oxidase antisense	bacterial and fungal pathogens like Cylindrosporium, Phoma, Sclerotinia
Metallothionein	bacterial and fungal pathogens like Cylindrosporium, Phoma, Sclerotinia
Ribonuclease	bacterial and fungal pathogens like Cylindrosporium, Phoma, Sclerotinia
Antifungal polypeptide AlyAFP	bacterial and fungal pathogens like Cylindrosporium, Phoma, Sclerotinia
oxalate oxidase	bacterial and fungal pathogens like Cylindrosporium, Phoma, Sclerotinia
glucose oxidase	bacterial and fungal pathogens like Cylindrosporium, Phoma, Sclerotinia
pyrrolnitrin synthesis genes	bacterial and fungal pathogens like Cylindrosporium, Phoma, Sclerotinia

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
serine/threonine kinases	bacterial and fungal pathogens like Cylindrosporium, Phoma, Sclerotinia
Cecropin B	bacterial and fungal pathogens like Cylindrosporium, Phoma, Sclerotinia
Phenylalanine ammonia lyase (PAL)	bacterial and fungal pathogens like Cylindrosporium, Phoma, Sclerotinia
Cf genes eg. Cf 9 Cf5 Cf4 Cf2	bacterial and fungal pathogens like Cylindrosporium, Phoma, Sclerotinia
Osmotin	bacterial and fungal pathogens like Cylindrosporium, Phoma, Sclerotinia
Alpha Hordothionin	bacterial and fungal pathogens like Cylindrosporium, Phoma, Sclerotinia
Systemin	bacterial and fungal pathogens like Cylindrosporium, Phoma, Sclerotinia
Polygalacturonase inhibitors	bacterial and fungal pathogens like Cylindrosporium, Phoma, Sclerotinia
Prf regulatory gene	bacterial and fungal pathogens like Cylindrosporium, Phoma, Sclerotinia
phytoalexins	bacterial and fungal pathogens like Cylindrosporium, Phoma, Sclerotinia
B-1,3-glucanase antisense	bacterial and fungal pathogens like Cylindrosporium, Phoma, Sclerotinia
receptor kinase	bacterial and fungal pathogens like Cylindrosporium, Phoma, Sclerotinia
Hypersensitive response eliciting polypeptide	bacterial and fungal pathogens like Cylindrosporium, Phoma, Sclerotinia
Systemic acquires resistance (SAR) genes	viral, bacterial, fungal, nematodal pathogens
Chitinases	bacterial and fungal pathogens like Cylindrosporium, Phoma, Sclerotinia
Barnase	bacterial and fungal pathogens like Cylindrosporium, Phoma,

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
	Sclerotinia, nematodes
Glucanases	bacterial and fungal pathogens like Cylindrosporium, Phoma, Sclerotinia
double stranded ribonuclease	viruses
Coat proteins	viruses
17kDa or 60 kDa protein	viruses
Nuclear inclusion proteins eg. a or b or Nucleoprotein	viruses
Pseudoubiquitin	viruses
Replicase	viruses
Bacillus thuringiensis toxins, VIP 3, Bacillus cereus toxins, Photorabdus and Xenorhabdus toxins	lepidoptera, aphids
3- Hydroxysteroid oxidase	lepidoptera, aphids
Peroxidase	lepidoptera, aphids
Aminopeptidase inhibitors eg. Leucine aminopeptidase inhibitor	lepidoptera, aphids
Lectines	lepidoptera, aphids
Protease Inhibitors eg cystatin, patatin, CPTI	lepidoptera, aphids
ribosome inactivating protein	lepidoptera, aphids
stilbene synthase	lepidoptera, aphids, diseases
HMG-CoA reductase	lepidoptera, aphids
Cyst nematode hatching stimulus	cyst nematodes
Barnase	nematodes eg root knot nematodes and cyst nematodes
CBI	root knot nematodes
Antifeeding principles induced at a nematode feeding site	nematodes eg root knot nematodes, root cyst nematodes

Table A11: Crop Brassica vegetable (cabbage, brussel sprouts, broccoli etc.)

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
Acetolactate synthase (ALS)	Sulfonylureas, Imidazolinones, Triazolopyrimidines, Pyrimidyloxybenzoates, Phtalides
AcetylCoA Carboxylase (ACCase)	Aryloxyphenoxyalkanecarboxylic acids, cyclohexanediones
Hydroxyphenylpyruvate dioxygenase (HPPD)	Isoxazoles such as Isoxaflutol or Isoxachlortol, Triones such as mesotrione or sulcotrione
Phosphinothricin acetyl transferase	Phosphinothricin
O-Methyl transferase	altered lignin levels
Glutamine synthetase	Glufosinate, Bialaphos
Adenylosuccinate Lyase (ADSL)	Inhibitors of IMP and AMP synthesis
Adenylosuccinate Synthase	Inhibitors of adenylosuccinate synthesis
Anthranilate Synthase	Inhibitors of tryptophan synthesis and catabolism
Nitrilase	3,5-dihalo-4-hydroxy-benzonitriles such as Bromoxynil and Ioxinyl
5-Enolpyruvyl-3phosphoshikimate Synthase (EPSPS)	Glyphosate or sulfosate
Glyphosate oxidoreductase	Glyphosate or sulfosate
Protoporphyrinogen oxidase (PROTOX)	Diphenylethers, cyclic imides, phenylpyrazoles, pyridin derivatives, phenopylate, oxadiazoles etc.
Cytochrome P450 eg. P450 SU1 or selection	Xenobiotics and herbicides such as Sulfonylureas
Polyphenol oxidase or Polyphenol oxidase antisense	bacterial and fungal pathogens
Metallothionein	bacterial and fungal pathogens
Ribonuclease	bacterial and fungal pathogens
Antifungal polypeptide AlyAFP	bacterial and fungal pathogens
oxalate oxidase	bacterial and fungal pathogens
glucose oxidase	bacterial and fungal pathogens

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
pyrrolnitrin synthesis genes	bacterial and fungal pathogens
serine/threonine kinases	bacterial and fungal pathogens
Cecropin B	bacterial and fungal pathogens
Phenylalanine ammonia lyase (PAL)	bacterial and fungal pathogens
Cf genes eg. Cf 9 Cf5 Cf4 Cf2	bacterial and fungal pathogens
Osmotin	bacterial and fungal pathogens
Alpha Hordothionin	bacterial and fungal pathogens
Systemin	bacterial and fungal pathogens
Polygalacturonase inhibitors	bacterial and fungal pathogens
Prf regulatory gene	bacterial and fungal pathogens
phytoalexins	bacterial and fungal pathogens
B-1,3-glucanase antisense	bacterial and fungal pathogens
receptor kinase	bacterial and fungal pathogens
Hypersensitive response eliciting polypeptide	bacterial and fungal pathogens
Systemic acquires resistance (SAR) genes	viral, bacterial, fungal, nematodal pathogens
Chitinases	bacterial and fungal pathogens
Barnase	bacterial and fungal pathogens
Glucanases	bacterial and fungal pathogens
double stranded ribonuclease	viruses
Coat proteins	viruses
17kDa or 60 kDa protein	viruses
Nuclear inclusion proteins eg. a or b or Nucleoprotein	viruses
Pseudoubiquitin	viruses
Replicase	viruses
Bacillus thuringiensis toxins, VIP 3, Bacillus cereus toxins, Photorabdus and Xenorhabdus toxins	lepidoptera, aphids
3- Hydroxysteroid oxidase	lepidoptera, aphids
Peroxidase	lepidoptera, aphids

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
Aminopeptidase inhibitors eg. Leucine aminopeptidase inhibitor	lepidoptera, aphids
Lectines	lepidoptera, aphids
Protease Inhibitors eg cystatin, patatin, CPTI	lepidoptera, aphids
ribosome inactivating protein	lepidoptera, aphids
stilbene synthase	lepidoptera, aphids, diseases
HMG-CoA reductase	lepidoptera, aphids
Cyst nematode hatching stimulus	cyst nematodes
Barnase	nematodes eg root knot nematodes and cyst nematodes
CBI	root knot nematodes
Antifeeding principles induced at a nematode feeding site	nematodes eg root knot nematodes, root cyst nematodes

Table A12 : Crop Pome fruits eg apples, pears

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
Acetolactate synthase (ALS)	Sulfonylureas, Imidazolinones, Triazolopyrimidines, Pyrimidyloxybenzoates, Phtalides
AcetylCoA Carboxylase (ACCase)	Aryloxyphenoxyalkanecarboxylic acids, cyclohexanediones
Hydroxyphenylpyruvate dioxygenase (HPPD)	Isoxazoles such as Isoxaflutol or Isoxachlortol, Triones such as mesotrione or sulcotrione
Phosphinothricin acetyl transferase	Phosphinothricin
O-Methyl transferase	altered lignin levels
Glutamine synthetase	Glufosinate, Bialaphos
Adenylosuccinate Lyase (ADSL)	Inhibitors of IMP and AMP synthesis
Adenylosuccinate Synthase	Inhibitors of adenylosuccinate synthesis
Anthranilate Synthase	Inhibitors of tryptophan synthesis and

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
	catabolism
Nitrilase	3,5-dihalo-4-hydroxy-benzonitriles such as Bromoxynil and Ioxinyl
5-Enolpyruvyl-3phosphoshikimate Synthase (EPSPS)	Glyphosate or sulfosate
Glyphosate oxidoreductase	Glyphosate or sulfosate
Protoporphyrinogen oxidase (PROTOX)	Diphenylethers, cyclic imides, phenylpyrazoles, pyridin derivatives, phenopylate, oxadiazoles etc.
Cytochrome P450 eg. P450 SU1 or selection	Xenobiotics and herbicides such as Sulfonylureas
Polyphenol oxidase or Polyphenol oxidase antisense	bacterial and fungal pathogens like apple scab or fireblight
Metallothionein	bacterial and fungal pathogens like apple scab or fireblight
Ribonuclease	bacterial and fungal pathogens like apple scab or fireblight
Antifungal polypeptide AlyAFP	bacterial and fungal pathogens like apple scab or fireblight
oxalate oxidase	bacterial and fungal pathogens like apple scab or fireblight
glucose oxidase	bacterial and fungal pathogens like apple scab or fireblight
pyrrolnitrin synthesis genes	bacterial and fungal pathogens like apple scab or fireblight
serine/threonine kinases	bacterial and fungal pathogens like apple scab or fireblight
Cecropin B	bacterial and fungal pathogens like apple scab or fireblight
Phenylalanine ammonia lyase (PAL)	bacterial and fungal pathogens like apple scab or fireblight
Cf genes eg. Cf 9 Cf5 Cf4 Cf2	bacterial and fungal pathogens like

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
	apple scab or fireblight
Osmotin	bacterial and fungal pathogens like apple scab or fireblight
Alpha Hordothionin	bacterial and fungal pathogens like apple scab or fireblight
Systemin	bacterial and fungal pathogens like apple scab or fireblight
Polygalacturonase inhibitors	bacterial and fungal pathogens like apple scab or fireblight
Prf regulatory gene	bacterial and fungal pathogens like apple scab or fireblight
phytoalexins	bacterial and fungal pathogens like apple scab or fireblight
B-1,3-glucanase antisense	bacterial and fungal pathogens like apple scab or fireblight
receptor kinase	bacterial and fungal pathogens like apple scab or fireblight
Hypersensitive response eliciting polypeptide	bacterial and fungal pathogens like apple scab or fireblight
Systemic acquires resistance (SAR) genes	viral, bacterial, fungal, nematodal pathogens
Lytic protein	bacterial and fungal pathogens like apple scab or fireblight
Lysozym	bacterial and fungal pathogens like apple scab or fireblight
Chitinases	bacterial and fungal pathogens like apple scab or fireblight
Barnase	bacterial and fungal pathogens like apple scab or fireblight
Glucanases	bacterial and fungal pathogens like apple scab or fireblight

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
double stranded ribonuclease	viruses
Coat proteins	viruses
17kDa or 60 kDa protein	viruses
Nuclear inclusion proteins eg. a or b or Nucleoprotein	viruses
Pseudoubiquitin	viruses
Replicase	viruses
Bacillus thuringiensis toxins, VIP 3, Bacillus cereus toxins, Photorabdus and Xenorhabdus toxins	lepidoptera, aphids, mites
3- Hydroxysteroid oxidase	lepidoptera, aphids, mites
Peroxidase	lepidoptera, aphids, mites
Aminopeptidase inhibitors eg. Leucine aminopeptidase inhibitor	lepidoptera, aphids, mites
Lectines	lepidoptera, aphids, mites
Protease Inhibitors eg cystatin, patatin, CPTI	lepidoptera, aphids , mites
ribosome inactivating protein	lepidoptera, aphids, mites
stilbene synthase	lepidoptera, aphids, diseases, mites
HMG-CoA reductase	lepidoptera, aphids, mites
Cyst nematode hatching stimulus	cyst nematodes
Barnase	nematodes eg root knot nematodes and cyst nematodes
CBI	root knot nematodes
Antifeeding principles induced at a nematode feeding site	nematodes eg root knot nematodes, root cyst nematodes

Table A13: Crop Melons

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
Acetolactate synthase (ALS)	Sulfonylureas, Imidazolinones, Triazolopyrimidines,

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
	Pyrimidyloxybenzoates, Phtalides
AcetylCoA Carboxylase (ACCase)	Aryloxyphenoxyalkanecarboxylic acids, cyclohexanediones
Hydroxyphenylpyruvate dioxygenase (HPPD)	Isoxazoles such as Isoxaflutol or Isoxachlortol, Triones such as mesotrione or sulcotrione
Phosphinothricin acetyl transferase	Phosphinothricin
O-Methyl transferase	altered lignin levels
Glutamine synthetase	Glufosinate, Bialaphos
Adenylosuccinate Lyase (ADSL)	Inhibitors of IMP and AMP synthesis
Adenylosuccinate Synthase	Inhibitors of adenylosuccinate synthesis
Anthranilate Synthase	Inhibitors of tryptophan synthesis and catabolism
Nitrilase	3,5-dihalo-4-hydroxy-benzonitriles such as Bromoxynil and Ioxinyl
5-Enolpyruvyl-3phosphoshikimate Synthase (EPSPS)	Glyphosate or sulfosate
Glyphosate oxidoreductase	Glyphosate or sulfosate
Protoporphyrinogen oxidase (PROTOX)	Diphenylethers, cyclic imides, phenylpyrazoles, pyridin derivatives, phenopylate, oxadiazoles etc.
Cytochrome P450 eg. P450 SU1 or selection	Xenobiotics and herbicides such as Sulfonylureas
Polyphenol oxidase or Polyphenol oxidase antisense	bacterial or fungal pathogens like phytophtora
Metallothionein	bacterial or fungal pathogens like phytophtora
Ribonuclease	bacterial or fungal pathogens like phytophtora
Antifungal polypeptide AlyAFP	bacterial or fungal pathogens like phytophtora

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
oxalate oxidase	bacterial or fungal pathogens like phytophtora
glucose oxidase	bacterial or fungal pathogens like phytophtora
pyrrolnitrin synthesis genes	bacterial or fungal pathogens like phytophtora
serine/threonine kinases	bacterial or fungal pathogens like phytophtora
Cecropin B	bacterial or fungal pathogens like phytophtora
Phenylalanine ammonia lyase (PAL)	bacterial or fungal pathogens like phytophtora
Cf genes eg. Cf 9 Cf5 Cf4 Cf2	bacterial or fungal pathogens like phytophtora
Osmotin	bacterial or fungal pathogens like phytophtora
Alpha Hordothionin	bacterial or fungal pathogens like phytophtora
Systemin	bacterial or fungal pathogens like phytophtora
Polygalacturonase inhibitors	bacterial or fungal pathogens like phytophtora
Prf regulatory gene	bacterial or fungal pathogens like phytophtora
phytoalexins	bacterial or fungal pathogens like phytophtora
B-1,3-glucanase antisense	bacterial or fungal pathogens like phytophtora
receptor kinase	bacterial or fungal pathogens like phytophtora
Hypersensitive response eliciting polypeptide	bacterial or fungal pathogens like phytophtora

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
Systemic acquires resistance (SAR) genes	viral, bacterial, fungal, nematodal pathogens
Lytic protein	bacterial or fungal pathogens like phytophtora
Lysozym	bacterial or fungal pathogens like phytophtora
Chitinases	bacterial or fungal pathogens like phytophtora
Barnase	bacterial or fungal pathogens like phytophtora
Glucanases	bacterial or fungal pathogens like phytophtora
double stranded ribonuclease	viruses as CMV,, PRSV, WMV2, SMV, ZYMV
Coat proteins	viruses as CMV,, PRSV, WMV2, SMV, ZYMV
17kDa or 60 kDa protein	viruses as CMV,, PRSV, WMV2, SMV, ZYMV
Nuclear inclusion proteins eg. a or b or Nucleoprotein	viruses as CMV,, PRSV, WMV2, SMV, ZYMV
Pseudoubiquitin	viruses as CMV,, PRSV, WMV2, SMV, ZYMV
Replicase	viruses as CMV,, PRSV, WMV2, SMV, ZYMV
Bacillus thuringiensis toxins, VIP 3, Bacillus cereus toxins, Photorabdus and Xenorhabdus toxins	lepidoptera, aphids, mites
3- Hydroxysteroid oxidase	lepidoptera, aphids, mites, whitefly
Peroxidase	lepidoptera, aphids, mites, whitefly
Aminopeptidase inhibitors eg. Leucine aminopeptidase inhibitor	lepidoptera, aphids, mites, whitefly

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
Lectines	lepidoptera, aphids, mites, whitefly
Protease Inhibitors eg cystatin, patatin, CPTI, virgiferin	lepidoptera, aphids, mites, whitefly
ribosome inactivating protein	lepidoptera, aphids, mites, whitefly
stilbene synthase	lepidoptera, aphids, mites, whitefly
HMG-CoA reductase	lepidoptera, aphids, mites, whitefly
Cyst nematode hatching stimulus	cyst nematodes
Barnase	nematodes eg root knot nematodes and cyst nematodes
CBI	root knot nematodes
Antifeeding principles induced at a nematode feeding site	nematodes eg root knot nematodes, root cyst nematodes

Table A14: Crop Banana

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
Acetolactate synthase (ALS)	Sulfonylureas, Imidazolinones, Triazolopyrimidines, Pyrimidyloxybenzoates, Phtalides
AcetylCoA Carboxylase (ACCase)	Aryloxyphenoxyalkanecarboxylic acids, cyclohexanediones
Hydroxyphenylpyruvate dioxygenase (HPPD)	Isoxazoles such as Isoxaflutol or Isoxachlortol, Triones such as mesotrione or sulcotrione
Phosphinothricin acetyl transferase	Phosphinothricin
O-Methyl transferase	altered lignin levels
Glutamine synthetase	Glufosinate, Bialaphos
Adenylosuccinate Lyase (ADSL)	Inhibitors of IMP and AMP synthesis
Adenylosuccinate Synthase	Inhibitors of adenylosuccinate synthesis
Anthranilate Synthase	Inhibitors of tryptophan synthesis and catabolism
Nitrilase	3,5-dihalo-4-hydroxy-benzonitriles such

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
	as Bromoxynil and Ioxinyl
5-Enolpyruvyl-3phosphoshikimate Synthase (EPSPS)	Glyphosate or sulfosate
Glyphosate oxidoreductase	Glyphosate or sulfosate
Protoporphyrinogen oxidase (PROTOX)	Diphenylethers, cyclic imides, phenylpyrazoles, pyridin derivatives, phenopylate, oxadiazoles etc.
Cytochrome P450 eg. P450 SU1 or selection	Xenobiotics and herbicides such as Sulfonylureas
Polyphenol oxidase or Polyphenol oxidase antisense	bacterial or fungal pathogens
Metallothionein	bacterial or fungal pathogens
Ribonuclease	bacterial or fungal pathogens
Antifungal polypeptide AlyAFP	bacterial or fungal pathogens
oxalate oxidase	bacterial or fungal pathogens
glucose oxidase	bacterial or fungal pathogens
pyrrolnitrin synthesis genes	bacterial or fungal pathogens
serine/threonine kinases	bacterial or fungal pathogens
Cecropin B	bacterial or fungal pathogens
Phenylalanine ammonia lyase (PAL)	bacterial or fungal pathogens
Cf genes eg. Cf 9 Cf5 Cf4 Cf2	bacterial or fungal pathogens
Osmotin	bacterial or fungal pathogens
Alpha Hordothionin	bacterial or fungal pathogens
Systemin	bacterial or fungal pathogens
Polygalacturonase inhibitors	bacterial or fungal pathogens
Prf regulatory gene	bacterial or fungal pathogens
phytoalexins	bacterial or fungal pathogens
B-1,3-glucanase antisense	bacterial or fungal pathogens
receptor kinase	bacterial or fungal pathogens
Hypersensitive response eliciting polypeptide	bacterial or fungal pathogens
Systemic acquires resistance (SAR)	viral, bacterial, fungal, nematodal

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
genes	pathogens
Lytic protein	bacterial or fungal pathogens
Lysozym	bacterial or fungal pathogens
Chitinases	bacterial or fungal pathogens
Barnase	bacterial or fungal pathogens
Glucanases	bacterial or fungal pathogens
double stranded ribonuclease	viruses as Banana bunchy top virus (BBTV)
Coat proteins	viruses as Banana bunchy top virus (BBTV)
17kDa or 60 kDa protein	viruses as Banana bunchy top virus (BBTV)
Nuclear inclusion proteins eg. a or b or Nucleoprotein	viruses as Banana bunchy top virus (BBTV)
Pseudoubiquitin	viruses as Banana bunchy top virus (BBTV)
Replicase	viruses as Banana bunchy top virus (BBTV)
Bacillus thuringiensis toxins, VIP 3, Bacillus cereus toxins, Photorabdus and Xenorhabdus toxins	lepidoptera, aphids, mites, nematodes
3- Hydroxysteroid oxidase	lepidoptera, aphids, mites, nematodes
Peroxidase	lepidoptera, aphids, mites, nematodes
Aminopeptidase inhibitors eg. Leucine aminopeptidase inhibitor	lepidoptera, aphids, mites, nematodes
Lectines	lepidoptera, aphids, mites, nematodes
Protease Inhibitors eg cystatin, patatin, CPTI, virgiferin	lepidoptera, aphids, mites, nematodes
ribosome inactivating protein	lepidoptera, aphids, mites, nematodes
stilbene synthase	lepidoptera, aphids, mites, nematodes
HMG-CoA reductase	lepidoptera, aphids, mites, nematodes
Cyst nematode hatching stimulus	cyst nematodes

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
Barnase	nematodes eg root knot nematodes and cyst nematodes
CBI	root knot nematodes
Antifeeding principles induced at a nematode feeding site	nematodes eg root knot nematodes, root cyst nematodes

Table A 15: Crop Cotton

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
Acetolactate synthase (ALS)	Sulfonylureas, Imidazolinones, Triazolopyrimidines, Pyrimidyloxybenzoates, Phtalides
AcetylCoA Carboxylase (ACCase)	Aryloxyphenoxyalkanecarboxylic acids, cyclohexanediones
Hydroxyphenylpyruvate dioxygenase (HPPD)	Isoxazoles such as Isoxaflutol or Isoxachlortol, Triones such as mesotrione or sulcotrione
Phosphinothricin acetyl transferase	Phosphinothricin
O-Methyl transferase	altered lignin levels
Glutamine synthetase	Glufosinate, Bialaphos
Adenylosuccinate Lyase (ADSL)	Inhibitors of IMP and AMP synthesis
Adenylosuccinate Synthase	Inhibitors of adenylosuccinate synthesis
Anthranilate Synthase	Inhibitors of tryptophan synthesis and catabolism
Nitrilase	3,5-dihalo-4-hydroxy-benzonitriles such as Bromoxynil and Ioxinyl
5-Enolpyruvyl-3phosphoshikimate Synthase (EPSPS)	Glyphosate or sulfosate
Glyphosate oxidoreductase	Glyphosate or sulfosate
Protoporphyrinogen oxidase (PROTOX)	Diphenylethers, cyclic imides, phenylpyrazoles, pyridin derivatives, phenopylate, oxadiazoles etc.

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
Cytochrome P450 eg. P450 SU1 or selection	Xenobiotics and herbicides such as Sulfonylureas
Polyphenol oxidase or Polyphenol oxidase antisense	bacterial or fungal pathogens
Metallothionein	bacterial or fungal pathogens
Ribonuclease	bacterial or fungal pathogens
Antifungal polypeptide AlyAFP	bacterial or fungal pathogens
oxalate oxidase	bacterial or fungal pathogens
glucose oxidase	bacterial or fungal pathogens
pyrrolnitrin synthesis genes	bacterial or fungal pathogens
serine/threonine kinases	bacterial or fungal pathogens
Cecropin B	bacterial or fungal pathogens
Phenylalanine ammonia lyase (PAL)	bacterial or fungal pathogens
Cf genes eg. Cf 9 Cf5 Cf4 Cf2	bacterial or fungal pathogens
Osmotin	bacterial or fungal pathogens
Alpha Hordothionin	bacterial or fungal pathogens
Systemin	bacterial or fungal pathogens
Polygalacturonase inhibitors	bacterial or fungal pathogens
Prf regulatory gene	bacterial or fungal pathogens
phytoalexins	bacterial or fungal pathogens
B-1,3-glucanase antisense	bacterial or fungal pathogens
receptor kinase	bacterial or fungal pathogens
Hypersensitive response eliciting polypeptide	bacterial or fungal pathogens
Systemic acquires resistance (SAR) genes	viral, bacterial, fungal, nematodal pathogens
Lytic protein	bacterial or fungal pathogens
Lysozym	bacterial or fungal pathogens
Chitinases	bacterial or fungal pathogens
Barnase	bacterial or fungal pathogens
Glucanases	bacterial or fungal pathogens
double stranded ribonuclease	viruses as wound tumor virus (WTV)

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
Coat proteins	viruses as wound tumor virus (WTV)
17kDa or 60 kDa protein	viruses as wound tumor virus (WTV)
Nuclear inclusion proteins eg. a or b or Nucleoprotein	viruses as wound tumor virus (WTV)
Pseudoubiquitin	viruses as wound tumor virus (WTV)
Replicase	viruses as wound tumor virus (WTV)
Bacillus thuringiensis toxins, VIP 3, Bacillus cereus toxins, Photorabdus and Xenorhabdus toxins	lepidoptera, aphids, mites, nematodes, whitefly
3- Hydroxysteroid oxidase	lepidoptera, aphids, mites, nematodes, whitefly
Peroxidase	lepidoptera, aphids, mites, nematodes, whitefly
Aminopeptidase inhibitors eg. Leucine aminopeptidase inhibitor	lepidoptera, aphids, mites, nematodes, whitefly
Lectines	lepidoptera, aphids, mites, nematodes, whitefly
Protease Inhibitors eg cystatin, patatin, CPTI, virgiferin	lepidoptera, aphids, mites, nematodes, whitefly
ribosome inactivating protein	lepidoptera, aphids, mites, nematodes, whitefly
stilbene synthase	lepidoptera, aphids, mites, nematodes, whitefly
HMG-CoA reductase	lepidoptera, aphids, mites, nematodes, whitefly
Cyst nematode hatching stimulus	cyst nematodes
Barnase	nematodes eg root knot nematodes and cyst nematodes
CBI	root knot nematodes
Antifeeding principles induced at a nematode feeding site	nematodes eg root knot nematodes, root cyst nematodes

Table A 16: Crop Sugarcane

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
Acetolactate synthase (ALS)	Sulfonylureas, Imidazolinones, Triazolopyrimidines, Pyrimidyloxybenzoates, Phtalides
AcetylCoA Carboxylase (ACCase)	Aryloxyphenoxyalkanecarboxylic acids, cyclohexanediones
Hydroxyphenylpyruvate dioxygenase (HPPD)	Isoxazoles such as Isoxaflutol or Isoxachlortol, Triones such as mesotrione or sulcotrione
Phosphinothricin acetyl transferase	Phosphinothricin
O-Methyl transferase	altered lignin levels
Glutamine synthetase	Glufosinate, Bialaphos
Adenylosuccinate Lyase (ADSL)	Inhibitors of IMP and AMP synthesis
Adenylosuccinate Synthase	Inhibitors of adenylosuccinate synthesis
Anthranilate Synthase	Inhibitors of tryptophan synthesis and catabolism
Nitrilase	3,5-dihalo-4-hydroxy-benzonitriles such as Bromoxynil and Ioxinyl
5-Enolpyruvyl-3phosphoshikimate Synthase (EPSPS)	Glyphosate or sulfosate
Glyphosate oxidoreductase	Glyphosate or sulfosate
Protoporphyrinogen oxidase (PROTOX)	Diphenylethers, cyclic imides, phenylpyrazoles, pyridin derivatives, phenopylate, oxadiazoles etc.
Cytochrome P450 eg. P450 SU1 or selection	Xenobiotics and herbicides such as Sulfonylureas
Polyphenol oxidase or Polyphenol oxidase antisense	bacterial or fungal pathogens
Metallothionein	bacterial or fungal pathogens
Ribonuclease	bacterial or fungal pathogens
Antifungal polypeptide AlyAFP	bacterial or fungal pathogens

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
oxalate oxidase	bacterial or fungal pathogens
glucose oxidase	bacterial or fungal pathogens
pyrrolnitrin synthesis genes	bacterial or fungal pathogens
serine/threonine kinases	bacterial or fungal pathogens
Cecropin B	bacterial or fungal pathogens
Phenylalanine ammonia lyase (PAL)	bacterial or fungal pathogens
Cf genes eg. Cf 9 Cf5 Cf4 Cf2	bacterial or fungal pathogens
Osmotin	bacterial or fungal pathogens
Alpha Hordothionin	bacterial or fungal pathogens
Systemin	bacterial or fungal pathogens
Polygalacturonase inhibitors	bacterial or fungal pathogens
Prf regulatory gene	bacterial or fungal pathogens
phytoalexins	bacterial or fungal pathogens
B-1,3-glucanase antisense	bacterial or fungal pathogens
receptor kinase	bacterial or fungal pathogens
Hypersensitive response eliciting polypeptide	bacterial or fungal pathogens
Systemic acquires resistance (SAR) genes	viral, bacterial, fungal, nematodal pathogens
Lytic protein	bacterial or fungal pathogens
Lysozym	bacterial or fungal pathogens eg clavibacter
Chitinases	bacterial or fungal pathogens
Barnase	bacterial or fungal pathogens
Glucanases	bacterial or fungal pathogens
double stranded ribonuclease	viruses as SCMV, SrMV
Coat proteins	viruses as SCMV, SrMV
17kDa or 60 kDa protein	viruses as SCMV, SrMV
Nuclear inclusion proteins eg. a or b or Nucleoprotein	viruses as SCMV, SrMV
Pseudoubiquitin	viruses as SCMV, SrMV
Replicase	viruses as SCMV, SrMV

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
Bacillus thuringiensis toxins, VIP 3, Bacillus cereus toxins, Photorabdus and Xenorhabdus toxins	lepidoptera, aphids, mites, nematodes, whitefly, beetles eg mexican rice borer
3- Hydroxysteroid oxidase	lepidoptera, aphids, mites, nematodes, whitefly, beetles eg mexican rice borer
Peroxidase	lepidoptera, aphids, mites. nematodes, whitefly, beetles eg mexican rice borer
Aminopeptidase inhibitors eg. Leucine aminopeptidase inhibitor	lepidoptera, aphids, mites, nematodes, whitefly, beetles eg mexican rice borer
Lectines	lepidoptera, aphids, mites, nematodes, whitefly, beetles eg mexican rice borer
Protease Inhibitors eg cystatin, patatin, CPTI, virgiferin	lepidoptera, aphids, mites, nematodes, whitefly, beetles eg mexican rice borer
ribosome inactivating protein	lepidoptera, aphids, mites, nematodes, whitefly, beetles eg mexican rice borer
stilbene synthase	lepidoptera, aphids, mites, nematodes, whitefly, beetles eg mexican rice borer
HMG-CoA reductase	lepidoptera, aphids, mites, nematodes, whitefly, beetles eg mexican rice borer
Cyst nematode hatching stimulus	cyst nematodes
Barnase	nematodes eg root knot nematodes and cyst nematodes
CBI	root knot nematodes
Antifeeding principles induced at a nematode feeding site	nematodes eg root knot nematodes, root cyst nematodes

Table A 17: Crop Sunflower

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
Acetolactate synthase (ALS)	Sulfonylureas, Imidazolinones,

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
	Triazolopyrimidines, Pyrimidyloxybenzoates, Phtalides
AcetylCoA Carboxylase (ACCase)	Aryloxyphenoxyalkanecarboxylic acids, cyclohexanediones
Hydroxyphenylpyruvate dioxygenase (HPPD)	Isoxazoles such as Isoxaflutol or Isoxachlortol, Triones such as mesotrione or sulcotrione
Phosphinothricin acetyl transferase	Phosphinothricin
O-Methyl transferase	altered lignin levels
Glutamine synthetase	Glufosinate, Bialaphos
Adenylosuccinate Lyase (ADSL)	Inhibitors of IMP and AMP synthesis
Adenylosuccinate Synthase	Inhibitors of adenylosuccinate synthesis
Anthranilate Synthase	Inhibitors of tryptophan synthesis and catabolism
Nitrilase	3,5-dihalo-4-hydroxy-benzonitriles such as Bromoxynil and Ioxinyl
5-Enolpyruvyl-3phosphoshikimate Synthase (EPSPS)	Glyphosate or sulfosate
Glyphosate oxidoreductase	Glyphosate or sulfosate
Protoporphyrinogen oxidase (PROTOX)	Diphenylethers, cyclic imides, phenylpyrazoles, pyridin derivatives, phenopylate, oxadiazoles etc.
Cytochrome P450 eg. P450 SU1 or selection	Xenobiotics and herbicides such as Sulfonylureas
Polyphenol oxidase or Polyphenol oxidase antisense	bacterial or fungal pathogens
Metallothionein	bacterial or fungal pathogens
Ribonuclease	bacterial or fungal pathogens
Antifungal polypeptide AlyAFP	bacterial or fungal pathogens
oxalate oxidase	bacterial or fungal pathogens eg sclerotinia
glucose oxidase	bacterial or fungal pathogens

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
pyrrolnitrin synthesis genes	bacterial or fungal pathogens
serine/threonine kinases	bacterial or fungal pathogens
Cecropin B	bacterial or fungal pathogens
Phenylalanine ammonia lyase (PAL)	bacterial or fungal pathogens
Cf genes eg. Cf 9 Cf5 Cf4 Cf2	bacterial or fungal pathogens
Osmotin	bacterial or fungal pathogens
Alpha Hordothionin	bacterial or fungal pathogens
Systemin	bacterial or fungal pathogens
Polygalacturonase inhibitors	bacterial or fungal pathogens
Prf regulatory gene	bacterial or fungal pathogens
phytoalexins	bacterial or fungal pathogens
B-1,3-glucanase antisense	bacterial or fungal pathogens
receptor kinase	bacterial or fungal pathogens
Hypersensitive response eliciting polypeptide	bacterial or fungal pathogens
Systemic acquires resistance (SAR) genes	viral, bacterial, fungal, nematodal pathogens
Lytic protein	bacterial or fungal pathogens
Lysozym	bacterial or fungal pathogens
Chitinases	bacterial or fungal pathogens
Barnase	bacterial or fungal pathogens
Glucanases	bacterial or fungal pathogens
double stranded ribonuclease	viruses as CMV, TMV
Coat proteins	viruses as CMV, TMV
17kDa or 60 kDa protein	viruses as CMV, TMV
Nuclear inclusion proteins eg. a or b or Nucleoprotein	viruses as CMV, TMV
Pseudoubiquitin	viruses as CMV, TMV
Replicase	viruses as CMV, TMV
Bacillus thuringiensis toxins, VIP 3, Bacillus cereus toxins, Photorabdus and Xenorhabdus toxins	lepidoptera, aphids, mites, nematodes, whitefly, beetles

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
3- Hydroxysteroid oxidase	lepidoptera, aphids, mites, nematodes, whitefly, beetles
Peroxidase	lepidoptera, aphids, mites, nematodes, whitefly, beetles
Aminopeptidase inhibitors eg. Leucine aminopeptidase inhibitor	lepidoptera, aphids, mites, nematodes, whitefly, beetles
Lectines	lepidoptera, aphids, mites, nematodes, whitefly, beetles
Protease Inhibitors eg cystatin, patatin, CPTI, virgiferin	lepidoptera, aphids, mites, nematodes, whitefly, beetles
ribosome inactivating protein	lepidoptera, aphids, mites, nematodes, whitefly, beetles
stilbene synthase	lepidoptera, aphids, mites, nematodes, whitefly, beetles
HMG-CoA reductase	lepidoptera, aphids, mites, nematodes, whitefly, beetles
Cyst nematode hatching stimulus	cyst nematodes
Barnase	nematodes eg root knot nematodes and cyst nematodes
CBI	root knot nematodes
Antifeeding principles induced at a nematode feeding site	nematodes eg root knot nematodes, root cyst nematodes

Table A 18: Crop Sugarbeet, Beet root

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
Acetolactate synthase (ALS)	Sulfonylureas, Imidazolinones, Triazolopyrimidines, Pyrimidyloxybenzoates, Phtalides
AcetylCoA Carboxylase (ACCase)	Aryloxyphenoxyalkanecarboxylic acids,

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
	cyclohexanediones
Hydroxyphenylpyruvate dioxygenase (HPPD)	Isoxazoles such as Isoxaflutol or Isoxachlortol, Triones such as mesotrione or sulcotrione
Phosphinothricin acetyl transferase	Phosphinothricin
O-Methyl transferase	altered lignin levels
Glutamine synthetase	Glufosinate, Bialaphos
Adenylosuccinate Lyase (ADSL)	Inhibitors of IMP and AMP synthesis
Adenylosuccinate Synthase	Inhibitors of adenylosuccinate synthesis
Anthranilate Synthase	Inhibitors of tryptophan synthesis and catabolism
Nitrilase	3,5-dihalo-4-hydroxy-benzonitriles such as Bromoxynil and Ioxinyl
5-Enolpyruvyl-3phosphoshikimate Synthase (EPSPS)	Glyphosate or sulfosate
Glyphosate oxidoreductase	Glyphosate or sulfosate
Protoporphyrinogen oxidase (PROTOX)	Diphenylethers, cyclic imides, phenylpyrazoles, pyridin derivatives, phenopylate, oxadiazoles etc.
Cytochrome P450 eg. P450 SU1 or selection	Xenobiotics and herbicides such as Sulfonylureas
Polyphenol oxidase or Polyphenol oxidase antisense	bacterial or fungal pathogens
Metallothionein	bacterial or fungal pathogens
Ribonuclease	bacterial or fungal pathogens
Antifungal polypeptide AlyAFP	bacterial or fungal pathogens
oxalate oxidase	bacterial or fungal pathogens eg sclerotinia
glucose oxidase	bacterial or fungal pathogens
pyrrolnitrin synthesis genes	bacterial or fungal pathogens
serine/threonine kinases	bacterial or fungal pathogens
Cecropin B	bacterial or fungal pathogens

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
Phenylalanine ammonia lyase (PAL)	bacterial or fungal pathogens
Cf genes eg. Cf 9 Cf5 Cf4 Cf2	bacterial or fungal pathogens
Osmotin	bacterial or fungal pathogens
Alpha Hordothionin	bacterial or fungal pathogens
Systemin	bacterial or fungal pathogens
Polygalacturonase inhibitors	bacterial or fungal pathogens
Prf regulatory gene	bacterial or fungal pathogens
phytoalexins	bacterial or fungal pathogens
B-1,3-glucanase antisense	bacterial or fungal pathogens
AX + WIN proteins	bacterial or fungal pathogens like Cercospora beticola
receptor kinase	bacterial or fungal pathogens
Hypersensitive response eliciting polypeptide	bacterial or fungal pathogens
Systemic acquires resistance (SAR) genes	viral, bacterial, fungal, nematodal pathogens
Lytic protein	bacterial or fungal pathogens
Lysozym	bacterial or fungal pathogens
Chitinases	bacterial or fungal pathogens
Barnase	bacterial or fungal pathogens
Glucanases	bacterial or fungal pathogens
double stranded ribonuclease	viruses as BNYVV
Coat proteins	viruses as BNYVV
17kDa or 60 kDa protein	viruses as BNYVV
Nuclear inclusion proteins eg. a or b or Nucleoprotein	viruses as BNYVV
Pseudoubiquitin	viruses as BNYVV
Replicase	viruses as BNYVV
Bacillus thuringiensis toxins, VIP 3, Bacillus cereus toxins, Photorabdus and Xenorhabdus toxins	lepidoptera, aphids, mites, nematodes, whitefly, beetles, rootflies
3- Hydroxysteroid oxidase	lepidoptera, aphids, mites, nematodes,

Effected target or expressed principle(s)	Crop phenotype / Tolerance to
	whitefly, beetles, rootflies
Peroxidase	lepidoptera, aphids, mites, nematodes, whitefly, beetles, rootflies
Aminopeptidase inhibitors eg. Leucine aminopeptidase inhibitor	lepidoptera, aphids, mites, nematodes, whitefly, beetles, rootflies
Lectines	lepidoptera, aphids, mites, nematodes, whitefly, beetles, rootflies
Protease Inhibitors eg cystatin, patatin, CPTI, virgiferin	lepidoptera, aphids, mites, nematodes, whitefly, beetles, rootflies
ribosome inactivating protein	lepidoptera, aphids, mites, nematodes, whitefly, beetles, rootflies
stilbene synthase	lepidoptera, aphids, mites, nematodes, whitefly, beetles, rootflies
HMG-CoA reductase	lepidoptera, aphids, mites, nematodes, whitefly, beetles, rootflies
Cyst nematode hatching stimulus	cyst nematodes
Barnase	nematodes eg root knot nematodes and cyst nematodes
Beet cyst nematode resistance locus	cyst nematodes
CBI	root knot nematodes
Antifeeding principles induced at a nematode feeding site	nematodes eg root knot nematodes, root cyst nematodes

The abovementioned animal pests which can be controlled by the method according to the invention include; for example, insects, representatives of the order acarina and representatives of the class nematoda; especially

from the order Lepidoptera Acleris spp., Adoxophyes spp., especially Adoxophyes reticulana; Aegeria spp., Agrotis spp., especially Agrotis spinifera; Alabama argillaceae, Amylois spp., Anticarsia gemmatalis, Archips spp., Argyrotaenia spp., Autographa spp., Busseola fusca, Cadra cautella, Carposina nipponensis, Chilo spp., Choristoneura spp., Clysia ambiguella, Cnaphalocrocis spp., Cnephasia spp., Cochylis spp., Coleophora spp., Crocidolomia binotalis, Cryptophlebia leucotreta, Cydia spp., especially Cydia pomonella;

Diatraea spp., Diparopsis castanea, Earias spp., Ephestia spp., especially E. Khüniella; Eucosma spp., Eupoecilia ambiguella, Euproctis spp., Euxoa spp., Grapholita spp., Hedya nubiferana, Heliothis spp., especially H. Virescens und H. zea; Hellula undalis, Hyphantria cunea, Keiferia lycopersicella, Leucoptera scitella, Lithocollethis spp., Lobesiaspp., Lymantria spp., Lyonetia spp., Malacosoma spp., Mamestra brassicae, Manduca sexta, Operophtera spp., Ostrinia nubilalis, Pammene spp., Pandemis spp., Panolis flammea, Pectinophora spp.. Phthorimaea operculella, Pieris rapae, Pieris spp., Plutella xylostella, Prays spp., Scirpophaga spp., Sesamia spp., Sparganothis spp., Spodopteralittoralis, Synanthedon spp., Thaumetopoea spp., Tortrix spp., Trichoplusia ni and Yponomeuta spp.;

from the order Coleoptera, for example Agriotes spp., Anthonomus spp., Atomaria linearis, Chaetocnema tibialis, Cosmopolites spp., Curculio spp., Dermestes spp., Diabrotica spp., Epilachna spp., Eremnus spp., Leptinotarsa decemlineata, Lissorhoptrus spp., Melolontha spp., Oryzaephilus spp., Otiorhynchus spp., Phlyctinus spp., Popillia spp., Psylliodes spp., Rhizopertha spp., Scarabeidae, Sitophilus spp., Sitotroga spp., Tenebrio spp., Tribolium spp. and Trogoderma spp.;

from the order Orthoptera, for example Blatta spp., Blattella spp., Gryllotalpa spp., Leucophaea maderae, Locusta spp., Periplaneta spp. and Schistocerca spp.;

from the order Isoptera, for example Reticulitermes spp.;

from the order Psocoptera, for example Liposcelis spp.;

from the order Anoplura, for example Haematopinus spp., Linognathus spp., Pediculus spp., Pemphigus spp. and Phylloxera spp.;

from the order Mallophaga, for example Damalinea spp. and Trichodectes spp.;

from the order Thysanoptera, for example Frankliniella spp., Hercinothrips spp., Taeniothrips spp., Thrips palmi, Thrips tabaci and Scirtothrips aurantii;

from the order Heteroptera, for example Cimex spp., Distantiella theobroma, Dysdercus spp., Euchistus spp. Eurygaster spp. Leptocorisa spp., Nezara spp., Piesma spp., Rhodnius spp., Sahlbergella singularis, Scotinophara spp. and Triatoma spp.;

from the order Homoptera, for example Aleurothrixus floccosus, Aleyrodes brassicae, Aonidiella aurantii, Aphididae, Aphis craccivora, A. fabae, A. gosypii; Aspidiotus spp., Bemisia tabaci, Ceroplaster spp., Chrysomphalus aonidium, Chrysomphalus dictyospermi, Coccus hesperidum, Empoasca spp., Eriosoma lanigerum, Erythroneura spp., Gascardia

spp., Laodelphax spp., Lecanium corni, Lepidosaphes spp., Macrosiphus spp., Myzus spp., especially M.persicae; Nephotettix spp., especially N. cincticeps; Nilaparvata spp., especially N. lugens; Paratoria spp., Pemphigus spp., Planococcus spp., Pseudaulacaspis spp., Pseudococcus spp., especially P. Fragilis, P. citriculus and P. comstocki; Psylla spp., especially P. pyri; Pulvinaria aethiopica, Quadraspidiotus spp., Rhopalosiphum spp., Saissetia spp., Scaphoideus spp., Schizaphis spp., Sitobion spp., Trialeurodes vaporariorum, Trioza erytreae and Unaspis citri;

from the order Hymenoptera, for example Acromyrmex, Atta spp., Cephus spp., Diprion spp., Diprionidae, Gilpinia polytoma, Hoplocampa spp., Lasius spp., Monomorium pharaonis, Neodiprion spp., Solenopsis spp. and Vespa spp.;

from the order Diptera, for example Aedes spp., Antherigona soccata, Bibio hortulanus, Calliphora erythrocephala, Ceratitis spp., Chrysomyia spp., Culex spp., Cuterebra spp., Dacus spp., Drosophila melanogaster, Fannia spp., Gastrophilus spp., Glossina spp., Hypoderma spp., Hyppobosca spp., Liriomyza spp., Lucilia spp., Melanagromyza spp., Musca spp., Oestrus spp., Orseolia spp., Oscinella frit, Pegomyia hyoscyami, Phorbia spp., Rhagoletis pomonella, Sciara spp., Stomoxys spp., Tabanus spp., Tannia spp. and Tipula spp.;

from the order Siphonaptera, for example Ceratophyllus spp. and Xenopsylla cheopis;

from the order Thysanura, for example Lepisma saccharina and

from the order Acarina, for example Acarus siro, Aceria sheldoni; Aculus spp., especially A. schlechtendali; Amblyomma spp., Argas spp., Boophilus spp., Brevipalpus spp., especially B. californicus and B. phoenicis; Bryobia praetiosa, Calipitrimerus spp., Chorioptes spp., Dermanyssus gallinae, Eotetranychus spp., especially E.carpini and E. orientalis; Eriophyes spp., especially E. vitis; Hyalomma spp., Ixodes spp., Olygonychus pratensis, Ornithodoros spp., Panonychus spp., especially P. ulmi and P. citri; Phyllocoptruta spp., especially P. oleivora; Polyphagotarsonemus spp., especially P. latus; Psoroptes spp., Rhipicephalus spp., Rhizoglyphus spp., Sarcoptes spp., Tarsonemus spp. and Tetranychus spp., in particular T. urticae, T. cinnabarinus and T. Kanzawai;

representatives of the class *Nematoda;*

(1) nematodes selected from the group consisting of root knot nematodes, cyst-forming nematodes, stem eelworms and foliar nematodes;

(2) nematodes selected from the group consisting of Anguina spp.; Aphelenchoides spp.; Ditylenchus spp.; Globodera spp., for example Globodera rostochiensis; Heterodera spp., for example Heterodera avenae, Heterodera glycines, Heterodera schachtii or Heterodera trifolii; Longidorus spp.; Meloidogyne spp., for example Meloidogyne incognita or Meloidogyne javanica; Pratylenchus, for example Pratylenchus neglectans or Pratylenchus penetrans; Radopholus spp., for example Radopholus similis; Trichodorus spp.; Tylenchulus, for example Tylenchulus semipenetrans; and Xiphinema spp.; or

(3) nematodes selected from the group consisting of Heterodera spp., for example Heterodera glycines; and Meloidogyne spp., for example Meloidogyne incognita.

The method according to the invention allows pests of the abovementioned type to be controlled, i.e. contained or destroyed, which occur, in particular, on transgenic plants, mainly useful plants and ornamentals in agriculture, in horticulture and in forests, or on parts, such as fruits, flowers, foliage, stalks, tubers or roots, of such plants, the protection against these pests in some cases even extending to plant parts which form at a later point in time.

The method according to the invention can be employed advantageously for controlling pests in rice, cereals such as maize or sorghum; in fruit, for example stone fruit, pome fruit and soft fruit such as apples, pears, plums, peaches, almonds, cherries or berries, for example strawberries, raspberries and blackberries; in legumes such as beans, lentils, peas or soya beans; in oil crops such as oilseed rape, mustard, poppies, olives, sunflowers, coconuts, castor-oil plants, cacao or peanuts; in the marrow family such as pumpkins, cucumbers or melons; in fibre plants such as cotton, flax, hemp or jute; in citrus fruit such as oranges, lemons, grapefruit or tangerines; in vegetables such as spinach, lettuce, asparagus, cabbage species, carrots, onions, tomatoes, potatoes, beet or capsicum; in the laurel family such as avocado, Cinnamonium or camphor; or in tobacco, nuts, coffee, egg plants, sugar cane, tea, pepper, grapevines, hops, the banana family, latex plants or ornamentals, mainly in maize, rice, cereals, soya beans, tomatoes, cotton, potatoes, sugar beet, rice and mustard; in particular in cotton, rice, soya beans, potatoes and maize.

It has emerged that the method according to the invention is valuable preventatively and/or curatively in the field of pest control even at low use concentrations of the pesticidal composition and that a very favourable biocidal spectrum is achieved thereby. Combined with a favourable compatibility of the composition employed with warm-blooded species,

fish and plants, the method according to the invention can be employed against all or individual developmental stages of normally-sensitive, but also of normally-resistant, animal pests such as insects and representatives of the order Acarina, depending on the species of the transgenic crop plant to be protected from attack by pests. The insecticidal and/or acaricidal effect of the method according to the invention may become apparent directly, i.e. in a destruction of the pests which occurs immediately or only after some time has elapsed, for example, during ecdysis. or indirectly, for example as a reduced oviposition and/or hatching rate, the good action corresponding to a destruction rate (mortality) of at least 40 to 50%.

Depending on the intended aims and the prevailing circumstances, the pesticides within the scope of invention , which are known per se, are emulsifiable concentrates, suspension concentrates, directly sprayable or dilutable solutions, spreadable pastes, dilute emulsions, wettable powders, soluble powders, dispersible powders, wettable powders, dusts, granules or encapsulations in polymeric substances which comprise a nitroimino- or nitroguanidino-compound.

The active ingredients are employed in these compositions together with at least one of the auxiliaries conventionally used in art of formulation, such as extenders, for example solvents or solid carriers, or such as surface-active compounds (surfactants).

Formulation auxiliaries which are used are, for example, solid carriers, solvents, stabilizers, "slow release" auxiliaries, colourants and, if appropriate, surface-active substances (surfactants). Suitable carriers and auxiliaries are all those substances which are conventionally used for crop protection products. Suitable auxiliaries such as solvents, solid carriers, surface-active compounds, non-ionic surfactants, cationic surfactants, anionic surfactants and other auxiliaries in the compositions employed according to the invention are, for example, those which have been described in EP-A-736 252.

These compositions for controlling pests can be formulated, for example, as wettable powders, dusts, granules, solutions, emulsifiable concentrates, emulsions, suspension concentrates or aerosols. For example, the compositions are of the type described in EP-A-736 252.

The action of the compositions within the scope of invention which comprise a nitroimino- or nitroguanidino-compound can be extended substantially and adapted to prevailing circumstances by adding other insecticidally, acaricidally and/or fungicidally active

ingredients. Suitable examples of added active ingredients are representatives of the following classes of active ingredients: organophosphorous compounds, nitrophenols and derivatives, formamidines, ureas, carbamates, pyrethroids, chlorinated hydrocarbons; especially preferred components in mixtures are, for example, abamectin, emamectin, spinosad, pymetrozine, fenoxycarb, Ti-435, fipronil, pyriproxyfen, diazinon or diafenthiuron.

As a rule, the compositions within the scope of invention comprise 0.1 to 99%, in particular 0.1 to 95 %, of a nitroimino- or nitroguanidino-compound and 1 to 99.9 %, in particular 5 to 99.9 %, of - at least - one solid or liquid auxiliary, it being possible, as a rule, for 0 to 25 %, in particular 0.1 to 20 %, of the compositions to be surfactants (% in each case meaning per cent by weight). While concentrated compositions are more preferred as commercial products, the end user will, as a rule, use dilute compositions which have considerably lower concentrations of active ingredient.

The compositions according to the invention may also comprise other solid or liquid auxiliaries, such as stabilisers, for example epoxidized or unepoxidized vegetable oils (for example epoxidized coconut oil, rapeseed oil or soya bean oil), antifoams, for example silicone oil, preservatives, viscosity regulators, binders and/or tackifiers, and also fertilizers or other active ingredients for achieving specific effects, for example, bactericides, fungicides, nematicides, molluscicides or herbicides.

The compositions according to the invention are produced in a known manner, for example prior to mixing with the auxiliary/auxiliaries by grinding, screening and/or compressing the active ingredient, for example to give a particular particle size, and by intimately mixing and/or grinding the active ingredient with the auxiliary/auxiliaries.

The method according to the invention for controlling pests of the abovementioned type is carried out in a manner known per se to those skilled in the art, depending on the intended aims and prevailing circumstances, that is to say by spraying, wetting, atomizing, dusting, brushing on, seed dressing, scattering or pouring of the composition. Typical use concentrations are between 0.1 and 1000 ppm, preferably between 0.1 and 500 ppm of active ingredient. The application rate may vary within wide ranges and depends on the soil constitution, the type of application (foliar application; seed dressing; application in the seed furrow), the transgenic crop plant, the pest to be controlled, the climatic circumstances prevailing in each case, and other factors determined by the type of application, timing of application and target crop. The application rates per hectare are generally 1 to 2000 g of

nitroimino- or nitroguanidino-compound per hectare, in particular 10 to 1000 g/ha, preferably 10 to 500 g/ha, especially preferably 10 to 200 g/ha.

A preferred type of application in the field of crop protection within the scope of invention is application to the foliage of the plants (foliar application), it being possible to adapt frequency and rate of application to the risk of infestation with the pest in question.. However, the active ingredient may also enter into the plants via the root system (systemic action), by drenching the site of the plants with a liquid composition or by incorporating the active ingredient in solid form into the site of the plants, for example into the soil, for example in the form of granules (soil application). In the case of paddy rice crops, such granules may be metered into the flooded paddy field.

The compositions according to invention are also suitable for protecting propagation material of transgenic plants, for example seed, such as fruits, tubers or kernels, or plant cuttings, from animal pests, in particular insects and representatives of the order Acarina. The propagation material can be treated with the composition prior to application, for example, seed being dressed prior to sowing. The active ingredient may also be applied to seed kernels (coating), either by soaking the kernels in a liquid composition or by coating them with a solid composition. The composition may also be applied to the site of application when applying the propagation material, for example into the seed furrow during sowing. These treatment methods for plant propagation material and the plant propagation material treated thus are a further subject of the invention.

Examples of formulations of nitroimino- or nitroguanidino-compounds which can be used in the method according to the invention , for instance solutions, granules, dusts, sprayable powders, emulsion concentrates, coated granules and suspension concentrates, are of the type as has been described in, for example, EP-A-580 553, Examples F1 to F10.

Biological examples

Table B

The following abreviations are used in the table:
Active Principle of transgenic plant: AP
Photorhabdus luminescens: PL
Xenorhabdus nematophilus: XN
Proteinase Inhibitors: PInh.

Plant lectins PLec.

Agglutinins: Aggl.

3-Hydroxysteroid oxidase: HO

Cholesteroloxidase: CO

Chitinase: CH

Glucanase: GL

Stilbensynthase SS

Table B:

	AP	Control of
B.1	CrylA(a)	Adoxophyes spp.
B.2	CrylA(a)	Agrotis spp.
B.3	CrylA(a)	Alabama argillaceae
B.4	CrylA(a)	Anticarsia gemmatalis
B.5	CrylA(a)	Chilo spp.
B.6	CrylA(a)	Clysia ambiguella
B.7	CrylA(a)	Crocidolomia binotalis
B.8	CrylA(a)	Cydia spp.
B.9	CrylA(a)	Diparopsis castanea
B.10	CrylA(a)	Earias spp.
B.11	CrylA(a)	Ephestia spp.
B.12	CrylA(a)	Heliothis spp.
B.13	CrylA(a)	Hellula undalis
B.14	CrylA(a)	Keiferia lycopersicella
B.15	CrylA(a)	Leucoptera scitella
B.16	CrylA(a)	Lithocollethis spp.
B.17	CrylA(a)	Lobesia botrana
B.18	CrylA(a)	Ostrinia nubilalis
B.19	CrylA(a)	Pandemis spp.
B.20	CrylA(a)	Pectinophora gossyp.
B.21	CrylA(a)	Phyllocnistis citrella
B.22	CrylA(a)	Pieris spp.
B.23	CrylA(a)	Plutella xylostella
B.24	CrylA(a)	Scirpophaga spp.
B.25	CrylA(a)	Sesamia spp.
B.26	CrylA(a)	Sparganothis spp.
B.27	CrylA(a)	Spodoptera spp.
B.28	CrylA(a)	Tortrix spp.
B.29	CrylA(a)	Trichoplusia ni
B.30	CrylA(a)	Agriotes spp.
B.31	CrylA(a)	Anthonomus grandis
B.32	CrylA(a)	Curculio spp.
B.33	CrylA(a)	Diabrotica balteata
B.34	CrylA(a)	Leptinotarsa spp.
B.35	CrylA(a)	Lissorhoptrus spp.
B.36	CrylA(a)	Otiorhynchus spp.
B.37	CrylA(a)	Aleurothrixus spp.

	AP	Control of
B.38	CrylA(a)	Aleyrodes spp.
B.39	CrylA(a)	Aonidiella spp.
B.40	CrylA(a)	Aphididae spp.
B.41	CrylA(a)	Aphis spp.
B.42	CrylA(a)	Bemisia tabaci
B.43	CrylA(a)	Empoasca spp.
B.44	CrylA(a)	Mycus spp.
B.45	CrylA(a)	Nephotettix spp.
B.46	CrylA(a)	Nilaparvata spp.
B.47	CrylA(a)	Pseudococcus spp.
B.48	CrylA(a)	Psylla spp.
B.49	CrylA(a)	Quadraspidiotus spp.
B.50	CrylA(a)	Schizaphis spp.
B.51	CrylA(a)	Trialeurodes spp.
B.52	CrylA(a)	Lyriomyza spp.
B.53	CrylA(a)	Oscinella spp.
B.54	CrylA(a)	Phorbia spp.
B.55	CrylA(a)	Frankliniella spp.
B.56	CrylA(a)	Thrips spp.
B.57	CrylA(a)	Scirtothrips aurantii
B.58	CrylA(a)	Aceria spp.
B.59	CrylA(a)	Aculus spp.
B.60	CrylA(a)	Brevipalpus spp.
B.61	CrylA(a)	Panonychus spp.
B.62	CrylA(a)	Phyllocoptruta spp.
B.63	CrylA(a)	Tetranychus spp.
B.64	CrylA(a)	Heterodera spp.
B.65	CrylA(a)	Meloidogyne spp.
B.66	CrylA(b)	Adoxophyes spp.
B.67	CrylA(b)	Agrotis spp.
B.68	CrylA(b)	Alabama argillaceae
B.69	CrylA(b)	Anticarsia gemmatalis
B.70	CrylA(b)	Chilo spp.
B.71	CrylA(b)	Clysia ambiguella
B.72	CrylA(b)	Crocidolomia binotalis
B.73	CrylA(b)	Cydia spp.
B.74	CrylA(b)	Diparopsis castanea
B.75	CrylA(b)	Earias spp.
B.76	CrylA(b)	Ephestia spp.
B.77	CrylA(b)	Heliothis spp.
B.78	CrylA(b)	Hellula undalis
B.79	CrylA(b)	Keiferia lycopersicella
B.80	CrylA(b)	Leucoptera scitella
B.81	CrylA(b)	Lithocollethis spp.
B.82	CrylA(b)	Lobesia botrana
B.83	CrylA(b)	Ostrinia nubilalis
B.84	CrylA(b)	Pandemis spp.
B.85	CrylA(b)	Pectinophora gossyp.
B.86	CrylA(b)	Phyllocnistis citrella
B.87	CrylA(b)	Pieris spp.
B.88	CrylA(b)	Plutella xylostella
B.89	CrylA(b)	Scirpophaga spp.
B.90	CrylA(b)	Sesamia spp.
B.91	CrylA(b)	Sparganothis spp.
B.92	CrylA(b)	Spodoptera spp.
B.93	CrylA(b)	Tortrix spp.
B.94	CrylA(b)	Trichoplusia ni

	AP	Control of
B.95	CrylA(b)	Agriotes spp.
B.96	CrylA(b)	Anthonomus grandis
B.97	CrylA(b)	Curculio spp.
B.98	CrylA(b)	Diabrotica balteata
B.99	CrylA(b)	Leptinotarsa spp.
B.100	CrylA(b)	Lissorhoptrus spp.
B.101	CrylA(b)	Otiorhynchus spp.
B.102	CrylA(b)	Aleurothrixus spp.
B.103	CrylA(b)	Aleyrodes spp.
B.104	CrylA(b)	Aonidiella spp.
B.105	CrylA(b)	Aphididae spp.
B.106	CrylA(b)	Aphis spp.
B.107	CrylA(b)	Bemisia tabaci
B.108	CrylA(b)	Empoasca spp.
B.109	CrylA(b)	Mycus spp.
B.110	CrylA(b)	Nephotettix spp.
B.111	CrylA(b)	Nilaparvata spp.
B.112	CrylA(b)	Pseudococcus spp.
B.113	CrylA(b)	Psylla spp.
B.114	CrylA(b)	Quadraspidiotus spp.
B.115	CrylA(b)	Schizaphis spp.
B.116	CrylA(b)	Trialeurodes spp.
B.117	CrylA(b)	Lyriomyza spp.
B.118	CrylA(b)	Oscinella spp.
B.119	CrylA(b)	Phorbia spp.
B.120	CrylA(b)	Frankliniella spp.
B.121	CrylA(b)	Thrips spp.
B.122	CrylA(b)	Scirtothrips aurantii
B.123	CrylA(b)	Aceria spp.
B.124	CrylA(b)	Aculus spp.
B.125	CrylA(b)	Brevipalpus spp.
B.126	CrylA(b)	Panonychus spp.
B.127	CrylA(b)	Phyllocoptruta spp.
B.128	CrylA(b)	Tetranychus spp.
B.129	CrylA(b)	Heterodera spp.
B.130	CrylA(b)	Meloidogyne spp.
B.131	CrylA(c)	Adoxophyes spp.
B.132	CrylA(c)	Agrotis spp.
B.133	CrylA(c)	Alabama argillaceae
B.134	CrylA(c)	Anticarsia gemmatalis
B.135	CrylA(c)	Chilo spp.
B.136	CrylA(c)	Clysia ambiguella
B.137	CrylA(c)	Crocidolomia binotalis
B.138	CrylA(c)	Cydia spp.
B.139	CrylA(c)	Diparopsis castanea
B.140	CrylA(c)	Earias spp.
B.141	CrylA(c)	Ephestia spp.
B.142	CrylA(c)	Heliothis spp.
B.143	CrylA(c)	Hellula undalis
B.144	CrylA(c)	Keiferia lycopersicella
B.145	CrylA(c)	Leucoptera scitella
B.146	CrylA(c)	Lithocollethis spp.
B.147	CrylA(c)	Lobesia botrana
B.148	CrylA(c)	Ostrinia nubilalis
B.149	CrylA(c)	Pandemis spp.
B.150	CrylA(c)	Pectinophora gossypiella.

	AP	Control of
B.151	CrylA(c)	Phyllocnistis citrella
B.152	CrylA(c)	Pieris spp.
B.153	CrylA(c)	Plutella xylostella
B.154	CrylA(c)	Scirpophaga spp.
B.155	CrylA(c)	Sesamia spp.
B.156	CrylA(c)	Sparganothis spp.
B.157	CrylA(c)	Spodoptera spp.
B.158	CrylA(c)	Tortrix spp.
B.159	CrylA(c)	Trichoplusia ni
B.160	CrylA(c)	Agriotes spp.
B.161	CrylA(c)	Anthonomus grandis
B.162	CrylA(c)	Curculio spp.
B.163	CrylA(c)	Diabrotica balteata
B.164	CrylA(c)	Leptinotarsa spp.
B.165	CrylA(c)	Lissorhoptrus spp.
B.166	CrylA(c)	Otiorhynchus spp.
B.167	CrylA(c)	Aleurothrixus spp.
B.168	CrylA(c)	Aleyrodes spp.
B.169	CrylA(c)	Aonidiella spp.
B.170	CrylA(c)	Aphididae spp.
B.171	CrylA(c)	Aphis spp.
B.172	CrylA(c)	Bemisia tabaci
B.173	CrylA(c)	Empoasca spp.
B.174	CrylA(c)	Mycus spp.
B.175	CrylA(c)	Nephotettix spp.
B.176	CrylA(c)	Nilaparvata spp.
B.177	CrylA(c)	Pseudococcus spp.
B.178	CrylA(c)	Psylla spp.
B.179	CrylA(c)	Quadraspidiotus spp.
B.180	CrylA(c)	Schizaphis spp.
B.181	CrylA(c)	Trialeurodes spp.
B.182	CrylA(c)	Lyriomyza spp.
B.183	CrylA(c)	Oscinella spp.
B.184	CrylA(c)	Phorbia spp.
B.185	CrylA(c)	Frankliniella spp.
B.186	CrylA(c)	Thrips spp.
B.187	CrylA(c)	Scirtothrips aurantii
B.188	CrylA(c)	Aceria spp.
B.189	CrylA(c)	Aculus spp.
B.190	CrylA(c)	Brevipalpus spp.
B.191	CrylA(c)	Panonychus spp.
B.192	CrylA(c)	Phyllocoptruta spp.
B.193	CrylA(c)	Tetranychus spp.
B.194	CrylA(c)	Heterodera spp.
B.195	CrylA(c)	Meloidogyne spp.
B.196	CrylIA	Adoxophyes spp.
B.197	CrylIA	Agrotis spp.
B.198	CrylIA	Alabama argillaceae
B.199	CrylIA	Anticarsia gemmatalis
B.200	CrylIA	Chilo spp.
B.201	CrylIA	Clysia ambiguella
B.202	CrylIA	Crocidolomia binotalis
B.203	CrylIA	Cydia spp.
B.204	CrylIA	Diparopsis castanea
B.205	CrylIA	Earias spp.
B.206	CrylIA	Ephestia spp.
B.207	CrylIA	Heliothis spp.
B.208	CrylIA	Hellula undalis

	AP	Control of
B.209	CryIIA	Keiferia lycopersicella
B.210	CryIIA	Leucoptera scitella
B.211	CryIIA	Lithocollethis spp.
B.212	CryIIA	Lobesia botrana
B.213	CryIIA	Ostrinia nubilalis
B.214	CryIIA	Pandemis spp.
B.215	CryIIA	Pectinophora gossyp.
B.216	CryIIA	Phyllocnistis citrella
B.217	CryIIA	Pieris spp.
B.218	CryIIA	Plutella xylostella
B.219	CryIIA	Scirpophaga spp.
B.220	CryIIA	Sesamia spp.
B.221	CryIIA	Sparganothis spp.
B.222	CryIIA	Spodoptera spp.
B.223	CryIIA	Tortrix spp.
B.224	CryIIA	Trichoplusia ni
B.225	CryIIA	Agriotes spp.
B.226	CryIIA	Anthonomus grandis
B.227	CryIIA	Curculio spp.
B.228	CryIIA	Diabrotica balteata
B.229	CryIIA	Leptinotarsa spp.
B.230	CryIIA	Lissorhoptrus spp.
B.231	CryIIA	Otiorhynchus spp.
B.232	CryIIA	Aleurothrixus spp.
B.233	CryIIA	Aleyrodes spp.
B.234	CryIIA	Aonidiella spp.
B.235	CryIIA	Aphididae spp.
B.236	CryIIA	Aphis spp.
B.237	CryIIA	Bemisia tabaci
B.238	CryIIA	Empoasca spp.
B.239	CryIIA	Mycus spp.
B.240	CryIIA	Nephotettix spp.
B.241	CryIIA	Nilaparvata spp.
B.242	CryIIA	Pseudococcus spp.
B.243	CryIIA	Psyila spp.
B.244	CryIIA	Quadraspidiotus spp.
B.245	CryIIA	Schizaphis spp.
B.246	CryIIA	Trialeurodes spp.
B.247	CryIIA	Lyriomyza spp.
B.248	CryIIA	Oscinella spp.
B.249	CryIIA	Phorbia spp.
B.250	CryIIA	Frankliniella spp.
B.251	CryIIA	Thrips spp.
B.252	CryIIA	Scirtothrips aurantii
B.253	CryIIA	Aceria spp.
B.254	CryIIA	Aculus spp.
B.255	CryIIA	Brevipalpus spp.
B.256	CryIIA	Panonychus spp.
B.257	CryIIA	Phyllocoptruta spp.
B.258	CryIIA	Tetranychus spp.
B.259	CryIIA	Heterodera spp.
B.260	CryIIA	Meloidogyne spp.
B.261	CryIIIA	Adoxophyes spp.
B.262	CryIIIA	Agrotis spp.
B.263	CryIIIA	Alabama argillaceae
B.264	CryIIIA	Anticarsia gemmatalis
B.265	CryIIIA	Chilo spp.
B.266	CryIIIA	Clysia ambiguella

	AP	Control of
B.267	CryIIIA	Crocidolomia binotalis
B.268	CryIIIA	Cydia spp.
B.269	CryIIIA	Diparopsis castanea
B.270	CryIIIA	Earias spp.
B.271	CryIIIA	Ephestia spp.
B.272	CryIIIA	Heliothis spp.
B.273	CryIIIA	Hellula undalis
B.274	CryIIIA	Keiferia lycopersicella
B.275	CryIIIA	Leucoptera scitella
B.276	CryIIIA	Lithocollethis spp.
B.277	CryIIIA	Lobesia botrana
B.278	CryIIIA	Ostrinia nubilalis
B.279	CryIIIA	Pandemis spp.
B.280	CryIIIA	Pectinophora gossyp.
B.281	CryIIIA	Phyllocnistis citrella
B.282	CryIIIA	Pieris spp.
B.283	CryIIIA	Plutella xylostella
B.284	CryIIIA	Scirpophaga spp.
B.285	CryIIIA	Sesamia spp.
B.286	CryIIIA	Sparganothis spp.
B.287	CryIIIA	Spodoptera spp.
B.288	CryIIIA	Tortrix spp.
B.289	CryIIIA	Trichoplusia ni
B.290	CryIIIA	Agriotes spp.
B.291	CryIIIA	Anthonomus grandis
B.292	CryIIIA	Curculio spp.
B.293	CryIIIA	Diabrotica balteata
B.294	CryIIIA	Leptinotarsa spp.
B.295	CryIIIA	Lissorhoptrus spp.
B.296	CryIIIA	Otiorhynchus spp.
B.297	CryIIIA	Aleurothrixus spp.
B.298	CryIIIA	Aleyrodes spp.
B.299	CryIIIA	Aonidiella spp.
B.300	CryIIIA	Aphididae spp.
B.301	CryIIIA	Aphis spp.
B.302	CryIIIA	Bemisia tabaci
B.303	CryIIIA	Empoasca spp.
B.304	CryIIIA	Mycus spp.
B.305	CryIIIA	Nephotettix spp.
B.306	CryIIIA	Nilaparvata spp.
B.307	CryIIIA	Pseudococcus spp.
B.308	CryIIIA	Psylla spp.
B.309	CryIIIA	Quadraspidiotus spp.
B.310	CryIIIA	Schizaphis spp.
B.311	CryIIIA	Trialeurodes spp.
B.312	CryIIIA	Lyriomyza spp.
B.313	CryIIIA	Oscinella spp.
B.314	CryIIIA	Phorbia spp.
B.315	CryIIIA	Frankliniella spp.
B.316	CryIIIA	Thrips spp.
B.317	CryIIIA	Scirtothrips aurantii
B.318	CryIIIA	Aceria spp.
B.319	CryIIIA	Aculus spp.
B.320	CryIIIA	Brevipalpus spp.
B.321	CryIIIA	Panonychus spp.
B.322	CryIIIA	Phyllocoptruta spp.
B.323	CryIIIA	Tetranychus spp.
B.324	CryIIIA	Heterodera spp.

	AP	Control of
B.325	CryIIIA	Meloidogyne spp.
B.326	CryIIIB2	Adoxophyes spp.
B.327	CryIIIB2	Agrotis spp.
B.328	CryIIIB2	Alabama argillaceae
B.329	CryIIIB2	Anticarsia gemmatalis
B.330	CryIIIB2	Chilo spp.
B.331	CryIIIB2	Clysia ambiguella
B.332	CryIIIB2	Crocidolomia binotalis
B.333	CryIIIB2	Cydia spp.
B.334	CryIIIB2	Diparopsis castanea
B.335	CryIIIB2	Earias spp.
B.336	CryIIIB2	Ephestia spp.
B.337	CryIIIB2	Heliothis spp.
B.338	CryIIIB2	Hellula undalis
B.339	CryIIIB2	Keiferia lycopersicella
B.340	CryIIIB2	Leucoptera scitella
B.341	CryIIIB2	Lithocollethis spp.
B.342	CryIIIB2	Lobesia botrana
B.343	CryIIIB2	Ostrinia nubilalis
B.344	CryIIIB2	Pandemis spp.
B.345	CryIIIB2	Pectinophora gossyp.
B.346	CryIIIB2	Phyllocnistis citrella
B.347	CryIIIB2	Pieris spp.
B.348	CryIIIB2	Plutella xylostella
B.349	CryIIIB2	Scirpophaga spp.
B.350	CryIIIB2	Sesamia spp.
B.351	CryIIIB2	Sparganothis spp.
B.352	CryIIIB2	Spodoptera spp.
B.353	CryIIIB2	Tortrix spp.
B.354	CryIIIB2	Trichoplusia ni
B.355	CryIIIB2	Agriotes spp.
B.356	CryIIIB2	Anthonomus grandis
B.357	CryIIIB2	Curculio spp.
B.358	CryIIIB2	Diabrotica balteata
B.359	CryIIIB2	Leptinotarsa spp.
B.360	CryIIIB2	Lissorhoptrus spp.
B.361	CryIIIB2	Otiorhynchus spp.
B.362	CryIIIB2	Aleurothrixus spp.
B.363	CryIIIB2	Aleyrodes spp.
B.364	CryIIIB2	Aonidiella spp.
B.365	CryIIIB2	Aphididae spp.
B.366	CryIIIB2	Aphis spp.
B.367	CryIIIB2	Bemisia tabaci
B.368	CryIIIB2	Empoasca spp.
B.369	CryIIIB2	Mycus spp.
B.370	CryIIIB2	Nephotettix spp.
B.371	CryIIIB2	Nilaparvata spp.
B.372	CryIIIB2	Pseudococcus spp.
B.373	CryIIIB2	Psylla spp.
B.374	CryIIIB2	Quadraspidiotus spp.
B.375	CryIIIB2	Schizaphis spp.
B.376	CryIIIB2	Trialeurodes spp.
B.377	CryIIIB2	Lyriomyza spp.
B.378	CryIIIB2	Oscinella spp.
B.379	CryIIIB2	Phorbia spp.
B.380	CryIIIB2	Frankliniella spp.

	AP	Control of
B.381	CryIIIB2	Thrips spp.
B.382	CryIIIB2	Scirtothrips aurantii
B.383	CryIIIB2	Aceria spp.
B.384	CryIIIB2	Aculus spp.
B.385	CryIIIB2	Brevipalpus spp.
B.386	CryIIIB2	Panonychus spp.
B.387	CryIIIB2	Phyllocoptruta spp.
B.388	CryIIIB2	Tetranychus spp.
B.389	CryIIIB2	Heterodera spp.
B.390	CryIIIB2	Meloidogyne spp.
B.391	CytA	Adoxophyes spp.
B.392	CytA	Agrotis spp.
B.393	CytA	Alabama argillaceae
B.394	CytA	Anticarsia gemmatalis
B.395	CytA	Chilo spp.
B.396	CytA	Clysia ambiguella
B.397	CytA	Crocidolomia binotalis
B.398	CytA	Cydia spp.
B.399	CytA	Diparopsis castanea
B.400	CytA	Earias spp.
B.401	CytA	Ephestia spp.
B.402	CytA	Heliothis spp.
B.403	CytA	Hellula undalis
B.404	CytA	Keiferia lycopersicella
B.405	CytA	Leucoptera scitella
B.406	CytA	Lithocollethis spp.
B.407	CytA	Lobesia botrana
B.408	CytA	Ostrinia nubilalis
B.409	CytA	Pandemis spp.
B.410	CytA	Pectinophora gossyp.
B.411	CytA	Phyllocnistis citrella
B.412	CytA	Pieris spp.
B.413	CytA	Plutella xylostella
B.414	CytA	Scirpophaga spp.
B.415	CytA	Sesamia spp.
B.416	CytA	Sparganothis spp.
B.417	CytA	Spodoptera spp.
B.418	CytA	Tortrix spp.
B.419	CytA	Trichoplusia ni
B.420	CytA	Agriotes spp.
B.421	CytA	Anthonomus grandis
B.422	CytA	Curculio spp.
B.423	CytA	Diabrotica balteata
B.424	CytA	Leptinotarsa spp.
B.425	CytA	Lissorhoptrus spp.
B.426	CytA	Otiorhynchus spp.
B.427	CytA	Aleurothrixus spp.
B.428	CytA	Aleyrodes spp.
B.429	CytA	Aonidiella spp.
B.430	CytA	Aphididae spp.
B.431	CytA	Aphis spp.
B.432	CytA	Bemisia tabaci
B.433	CytA	Empoasca spp.
B.434	CytA	Mycus spp.
B.435	CytA	Nephotettix spp.
B.436	CytA	Nilaparvata spp.
B.437	CytA	Pseudococcus spp.

	AP	Control of
B.438	CytA	Psylla spp.
B.439	CytA	Quadraspidiotus spp.
B.440	CytA	Schizaphis spp.
B.441	CytA	Trialeurodes spp.
B.442	CytA	Lyriomyza spp.
B.443	CytA	Oscinella spp.
B.444	CytA	Phorbia spp.
B.445	CytA	Frankliniella spp.
B.446	CytA	Thrips spp.
B.447	CytA	Scirtothrips aurantii
B.448	CytA	Aceria spp.
B.449	CytA	Aculus spp.
B.450	CytA	Brevipalpus spp.
B.451	CytA	Panonychus spp.
B.452	CytA	Phyllocoptruta spp.
B.453	CytA	Tetranychus spp.
B.454	CytA	Heterodera spp.
B.455	CytA	Meloidogyne spp.
B.456	VIP3	Adoxophyes spp.
B.457	VIP3	Agrotis spp.
B.458	VIP3	Alabama argillaceae
B.459	VIP3	Anticarsia gemmatalis
B.460	VIP3	Chilo spp.
B.461	VIP3	Clysia ambiguella
B.462	VIP3	Crocidolomia binotalis
B.463	VIP3	Cydia spp.
B.464	VIP3	Diparopsis castanea
B.465	VIP3	Earias spp.
B.466	VIP3	Ephestia spp.
B.467	VIP3	Heliothis spp.
B.468	VIP3	Hellula undalis
B.469	VIP3	Keiferia lycopersicella
B.470	VIP3	Leucoptera scitella
B.471	VIP3	Lithocollethis spp.
B.472	VIP3	Lobesia botrana
B.473	VIP3	Ostrinia nubilalis
B.474	VIP3	Pandemis spp.
B.475	VIP3	Pectinophora gossyp.
B.476	VIP3	Phyllocnistis citrella
B.477	VIP3	Pieris spp.
B.478	VIP3	Plutella xylostella
B.479	VIP3	Scirpophaga spp.
B.480	VIP3	Sesamia spp.
B.481	VIP3	Sparganothis spp.
B.482	VIP3	Spodoptera spp.
B.483	VIP3	Tortrix spp.
B.484	VIP3	Trichoplusia ni
B.485	VIP3	Agriotes spp.
B.486	VIP3	Anthonomus grandis
B.487	VIP3	Curculio spp.
B.488	VIP3	Diabrotica balteata
B.489	VIP3	Leptinotarsa spp.
B.490	VIP3	Lissorhoptrus spp.
B.491	VIP3	Otiorhynchus spp.
B.492	VIP3	Aleurothrixus spp.
B.493	VIP3	Aleyrodes spp.

	AP	Control of
B.494	VIP3	Aonidiella spp.
B.495	VIP3	Aphididae spp.
B.496	VIP3	Aphis spp.
B.497	VIP3	Bemisia tabaci
B.498	VIP3	Empoasca spp.
B.499	VIP3	Mycus spp.
B.500	VIP3	Nephotettix spp.
B.501	VIP3	Nilaparvata spp.
B.502	VIP3	Pseudococcus spp.
B.503	VIP3	Psylla spp.
B.504	VIP3	Quadraspidiotus spp.
B.505	VIP3	Schizaphis spp.
B.506	VIP3	Trialeurodes spp.
B.507	VIP3	Lyriomyza spp.
B.508	VIP3	Oscinella spp.
B.509	VIP3	Phorbia spp.
B.510	VIP3	Frankliniella spp.
B.511	VIP3	Thrips spp.
B.512	VIP3	Scirtothrips aurantii
B.513	VIP3	Aceria spp.
B.514	VIP3	Aculus spp.
B.515	VIP3	Brevipalpus spp.
B.516	VIP3	Panonychus spp.
B.517	VIP3	Phyllocoptruta spp.
B.518	VIP3	Tetranychus spp.
B.519	VIP3	Heterodera spp.
B.520	VIP3	Meloidogyne spp.
B.521	GL	Adoxophyes spp.
B.522	GL	Agrotis spp.
B.523	GL	Alabama argillaceae
B.524	GL	Anticarsia gemmatalis
B.525	GL	Chilo spp.
B.526	GL	Clysia ambiguella
B.527	GL	Crocidolomia binotalis
B.528	GL	Cydia spp.
B.529	GL	Diparopsis castanea
B.530	GL	Earias spp.
B.531	GL	Ephestia spp.
B.532	GL	Heliothis spp.
B.533	GL	Hellula undalis
B.534	GL	Keiferia lycopersicella
B.535	GL	Leucoptera scitella
B.536	GL	Lithocollethis spp.
B.537	GL	Lobesia botrana
B.538	GL	Ostrinia nubilalis
B.539	GL	Pandemis spp.
B.540	GL	Pectinophora gossyp.
B.541	GL	Phyllocnistis citrella
B.542	GL	Pieris spp.
B.543	GL	Plutella xylostella
B.544	GL	Scirpophaga spp.
B.545	GL	Sesamia spp.
B.546	GL	Sparganothis spp.
B.547	GL	Spodoptera spp.
B.548	GL	Tortrix spp.
B.549	GL	Trichoplusia ni
B.550	GL	Agriotes spp.

	AP	Control of
B.551	GL	Anthonomus grandis
B.552	GL	Curculio spp.
B.553	GL	Diabrotica balteata
B.554	GL	Leptinotarsa spp.
B.555	GL	Lissorhoptrus spp.
B.556	GL	Otiorhynchus spp.
B.557	GL	Aleurothrixus spp.
B.558	GL	Aleyrodes spp.
B.559	GL	Aonidiella spp.
B.560	GL	Aphididae spp.
B.561	GL	Aphis spp.
B.562	GL	Bemisia tabaci
B.563	GL	Empoasca spp.
B.564	GL	Mycus spp.
B.565	GL	Nephotettix spp.
B.566	GL	Nilaparvata spp.
B.567	GL	Pseudococcus spp.
B.568	GL	Psylla spp.
B.569	GL	Quadraspidiotus spp.
B.570	GL	Schizaphis spp.
B.571	GL	Trialeurodes spp.
B.572	GL	Lyriomyza spp.
B.573	GL	Oscinella spp.
B.574	GL	Phorbia spp.
B.575	GL	Frankliniella spp.
B.576	GL	Thrips spp.
B.577	GL	Scirtothrips aurantii
B.578	GL	Aceria spp.
B.579	GL	Aculus spp.
B.580	GL	Brevipalpus spp.
B.581	GL	Panonychus spp.
B.582	GL	Phyllocoptruta spp.
B.583	GL	Tetranychus spp.
B.584	GL	Heterodera spp.
B.585	GL	Meloidogyne spp.
B.586	PL	Adoxophyes spp.
B.587	PL	Agrotis spp.
B.588	PL	Alabama argillaceae
B.589	PL	Anticarsia gemmatalis
B.590	PL	Chilo spp.
B.591	PL	Clysia ambiguella
B.592	PL	Crocidolomia binotalis
B.593	PL	Cydia spp.
B.594	PL	Diparopsis castanea
B.595	PL	Earias spp.
B.596	PL	Ephestia spp.
B.597	PL	Heliothis spp.
B.598	PL	Hellula undalis
B.599	PL	Keiferia lycopersicella
B.600	PL	Leucoptera scitella
B.601	PL	Lithocollethis spp.
B.602	PL	Lobesia botrana
B.603	PL	Ostrinia nubilalis
B.604	PL	Pandemis spp.
B.605	PL	Pectinophora gossyp.
B.606	PL	Phyllocnistis citrella

	AP	Control of
B.607	PL	Pieris spp.
B.608	PL	Plutella xylostella
B.609	PL	Scirpophaga spp.
B.610	PL	Sesamia spp.
B.611	PL	Sparganothis spp.
B.612	PL	Spodoptera spp.
B.613	PL	Tortrix spp.
B.614	PL	Trichoplusia ni
B.615	PL	Agriotes spp.
B.616	PL	Anthonomus grandis
B.617	PL	Curculio spp.
B.618	PL	Diabrotica balteata
B.619	PL	Leptinotarsa spp.
B.620	PL	Lissorhoptrus spp.
B.621	PL	Otiorhynchus spp.
B.622	PL	Aleurothrixus spp.
B.623	PL	Aleyrodes spp.
B.624	PL	Aonidiella spp.
B.625	PL	Aphididae spp.
B.626	PL	Aphis spp.
B.627	PL	Bemisia tabaci
B.628	PL	Empoasca spp.
B.629	PL	Mycus spp.
B.630	PL	Nephotettix spp.
B.631	PL	Nilaparvata spp.
B.632	PL	Pseudococcus spp.
B.633	PL	Psylla spp.
B.634	PL	Quadraspidiotus spp.
B.635	PL	Schizaphis spp.
B.636	PL	Trialeurodes spp.
B.637	PL	Lyriomyza spp.
B.638	PL	Oscinella spp.
B.639	PL	Phorbia spp.
B.640	PL	Frankliniella spp.
B.641	PL	Thrips spp.
B.642	PL	Scirtothrips aurantii
B.643	PL	Aceria spp.
B.644	PL	Aculus spp.
B.645	PL	Brevipalpus spp.
B.646	PL	Panonychus spp.
B.647	PL	Phyllocoptruta spp.
B.648	PL	Tetranychus spp.
B.649	PL	Heterodera spp.
B.650	PL	Meloidogyne spp.
B.651	XN	Adoxophyes spp.
B.652	XN	Agrotis spp.
B.653	XN	Alabama argillaceae
B.654	XN	Anticarsia gemmatalis
B.655	XN	Chilo spp.
B.656	XN	Clysia ambiguella
B.657	XN	Crocidolomia binotalis
B.658	XN	Cydia spp.
B.659	XN	Diparopsis castanea
B.660	XN	Earias spp.
B.661	XN	Ephestia spp.
B.662	XN	Heliothis spp.
B.663	XN	Hellula undalis
B.664	XN	Keiferia

	AP	Control of
		lycopersicella
B.665	XN	Leucoptera scitella
B.666	XN	Lithocollethis spp.
B.667	XN	Lobesia botrana
B.668	XN	Ostrinia nubilalis
B.669	XN	Pandemis spp.
B.670	XN	Pectinophora gossyp.
B.671	XN	Phyllocnistis citrella
B.672	XN	Pieris spp.
B.673	XN	Plutella xylostella
B.674	XN	Scirpophaga spp.
B.675	XN	Sesamia spp.
B.676	XN	Sparganothis spp.
B.677	XN	Spodoptera spp.
B.678	XN	Tortrix spp.
B.679	XN	Trichoplusia ni
B.680	XN	Agriotes spp.
B.681	XN	Anthonomus grandis
B.682	XN	Curculio spp.
B.683	XN	Diabrotica balteata
B.684	XN	Leptinotarsa spp.
B.685	XN	Lissorhoptrus spp.
B.686	XN	Otiorhynchus spp.
B.687	XN	Aleurothrixus spp.
B.688	XN	Aleyrodes spp.
B.689	XN	Aonidiella spp.
B.690	XN	Aphididae spp.
B.691	XN	Aphis spp.
B.692	XN	Bemisia tabaci
B.693	XN	Empoasca spp.
B.694	XN	Mycus spp.
B.695	XN	Nephotettix spp.
B.696	XN	Nilaparvata spp.
B.697	XN	Pseudococcus spp.
B.698	XN	Psylla spp.
B.699	XN	Quadraspidiotus spp.
B.700	XN	Schizaphis spp.
B.701	XN	Trialeurodes spp.
B.702	XN	Lyriomyza spp.
B.703	XN	Oscinella spp.
B.704	XN	Phorbia spp.
B.705	XN	Frankliniella spp.
B.706	XN	Thrips spp.
B.707	XN	Scirtothrips aurantii
B.708	XN	Aceria spp.
B.709	XN	Aculus spp.
B.710	XN	Brevipalpus spp.
B.711	XN	Panonychus spp.
B.712	XN	Phyllocoptruta spp.
B.713	XN	Tetranychus spp.
B.714	XN	Heterodera spp.
B.715	XN	Meloidogyne spp.
B.716	PInh.	Adoxophyes spp.
B.717	PInh.	Agrotis spp.
B.718	PInh.	Alabama argillaceae
B.719	PInh.	Anticarsia gemmatalis
B.720	PInh.	Chilo spp.
B.721	PInh.	Clysia ambiguella
B.722	PInh.	Crocidolomia

	AP	Control of
		binotalis
B.723	PInh.	Cydia spp.
B.724	PInh.	Diparopsis castanea
B.725	PInh.	Earias spp.
B.726	PInh.	Ephestia spp.
B.727	PInh.	Heliothis spp.
B.728	PInh.	Hellula undalis
B.729	PInh.	Keiferia lycopersicella
B.730	PInh.	Leucoptera scitella
B.731	PInh.	Lithocollethis spp.
B.732	PInh.	Lobesia botrana
B.733	PInh.	Ostrinia nubilalis
B.734	PInh.	Pandemis spp.
B.735	PInh.	Pectinophora gossyp.
B.736	PInh.	Phyllocnistis citrella
B.737	PInh.	Pieris spp.
B.738	PInh.	Plutella xylostella
B.739	PInh.	Scirpophaga spp.
B.740	PInh.	Sesamia spp.
B.741	PInh.	Sparganothis spp.
B.742	PInh.	Spodoptera spp.
B.743	PInh.	Tortrix spp.
B.744	PInh.	Trichoplusia ni
B.745	PInh.	Agriotes spp.
B.746	PInh.	Anthonomus grandis
B.747	PInh.	Curculio spp.
B.748	PInh.	Diabrotica balteata
B.749	PInh.	Leptinotarsa spp.
B.750	PInh.	Lissorhoptrus spp.
B.751	PInh.	Otiorhynchus spp.
B.752	PInh.	Aleurothrixus spp.
B.753	PInh.	Aleyrodes spp.
B.754	PInh.	Aonidiella spp.
B.755	PInh.	Aphididae spp.
B.756	PInh.	Aphis spp.
B.757	PInh.	Bemisia tabaci
B.758	PInh.	Empoasca spp.
B.759	PInh.	Mycus spp.
B.760	PInh.	Nephotettix spp.
B.761	PInh.	Nilaparvata spp.
B.762	PInh.	Pseudococcus spp.
B.763	PInh.	Psylla spp.
B.764	PInh.	Quadraspidiotus spp.
B.765	PInh.	Schizaphis spp.
B.766	PInh.	Trialeurodes spp.
B.767	PInh.	Lyriomyza spp.
B.768	PInh.	Oscinella spp.
B.769	PInh.	Phorbia spp.
B.770	PInh.	Frankliniella spp.
B.771	PInh.	Thrips spp.
B.772	PInh.	Scirtothrips aurantii
B.773	PInh.	Aceria spp.
B.774	PInh.	Aculus spp.
B.775	PInh.	Brevipalpus spp.
B.776	PInh.	Panonychus spp.
B.777	PInh.	Phyllocoptruta spp.
B.778	PInh.	Tetranychus spp.
B.779	PInh.	Heterodera spp.
B.780	PInh.	Meloidogyne spp.

	AP	Control of
B.781	PLec.	Adoxophyes spp.
B.782	PLec.	Agrotis spp.
B.783	PLec.	Alabama argillaceae
B.784	PLec.	Anticarsia gemmatalis
B.785	PLec.	Chilo spp.
B.786	PLec.	Clysia ambiguella
B.787	PLec.	Crocidolomia binotalis
B.788	PLec.	Cydia spp.
B.789	PLec.	Diparopsis castanea
B.790	PLec.	Earias spp.
B.791	PLec.	Ephestia spp.
B.792	PLec.	Heliothis spp.
B.793	PLec.	Hellula undalis
B.794	PLec.	Keiferia lycopersicella
B.795	PLec.	Leucoptera scitella
B.796	PLec.	Lithocollethis spp.
B.797	PLec.	Lobesia botrana
B.798	PLec.	Ostrinia nubilalis
B.799	PLec.	Pandemis spp.
B.800	PLec.	Pectinophora gossyp.
B.801	PLec.	Phyllocnistis citrella
B.802	PLec.	Pieris spp.
B.803	PLec.	Plutella xylostella
B.804	PLec.	Scirpophaga spp.
B.805	PLec.	Sesamia spp.
B.806	PLec.	Sparganothis spp.
B.807	PLec.	Spodoptera spp.
B.808	PLec.	Tortrix spp.
B.809	PLec.	Trichoplusia ni
B.810	PLec.	Agriotes spp.
B.811	PLec.	Anthonomus grandis
B.812	PLec.	Curculio spp.
B.813	PLec.	Diabrotica balteata
B.814	PLec.	Leptinotarsa spp.
B.815	PLec.	Lissorhoptrus spp.
B.816	PLec.	Otiorhynchus spp.
B.817	PLec.	Aleurothrixus spp.
B.818	PLec.	Aleyrodes spp.
B.819	PLec.	Aonidiella spp.
B.820	PLec.	Aphididae spp.
B.821	PLec.	Aphis spp.
B.822	PLec.	Bemisia tabaci
B.823	PLec.	Empoasca spp.
B.824	PLec.	Mycus spp.
B.825	PLec.	Nephotettix spp.
B.826	PLec.	Nilaparvata spp.
B.827	PLec.	Pseudococcus spp.
B.828	PLec.	Psylla spp.
B.829	PLec.	Quadraspidiotus spp.
B.830	PLec.	Schizaphis spp.
B.831	PLec.	Trialeurodes spp.
B.832	PLec.	Lyriomyza spp.
B.833	PLec.	Oscinella spp.
B.834	PLec.	Phorbia spp.
B.835	PLec.	Frankliniella spp.
B.836	PLec.	Thrips spp.

	AP	Control of
B.837	PLec.	Scirtothrips aurantii
B.838	PLec.	Aceria spp.
B.839	PLec.	Aculus spp.
B.840	PLec.	Brevipalpus spp.
B.841	PLec.	Panonychus spp.
B.842	PLec.	Phyllocoptruta spp.
B.843	PLec.	Tetranychus spp.
B.844	PLec.	Heterodera spp.
B.845	PLec.	Meloidogyne spp.
B.846	Aggl.	Adoxophyes spp.
B.847	Aggl.	Agrotis spp.
B.848	Aggl.	Alabama argillaceae
B.849	Aggl.	Anticarsia gemmatalis
B.850	Aggl.	Chilo spp.
B.851	Aggl.	Clysia ambiguella
B.852	Aggl.	Crocidolomia binotalis
B.853	Aggl.	Cydia spp.
B.854	Aggl.	Diparopsis castanea
B.855	Aggl.	Earias spp.
B.856	Aggl.	Ephestia spp.
B.857	Aggl.	Heliothis spp.
B.858	Aggl.	Hellula undalis
B.859	Aggl.	Keiferia lycopersicella
B.860	Aggl.	Leucoptera scitella
B.861	Aggl.	Lithocollethis spp.
B.862	Aggl.	Lobesia botrana
B.863	Aggl.	Ostrinia nubilalis
B.864	Aggl.	Pandemis spp.
B.865	Aggl.	Pectinophora gossyp.
B.866	Aggl.	Phyllocnistis citrella
B.867	Aggl.	Pieris spp.
B.868	Aggl.	Plutella xylostella
B.869	Aggl.	Scirpophaga spp.
B.870	Aggl.	Sesamia spp.
B.871	Aggl.	Sparganothis spp.
B.872	Aggl.	Spodoptera spp.
B.873	Aggl.	Tortrix spp.
B.874	Aggl.	Trichoplusia ni
B.875	Aggl.	Agriotes spp.
B.876	Aggl.	Anthonomus grandis
B.877	Aggl.	Curculio spp.
B.878	Aggl.	Diabrotica balteata
B.879	Aggl.	Leptinotarsa spp.
B.880	Aggl.	Lissorhoptrus spp.
B.881	Aggl.	Otiorhynchus spp.
B.882	Aggl.	Aleurothrixus spp.
B.883	Aggl.	Aleyrodes spp.
B.884	Aggl.	Aonidiella spp.
B.885	Aggl.	Aphididae spp.
B.886	Aggl.	Aphis spp.
B.887	Aggl.	Bemisia tabaci
B.888	Aggl.	Empoasca spp.
B.889	Aggl.	Mycus spp.
B.890	Aggl.	Nephotettix spp.
B.891	Aggl.	Nilaparvata spp.
B.892	Aggl.	Pseudococcus spp.
B.893	Aggl.	Psylla spp.

	AP	Control of
B.894	Aggl.	Quadraspidiotus spp.
B.895	Aggl.	Schizaphis spp.
B.896	Aggl.	Trialeurodes spp.
B.897	Aggl.	Lyriomyza spp.
B.898	Aggl.	Oscinella spp.
B.899	Aggl.	Phorbia spp.
B.900	Aggl.	Frankliniella spp.
B.901	Aggl.	Thrips spp.
B.902	Aggl.	Scirtothrips aurantii
B.903	Aggl.	Aceria spp.
B.904	Aggl.	Aculus spp.
B.905	Aggl.	Brevipalpus spp.
B.906	Aggl.	Panonychus spp.
B.907	Aggl.	Phyllocoptruta spp.
B.908	Aggl.	Tetranychus spp.
B.909	Aggl.	Heterodera spp.
B.910	Aggl.	Meloidogyne spp.
B.911	CO	Adoxophyes spp.
B.912	CO	Agrotis spp.
B.913	CO	Alabama argillaceae
B.914	CO	Anticarsia gemmatalis
B.915	CO	Chilo spp.
B.916	CO	Clysia ambiguella
B.917	CO	Crocidolomia binotalis
B.918	CO	Cydia spp.
B.919	CO	Diparopsis castanea
B.920	CO	Earias spp.
B.921	CO	Ephestia spp.
B.922	CO	Heliothis spp.
B.923	CO	Hellula undalis
B.924	CO	Keiferia lycopersicella
B.925	CO	Leucoptera scitella
B.926	CO	Lithocollethis spp.
B.927	CO	Lobesia botrana
B.928	CO	Ostrinia nubilalis
B.929	CO	Pandemis spp.
B.930	CO	Pectinophora gossyp.
B.931	CO	Phyllocnistis citrella
B.932	CO	Pieris spp.
B.933	CO	Plutella xylostella
B.934	CO	Scirpophaga spp.
B.935	CO	Sesamia spp.
B.936	CO	Sparganothis spp.
B.937	CO	Spodoptera spp.
B.938	CO	Tortrix spp.
B.939	CO	Trichoplusia ni
B.940	CO	Agriotes spp.
B.941	CO	Anthonomus grandis
B.942	CO	Curculio spp.
B.943	CO	Diabrotica balteata
B.944	CO	Leptinotarsa spp.
B.945	CO	Lissorhoptrus spp.
B.946	CO	Otiorhynchus spp.
B.947	CO	Aleurothrixus spp.
B.948	CO	Aleyrodes spp.
B.949	CO	Aonidiella spp.

	AP	Control of
B.950	CO	Aphididae spp.
B.951	CO	Aphis spp.
B.952	CO	Bemisia tabaci
B.953	CO	Empoasca spp.
B.954	CO	Mycus spp.
B.955	CO	Nephotettix spp.
B.956	CO	Nilaparvata spp.
B.957	CO	Pseudococcus spp.
B.958	CO	Psylla spp.
B.959	CO	Quadraspidiotus spp.
B.960	CO	Schizaphis spp.
B.961	CO	Trialeurodes spp.
B.962	CO	Lyriomyza spp.
B.963	CO	Oscinella spp.
B.964	CO	Phorbia spp.
B.965	CO	Frankliniella spp.
B.966	CO	Thrips spp.
B.967	CO	Scirtothrips aurantii
B.968	CO	Aceria spp.
B.969	CO	Aculus spp.
B.970	CO	Brevipalpus spp.
B.971	CO	Panonychus spp.
B.972	CO	Phyllocoptruta spp.
B.973	CO	Tetranychus spp.
B.974	CO	Heterodera spp.
B.975	CO	Meloidogyne spp.
B.976	CH	Adoxophyes spp.
B.977	CH	Agrotis spp.
B.978	CH	Alabama argillaceae
B.979	CH	Anticarsia gemmatalis
B.980	CH	Chilo spp.
B.981	CH	Clysia ambiguella
B.982	CH	Crocidolomia binotalis
B.983	CH	Cydia spp.
B.984	CH	Diparopsis castanea
B.985	CH	Earias spp.
B.986	CH	Ephestia spp.
B.987	CH	Heliothis spp.
B.988	CH	Hellula undalis
B.989	CH	Keiferia lycopersicella
B.990	CH	Leucoptera scitella
B.991	CH	Lithocollethis spp.
B.992	CH	Lobesia botrana
B.993	CH	Ostrinia nubilalis
B.994	CH	Pandemis spp.
B.995	CH	Pectinophora gossyp.
B.996	CH	Phyllocnistis citrella
B.997	CH	Pieris spp.
B.998	CH	Plutella xylostella
B.999	CH	Scirpophaga spp.
B.1000	CH	Sesamia spp.
B.1001	CH	Sparganothis spp.
B.1002	CH	Spodoptera spp.
B.1003	CH	Tortrix spp.
B.1004	CH	Trichoplusia ni
B.1005	CH	Agriotes spp.
B.1006	CH	Anthonomus

	AP	Control of
		grandis
B.1007	CH	Curculio spp.
B.1008	CH	Diabrotica balteata
B.1009	CH	Leptinotarsa spp.
B.1010	CH	Lissorhoptrus spp.
B.1011	CH	Otiorhynohus spp.
B.1012	CH	Aleurothrixus spp.
B.1013	CH	Aleyrodes spp.
B.1014	CH	Aonidiella spp.
B.1015	CH	Aphididae spp.
B.1016	CH	Aphis spp.
B.1017	CH	Bemisia tabaci
B.1018	CH	Empoasca spp.
B.1019	CH	Mycus spp.
B.1020	CH	Nephotettix spp.
B.1021	CH	Nilaparvata spp.
B.1022	CH	Pseudococcus spp.
B.1023	CH	Psylla spp.
B.1024	CH	Quadraspidiotus
		spp.
B.1025	CH	Schizaphis spp.
B.1026	CH	Trialeurodes spp.
B.1027	CH	Lyriomyza spp.
B.1028	CH	Oscinella spp.
B.1029	CH	Phorbia spp.
B.1030	CH	Frankliniella spp.
B.1031	CH	Thrips spp.
B.1032	CH	Scirtothrips aurantii
B.1033	CH	Aceria spp.
B.1034	CH	Aculus spp.
B.1035	CH	Brevipalpus spp.
B.1036	CH	Panonychus spp.

	AP	Control of
B.1037	CH	Phyllocoptruta spp.
B.1038	CH	Tetranychus spp.
B.1039	CH	Heterodera spp.
B.1040	CH	Meloidogyne spp.
B.1041	SS	Adoxophyes spp.
B.1042	SS	Agrotis spp.
B.1043	SS	Alabama
		argillaceae
B.1044	SS	Anticarsia
		gemmatalis
B.1045	SS	Chilo spp.
B.1046	SS	Clysia ambiguella
B.1047	SS	Crocidolomia
		binotalis
B.1048	SS	Cydia spp.
B.1049	SS	Diparopsis
		castanea
B.1050	SS	Earias spp.
B.1051	SS	Ephestia spp.
B.1052	SS	Heliothis spp.
B.1053	SS	Hellula undalis
B.1054	SS	Keiferia
		lycopersicella
B.1055	SS	Leucoptera scitella
B.1056	SS	Lithocollethis spp.
B.1057	SS	Lobesia botrana
B.1058	SS	Ostrinia nubilalis
B.1059	SS	Pandemis spp.
B.1060	SS	Pectinophora
		gossyp.
B.1061	SS	Phyllocnistis citrella
B.1062	SS	Pieris spp.

	AP	Control of
B.1063	SS	Plutella xylostella
B.1064	SS	Scirpophaga spp.
B.1065	SS	Sesamia spp.
B.1066	SS	Sparganothis spp.
B.1067	SS	Spodoptera spp.
B.1068	SS	Tortrix spp.
B.1069	SS	Trichoplusia ni
B.1070	SS	Agriotes spp.
B.1071	SS	Anthonomus grandis
B.1072	SS	Curculio spp.
B.1073	SS	Diabrotica balteata
B.1074	SS	Leptinotarsa spp.
B.1075	SS	Lissorhoptrus spp.
B.1076	SS	Otiorhynchus spp.
B.1077	SS	Aleurothrixus spp.
B.1078	SS	Aleyrodes spp.
B.1079	SS	Aonidiella spp.
B.1080	SS	Aphididae spp.
B.1081	SS	Aphis spp.
B.1082	SS	Bemisia tabaci
B.1083	SS	Empoasca spp.
B.1084	SS	Mycus spp.
B.1085	SS	Nephotettix spp.
B.1086	SS	Nilaparvata spp.
B.1087	SS	Pseudococcus spp.
B.1088	SS	Psylla spp.
B.1089	SS	Quadraspidiotus spp.
B.1090	SS	Schizaphis spp.
B.1091	SS	Trialeurodes spp.
B.1092	SS	Lyriomyza spp.
B.1093	SS	Oscinella spp.
B.1094	SS	Phorbia spp.
B.1095	SS	Frankliniella spp.
B.1096	SS	Thrips spp.
B.1097	SS	Scirtothrips aurantii
B.1098	SS	Aceria spp.
B.1099	SS	Aculus spp.
B.1100	SS	Brevipalpus spp.
B.1101	SS	Panonychus spp.
B.1102	SS	Phyllocoptruta spp.
B.1103	SS	Tetranychus spp.
B.1104	SS	Heterodera spp.
B.1105	SS	Meloidogyne spp.
B.1106	HO	Adoxophyes spp.
B.1107	HO	Agrotis spp.
B.1108	HO	Alabama argillaceae
B.1109	HO	Anticarsia gemmatalis
B.1110	HO	Chilo spp.
B.1111	HO	Clysia ambiguella
B.1112	HO	Crocidolomia binotalis
B.1113	HO	Cydia spp.
B.1114	HO	Diparopsis castanea
B.1115	HO	Earias spp.
B.1116	HO	Ephestia spp.
B.1117	HO	Heliothis spp.
B.1118	HO	Hellula undalis
B.1119	HO	Keiferia lycopersicella

	AP	Control of
B.1120	HO	Leucoptera scitella
B.1121	HO	Lithocollethis spp.
B.1122	HO	Lobesia botrana
B.1123	HO	Ostrinia nubilalis
B.1124	HO	Pandemis spp.
B.1125	HO	Pectinophora gossypiella
B.1126	HO	Phyllocnistis citrella
B.1127	HO	Pieris spp.
B.1128	HO	Plutella xylostella
B.1129	HO	Scirpophaga spp.
B.1130	HO	Sesamia spp.
B.1131	HO	Sparganothis spp.
B.1132	HO	Spodoptera spp.
B.1133	HO	Tortrix spp.
B.1134	HO	Trichoplusia ni
B.1135	HO	Agriotes spp.
B.1136	HO	Anthonomus grandis
B.1137	HO	Curculio spp.
B.1138	HO	Diabrotica balteata
B.1139	HO	Leptinotarsa spp.
B.1140	HO	Lissorhoptrus spp.
B.1141	HO	Otiorhynchus spp.
B.1142	HO	Aleurothrixus spp.
B.1143	HO	Aleyrodes spp.
B.1144	HO	Aonidiella spp.
B.1145	HO	Aphididae spp.
B.1146	HO	Aphis spp.
B.1147	HO	Bemisia tabaci
B.1148	HO	Empoasca spp.
B.1149	HO	Mycus spp.
B.1150	HO	Nephotettix spp.
B.1151	HO	Nilaparvata spp.
B.1152	HO	Pseudococcus spp.
B.1153	HO	Psylla spp.
B.1154	HO	Quadraspidiotus spp.
B.1155	HO	Schizaphis spp.
B.1156	HO	Trialeurodes spp.
B.1157	HO	Lyriomyza spp.
B.1158	HO	Oscinella spp.
B.1159	HO	Phorbia spp.
B.1160	HO	Frankliniella spp.
B.1161	HO	Thrips spp.
B.1162	HO	Scirtothrips aurantii
B.1163	HO	Aceria spp.
B.1164	HO	Aculus spp.
B.1165	HO	Brevipalpus spp.
B.1166	HO	Panonychus spp.
B.1167	HO	Phyllocoptruta spp.
B.1168	HO	Tetranychus spp.
B.1169	HO	Heterodera spp.
B.1170	HO	Meloidogyne spp.

Biological Examples

Table 1: A method of controlling pests comprising the application of thiamethoxam to transgenic cotton, wherein the combination of the active principle expressed by the

transgenic plant and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B.

Table 2: A method of controlling pests comprising the application of thiamethoxam to transgenic rice, wherein the combination of the active principle expressed by the transgenic plant and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B.

Table 3: A method of controlling pests comprising the application of thiamethoxam to transgenic potatoes, wherein the combination of the active principle expressed by the transgenic plant and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B.

Table 4: A method of controlling pests comprising the application of thiamethoxam to transgenic brassica, wherein the combination of the active principle expressed by the transgenic plant and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B.

Table 5: A method of controlling pests comprising the application of thiamethoxam to transgenic tomatoes, wherein the combination of the active principle expressed by the transgenic plant and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B.

Table 6: A method of controlling pests comprising the application of thiamethoxam to transgenic cucurbits, wherein the combination of the active principle expressed by the transgenic plant and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B.

Table 7: A method of controlling pests comprising the application of thiamethoxam to transgenic soybeans, wherein the combination of the active principle expressed by the transgenic plant and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B.

Table 8: A method of controlling pests comprising the application of thiamethoxam to transgenic maize, wherein the combination of the active principle expressed by the transgenic plant and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B.

Table 9: A method of controlling pests comprising the application of thiamethoxam to transgenic wheat, wherein the combination of the active principle expressed by the transgenic plant and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B.

Table 10: A method of controlling pests comprising the application of thiamethoxam to transgenic bananas, wherein the combination of the active principle expressed by the transgenic plant and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B.

Table 11: A method of controlling pests comprising the application of thiamethoxam to transgenic citrus trees, wherein the combination of the active principle expressed by the transgenic plant and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B.

Table 12: A method of controlling pests comprising the application of thiamethoxam to transgenic pome fruit trees, wherein the combination of the active principle expressed by the transgenic plant and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B.

Table 13: A method of controlling pests comprising the application of thiamethoxam to transgenic peppers, wherein the combination of the active principle expressed by the transgenic plant and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B.

Table 14: A method of controlling pests comprising the application of imidacloprid to transgenic cotton, wherein the combination of the active principle expressed by the transgenic plant and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B.

Table 15: A method of controlling pests comprising the application of imidacloprid to transgenic rice, wherein the combination of the active principle expressed by the transgenic plant and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B.

Table 16: A method of controlling pests comprising the application of imidacloprid to transgenic potatoes, wherein the combination of the active principle expressed by the

transgenic plant and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B.

Table 17: A method of controlling pests comprising the application of imidacloprid to transgenic tomatoes, wherein the combination of the active principle expressed by the transgenic plant and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B.

Table 18: A method of controlling pests comprising the application of imidacloprid to transgenic cucurbits, wherein the combination of the active principle expressed by the transgenic plant and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B.

Table 19: A method of controlling pests comprising the application of imidacloprid to transgenic soybeans, wherein the combination of the active principle expressed by the transgenic plant and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B.

Table 20: A method of controlling pests comprising the application of imidacloprid to transgenic maize, wherein the combination of the active principle expressed by the transgenic plant and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B.

Table 21: A method of controlling pests comprising the application of imidacloprid to transgenic wheat, wherein the combination of the active principle expressed by the transgenic plant and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B.

Table 22: A method of controlling pests comprising the application of imidacloprid to transgenic bananas, wherein the combination of the active principle expressed by the transgenic plant and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B.

Table 23: A method of controlling pests comprising the application of imidacloprid to transgenic orange trees, wherein the combination of the active principle expressed by the transgenic plant and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B.

Table 24: A method of controlling pests comprising the application of imidacloprid to transgenic pome fruit, wherein the combination of the active principle expressed by the transgenic plant and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B.

Table 25: A method of controlling pests comprising the application of imidacloprid to transgenic cucurbits, wherein the combination of the active principle expressed by the transgenic plant and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B.

Table 26: A method of controlling pests comprising the application of imidacloprid to transgenic peppers, wherein the combination of the active principle expressed by the transgenic plant and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B.

Table 27: A method of controlling pests comprising the application of Ti-435 to transgenic cotton, wherein the combination of the active principle expressed by the transgenic plant and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B.

Table 28: A method of controlling pests comprising the application of Ti-435 to transgenic rice, wherein the combination of the active principle expressed by the transgenic plant and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B.

Table 29: A method of controlling pests comprising the application of Ti-435 to transgenic potatoes, wherein the combination of the active principle expressed by the transgenic plant and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B.

Table 30: A method of controlling pests comprising the application of Ti-435 to transgenic brassica, wherein the combination of the active principle expressed by the transgenic plant and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B.

Table 31: A method of controlling pests comprising the application of Ti-435 to transgenic tomatoes, wherein the combination of the active principle expressed by the transgenic plant

and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B.

Table 32: A method of controlling pests comprising the application of Ti-435 to transgenic cucurbits, wherein the combination of the active principle expressed by the transgenic plant and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B.

Table 33: A method of controlling pests comprising the application of Ti-435 to transgenic soybeans, wherein the combination of the active principle expressed by the transgenic plant and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B.

Table 34: A method of controlling pests comprising the application of Ti-435 to transgenic maize, wherein the combination of the active principle expressed by the transgenic plant and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B.

Table 35: A method of controlling pests comprising the application of Ti-435 to transgenic wheat, wherein the combination of the active principle expressed by the transgenic plant and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B.

Table 36: A method of controlling pests comprising the application of Ti-435 to transgenic bananas, wherein the combination of the active principle expressed by the transgenic plant and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B.

Table 37: A method of controlling pests comprising the application of Ti-435 to transgenic citrus trees, wherein the combination of the active principle expressed by the transgenic plant and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B.

Table 38: A method of controlling pests comprising the application of Ti-435 to transgenic pome fruit trees, wherein the combination of the active principle expressed by the transgenic plant and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B.

Table 39: A method of controlling pests comprising the application of thiacloprid to transgenic cotton, wherein the combination of the active principle expressed by the transgenic plant and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B.

Table 40: A method of controlling pests comprising the application of thiacloprid to transgenic rice, wherein the combination of the active principle expressed by the transgenic plant and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B.

Table 41: A method of controlling pests comprising the application of thiacloprid to transgenic potatoes, wherein the combination of the active principle expressed by the transgenic plant and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B.

Table 42: A method of controlling pests comprising the application of thiacloprid to transgenic brassica, wherein the combination of the active principle expressed by the transgenic plant and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B.

Table 43: A method of controlling pests comprising the application of thiacloprid to transgenic tomatoes, wherein the combination of the active principle expressed by the transgenic plant and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B.

Table 44: A method of controlling pests comprising the application of thiacloprid to transgenic cucurbits, wherein the combination of the active principle expressed by the transgenic plant and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B.

Table 45: A method of controlling pests comprising the application of thiacloprid to transgenic soybeans, wherein the combination of the active principle expressed by the transgenic plant and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B.

Table 46: A method of controlling pests comprising the application of thiacloprid to transgenic maize, wherein the combination of the active principle expressed by the

transgenic plant and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B.

Table 47: A method of controlling pests comprising the application of thiacloprid to transgenic wheat, wherein the combination of the active principle expressed by the transgenic plant and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B.

Table 48: A method of controlling pests comprising the application of thiacloprid to transgenic bananas, wherein the combination of the active principle expressed by the transgenic plant and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B.

Table C:

Abbreviations:

Acetyl-COA Carboxylase: ACCase

Acetolactate Synthase: ALS

Hydroxyphenylpyruvat dioxygenase: HPPD

Inhibition of protein synthesis: IPS

Hormone mimic: HO

Glutamine Synthetase: GS

Protoporphyrinogen oxidase: PROTOX

5-Enolpyruvyl-3-Phosphoshikimate Synthase: EPSPS

	Principle	Tolerant to	Crop
C.1	ALS	Sulfonylureas etc.***	Cotton
C.2	ALS	Sulfonylureas etc. ***	Rice
C.3	ALS	Sulfonylureas etc. ***	Brassica
C.4	ALS	Sulfonylureas etc. ***	Potatoes
C.5	ALS	Sulfonylureas etc. ***	Tomatoes
C.6	ALS	Sulfonylureas etc. ***	Cucurbits
C.7	ALS	Sulfonylureas etc. ***	Soybeans
C.8	ALS	Sulfonylureas etc. ***	Maize
C.9	ALS	Sulfonylureas etc. ***	Wheat
C.10	ALS	Sulfonylureas etc. ***	pome fruit

	Principle	Tolerant to	Crop
C.11	ALS	Sulfonylureas etc. ***	stone fruit
C.12	ALS	Sulfonylureas etc. ***	citrus
C.13	ACCase	+++	Cotton
C.14	ACCase	+++	Rice
C.15	ACCase	+++	Brassica
C.16	ACCase	+++	Potatoes
C.17	ACCase	+++	Tomatoes
C.18	ACCase	+++	Cucurbits
C.19	ACCase	+++	Soybeans
C.20	ACCase	+++	Maize
C.21	ACCase	+++	Wheat
C.22	ACCase	+++	pome fruit
C.23	ACCase	+++	stone fruit
C.24	ACCase	+++	citrus
C.25	HPPD	Isoxaflutol, Isoxachlotol, Sulcotrion, Mesotrion	Cotton
C.26	HPPD	Isoxaflutol, Isoxachlotol, Sulcotrion, Mesotrion	Rice
C.27	HPPD	Isoxaflutol, Isoxachlotol, Sulcotrion, Mesotrion	Brassica
C.28	HPPD	Isoxaflutol, Isoxachlotol, Sulcotrion, Mesotrion	Potatoes
C.29	HPPD	Isoxaflutol, Isoxachlotol, Sulcotrion, Mesotrion	Tomatoes
C.30	HPPD	Isoxaflutol, Isoxachlotol, Sulcotrion, Mesotrion	Cucurbits
C.31	HPPD	Isoxaflutol, Isoxachlotol, Sulcotrion, Mesotrion	Soybeans
C.32	HPPD	Isoxaflutol, Isoxachlotol, Sulcotrion, Mesotrion	Maize
C.33	HPPD	Isoxaflutol, Isoxachlotol, Sulcotrion, Mesotrion	Wheat
C.34	HPPD	Isoxaflutol, Isoxachlotol, Sulcotrion, Mesotrion	pome fruit
C.35	HPPD	Isoxaflutol, Isoxachlotol, Sulcotrion, Mesotrion	stone fruit
C.36	HPPD	Isoxaflutol, Isoxachlotol, Sulcotrion, Mesotrion	citrus
C.37	Nitrilase	Bromoxynil, Ioxynil	Cotton
C.38	Nitrilase	Bromoxynil, Ioxynil	Rice
C.39	Nitrilase	Bromoxynil, Ioxynil	Brassica
C.40	Nitrilase	Bromoxynil, Ioxynil	Potatoes
C.41	Nitrilase	Bromoxynil, Ioxynil	Tomatoes
C.42	Nitrilase	Bromoxynil, Ioxynil	Cucurbits

	Principle	Tolerant to	Crop
C.43	Nitrilase	Bromoxynil, Ioxynil	Soybeans
C.44	Nitrilase	Bromoxynil, Ioxynil	Maize
C.45	Nitrilase	Bromoxynil, Ioxynil	Wheat
C.46	Nitrilase	Bromoxynil, Ioxynil	pome fruit
C.47	Nitrilase	Bromoxynil, Ioxynil	stone fruit
C.48	Nitrilase	Bromoxynil, Ioxynil	citrus
C.49	IPS	Chloroactanilides &&&	Cotton
C.50	IPS	Chloroactanilides &&&	Rice
C.51	IPS	Chloroactanilide &&&s	Brassica
C.52	IPS	Chloroactanilides &&&	Potatoes
C.53	IPS	Chloroactanilides &&&	Tomatoes
C.54	IPS	Chloroactanilides &&&	Cucurbits
C.55	IPS	Chloroactanilides &&&	Soybeans
C.56	IPS	Chloroactanilides &&&	Maize
C.57	IPS	Chloroactanilides &&&	Wheat
C.58	IPS	Chloroactanilides &&&	pome fruit
C.59	IPS	Chloroactanilides &&&	stone fruit
C.60	IPS	Chloroactanilides &&&	citrus
C.61	HOM	2,4-D, Mecoprop-P	Cotton
C.62	HOM	2,4-D, Mecoprop-P	Rice
C.63	HOM	2,4-D, Mecoprop-P	Brassica
C.64	HOM	2,4-D, Mecoprop-P	Potatoes
C.65	HOM	2,4-D, Mecoprop-P	Tomatoes
C.66	HOM	2,4-D, Mecoprop-P	Cucurbits
C.67	HOM	2,4-D, Mecoprop-P	Soybeans
C.68	HOM	2,4-D, Mecoprop-P	Maize
C.69	HOM	2,4-D, Mecoprop-P	Wheat
C.70	HOM	2,4-D, Mecoprop-P	pome fruit
C.71	HOM	2,4-D, Mecoprop-P	stone fruit
C.72	HOM	2,4-D, Mecoprop-P	citrus
C.73	PROTOX	Protox inhibitors ///	Cotton
C.74	PROTOX	Protox inhibitors ///	Rice

	Principle	Tolerant to	Crop
C.75	PROTOX	Protox inhibitors ///	Brassica
C.76	PROTOX	Protox inhibitors ///	Potatoes
C.77	PROTOX	Protox inhibitors ///	Tomatoes
C.78	PROTOX	Protox inhibitors ///	Cucurbits
C.79	PROTOX	Protox inhibitors ///	Soybeans
C.80	PROTOX	Protox inhibitors ///	Maize
C.81	PROTOX	Protox inhibitors ///	Wheat
C.82	PROTOX	Protox inhibitors ///	pome fruit
C.83	PROTOX	Protox inhibitors ///	stone fruit
C.84	PROTOX	Protox inhibitors ///	citrus
C.85	EPSPS	Glyphosate and /or Sulphosate	Cotton
C.86	EPSPS	Glyphosate and /or Sulphosate	Rice
C.87	EPSPS	Glyphosate and /or Sulphosate	Brassica
C.88	EPSPS	Glyphosate and /or Sulphosate	Potatoes
C.89	EPSPS	Glyphosate and /or Sulphosate	Tomatoes
C.90	EPSPS	Glyphosate and /or Sulphosate	Cucurbits
C.91	EPSPS	Glyphosate and /or Sulphosate	Soybeans
C.92	EPSPS	Glyphosate and /or Sulphosate	Maize
C.93	EPSPS	Glyphosate and /or Sulphosate	Wheat
C.94	EPSPS	Glyphosate and /or Sulphosate	pome fruit
C.95	EPSPS	Glyphosate and /or Sulphosate	stone fruit
C.96	EPSPS	Glyphosate and /or Sulphosate	citrus
C.97	GS	Gluphosinate and /or Bialaphos	Cotton
C.98	GS	Gluphosinate and /or Bialaphos	Rice
C.99	GS	Gluphosinate and /or Bialaphos	Brassica
C.100	GS	Gluphosinate and /or Bialaphos	Potatoes
C.101	GS	Gluphosinate and /or Bialaphos	Tomatoes
C.102	GS	Gluphosinate and /or Bialaphos	Cucurbits
C.103	GS	Gluphosinate and /or Bialaphos	Soybeans
C.104	GS	Gluphosinate and /or Bialaphos	Maize
C.105	GS	Gluphosinate and /or Bialaphos	Wheat
C.106	GS	Gluphosinate and /or Bialaphos	pome fruit

	Principle	Tolerant to	Crop
C.107	GS	Gluphosinate and /or Bialaphos	stone fruit
C.108	GS	Gluphosinate and /or Bialaphos	citrus

*** Included are Sulfonylureas, Imidazolinones, Triazolopyrimidines, Dimethoxypyrimidines and N-Acylsulfonamides:
Sulfonylureas such as Chlorsulfuron, Chlorimuron, Ethamethsulfuron, Metsulfuron, Primisulfuron, Prosulfuron, Triasulfuron, Cinosulfuron, Trifusulfuron, Oxasulfuron, Bensulfuron, Tribenuron, ACC 322140, Fluzasulfuron, Ethoxysulfuron, Fluzasdulfuron, Nicosulfuron, Rimsulfuron, Thifensulfuron, Pyrazosulfuron, Clopyrasulfuron, NC 330, Azimsulfuron, Imazosulfuron, Sulfosulfuron, Amidosulfuron, Flupyrsulfuron, CGA 362622

Imidazolinones such as Imazamethabenz, Imazaquin, Imazamethypyr, Imazethapyr, Imazapyr and Imazamox;

Triazolopyrimidines such as DE 511, Flumetsulam and Chloransulam;

Dimethoxypyrimidines such as Pyrithiobac, Pyriminobac, Bispyribac and Pyribenzoxim.

+++ Tolerant to Diclofop-methyl, Fluazifop-P-butyl, Haloxyfop-P-methyl, Haloxyfop-P-ethyl, Quizalafop-P-ethyl , clodinafop propargyl, fenoxaprop - -ethyl, - Tepraloxydim, Alloxydim, Sethoxydim, Cycloxydim, Cloproxydim, Tralkoxydim, Butoxydim, Caloxydim, Clefoxydim, Clethodim.

&&& Chloroacetanilides such as Alachlor Acetochlor, Dimethenamid

/// Protox inhibitors: For instance diphenyethers such as Acifluorfen, Aclonifen, Bifenox, Chlornitrofen, Ethoxyfen, Fluoroglycofen, Fomesafen, Lactofen, Oxyfluorfen; Imides such as Azafenidin, Carfentrazone-ethyl, Cinidon-ethyl, Flumiclorac-pentyl, Flumioxazin, Fluthiacet-methyl, Oxadiargyl, Oxadiazon, Pentoxazone, Sulfentrazone, Imides and others,such as Flumipropyn, Flupropacil, Nipyraclofen and Thidiazimin; and further Fluazolate and Pyraflufen-ethyl

Biological Examples

Table 49: A method of controlling representatives of the genus Adoxophyes comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the

combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 50: A method of controlling representatives of the genus Agrotis comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 51: A method of controlling Alabama argillaceae comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 52: A method of controlling Anticarsia gemmatalis comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 53: A method of controlling representatives of the genus Chilo comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 54: A method of controlling Clysia ambiguella comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 55: A method of controlling representatives of the genus Cnephalocrocis comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 56: A method of controlling Crocidolomia binotalis comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 57: A method of controlling representatives of the genus Cydia comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 58: A method of controlling Diparopsis castanea comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 59: A method of controlling representatives of the genus Earias comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 60: A method of controlling representatives of the genus Ephestia comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 61: A method of controlling representatives of the genus Heliothis comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 62: A method of controlling Hellula undalis comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 63: A method of controlling Keiferia lycopersicella comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 64: A method of controlling Leucoptera scitella comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the

active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 65: A method of controlling representatives of the genus Lithocollethis comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 66: A method of controlling Lobesia botrana comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 67: A method of controlling Ostrinia nubilalis comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 68: A method of controlling representatives of the genus Pandemis comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 69: A method of controlling Pectinophora gossypiella comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 70: A method of controlling Phyllocnistis citrella comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 71: A method of controlling representatives of the genus Pieris comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 72: A method of controlling Plutella xylostella comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 73: A method of controlling representatives of the genus Scirpophaga comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 74: A method of controlling representatives of the genus Sesamia comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 75: A method of controlling representatives of the genus Sparganothis comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 76: A method of controlling representatives of the genus Spodoptera comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 77: A method of controlling representatives of the genus Tortrix comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 78: A method of controlling Trichoplusia ni comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 79: A method of controlling representatives of the genus Agriotes comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the

combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 80: A method of controlling Anthonomus grandis comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 81: A method of controlling representatives of the genus Curculio comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 82: A method of controlling Diabrotica balteata comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 83: A method of controlling representatives of the genus Leptinotarsa comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 84: A method of controlling representatives of the genus Lissorhoptrus comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 85: A method of controlling representatives of the genus Otiorhynchus comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 86: A method of controlling representatives of the genus Aleurothrixus comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 87: A method of controlling representatives of the genus Aleyrodes comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 88: A method of controlling representatives of the genus Aonidiella comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 89: A method of controlling representatives of the family Aphididae comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 90: A method of controlling representatives of the genus Aphis comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 91: A method of controlling Bemisia tabaci comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 92: A method of controlling representatives of the genus Empoasca comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 93: A method of controlling representatives of the genus Mycus comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 94: A method of controlling representatives of the genus Nephotettix comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the

combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 95: A method of controlling representatives of the genus Nilaparvata comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 96: A method of controlling representatives of the genus Pseudococcus comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 97: A method of controlling representatives of the genus Psylla comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 98: A method of controlling representatives of the genus Quadraspidiotus comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 99: A method of controlling representatives of the genus Schizaphis comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 100: A method of controlling representatives of the genus Trialeurodes comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 101: A method of controlling representatives of the genus Lyriomyza comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 102: A method of controlling representatives of the genus Oscinella comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 103: A method of controlling representatives of the genus Phorbia comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 104: A method of controlling representatives of the genus Frankliniella comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 105: A method of controlling representatives of the genus Thrips comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 106: A method of controlling Scirtothrips aurantii comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 107: A method of controlling representatives of the genus Aceria comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 108: A method of controlling representatives of the genus Aculus comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 109: A method of controlling representatives of the genus Brevipalpus comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the

combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 110: A method of controlling representatives of the genus Panonychus comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 111: A method of controlling representatives of the genus Phyllocoptruta comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 112: A method of controlling representatives of the genus Tetranychus comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 113: A method of controlling representatives of the genus Heterodera comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 114: A method of controlling representatives of the genus Meloidogyne comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 115: A method of controlling Mamestra brassica comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 116: A method of controlling representatives of the genus Adoxophyes comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 117: A method of controlling representatives of the genus Agrotis comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 118: A method of controlling Alabama argillaceae comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 119: A method of controlling Anticarsia gemmatalis comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 120: A method of controlling representatives of the genus Chilo comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 121: A method of controlling Clysia ambiguella comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 122: A method of controlling representatives of the genus Cnephalocrocis comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 123: : A method of controlling Crocidolomia binotalis comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 124: A method of controlling representatives of the genus Cydia comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the

combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 125: A method of controlling Diparopsis castanea comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 126: A method of controlling representatives of the genus Earias comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 127: A method of controlling representatives of the genus Ephestia comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 128: A method of controlling representatives of the genus Heliothis of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 129: A method of controlling Hellula undalis comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 130: A method of controlling Keiferia lycopersicella comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 131: A method of controlling Leucoptera scitella comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 132: A method of controlling representatives of the genus Lithocollethis comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 133: A method of controlling Lobesia botrana comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 134: A method of controlling Ostrinia nubilalis comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 135: A method of controlling representatives of the genus Pandemis comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 136: A method of controlling Pectinophora gossypiella comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 137: A method of controlling Phyllocnistis citrella comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 138: A method of controlling representatives of the genus Pieris comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 139: A method of controlling Plutella xylostella comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the

active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 140: A method of controlling representatives of the genus Scirpophaga comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 141: A method of controlling representatives of the genus Sesamia comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 142: A method of controlling representatives of the genus Sparganothis comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 143: A method of controlling representatives of the genus Spodoptera comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 144: A method of controlling representatives of the genus Tortrix comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 145: A method of controlling Trichoplusia ni comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 146: A method of controlling representatives of the genus Agriotes comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 147: A method of controlling Anthonomus grandis comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 148: A method of controlling representatives of the genus Curculio comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 149: A method of controlling Diabrotica balteata comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 150: A method of controlling representatives of the genus Leptinotarsa comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 151: A method of controlling representatives of the genus Lissorhoptrus comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 152: A method of controlling representatives of the genus Otiorhynchus comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 153: A method of controlling representatives of the genus Aleurothrixus comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 154: A method of controlling representatives of the genus Aleyrodes comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the

combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 155: A method of controlling representatives of the genus Aonidiella comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 156: A method of controlling representatives of the family Aphididae comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 157: A method of controlling representatives of the genus Aphis comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 158: A method of controlling Bemisia tabaci comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 159: A method of controlling representatives of the genus Empoasca comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 160: A method of controlling representatives of the genus Mycus comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 161: A method of controlling representatives of the genus Nephotettix comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 162: A method of controlling representatives of the genus Nilaparvata comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 163: A method of controlling representatives of the genus Pseudococcus comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 164: A method of controlling representatives of the genus Psylla comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 165: A method of controlling representatives of the genus Quadraspidiotus comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 166: A method of controlling representatives of the genus Schizaphis comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 167: A method of controlling representatives of the genus Trialeurodes comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 168: A method of controlling representatives of the genus Lyriomyza comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 169: A method of controlling representatives of the genus Oscinella comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the

combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 170: A method of controlling representatives of the genus Phorbia comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 171: A method of controlling representatives of the genus Frankliniella comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 172: A method of controlling representatives of the genus Thrips comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 173: A method of controlling Scirtothrips aurantii comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 174: A method of controlling representatives of the genus Aceria comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 175: A method of controlling representatives of the genus Aculus comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 176: A method of controlling representatives of the genus Brevipalpus comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 177: A method of controlling representatives of the genus Panonychus comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 178: A method of controlling representatives of the genus Phyllocoptruta comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 179: A method of controlling representatives of the genus Tetranychus comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 180: A method of controlling representatives of the genus Heterodera comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 181: A method of controlling representatives of the genus Meloidogyne comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 182: A method of controlling representatives of the genus Adoxophyes comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 183: A method of controlling representatives of the genus Agrotis comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 184: A method of controlling Alabama argillaceae comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle

expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 185: A method of controlling Anticarsia gemmatalis comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 186: A method of controlling representatives of the genus Chilo comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 187: A method of controlling Clysia ambiguella comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 188: A method of controlling Crocidolomia binotalis comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 189: A method of controlling representatives of the genus Cydia comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 190: A method of controlling Diparopsis castanea comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 191: A method of controlling representatives of the genus Earias comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 192: A method of controlling representatives of the genus Ephestia comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 193: A method of controlling representatives of the genus Heliothis of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 194: A method of controlling Hellula undalis comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 195: A method of controlling Keiferia lycopersicella comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 196: A method of controlling Leucoptera scitella comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 197: A method of controlling representatives of the genus Lithocollethis comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 198: A method of controlling Lobesia botrana comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 199: A method of controlling Ostrinia nubilalis comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle

expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 200: A method of controlling representatives of the genus Pandemis comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 201: A method of controlling Pectinophora gossypiella comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 202: A method of controlling Phyllocnistis citrella comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 203: A method of controlling representatives of the genus Pieris comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 204: A method of controlling Plutella xylostella comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 205: A method of controlling representatives of the genus Scirpophaga comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 206: A method of controlling representatives of the genus Sesamia comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 207: A method of controlling representatives of the genus Sparganothis comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 208: A method of controlling representatives of the genus Spodoptera comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 209: A method of controlling representatives of the genus Tortrix comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 210: A method of controlling Trichoplusia ni comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 211: A method of controlling representatives of the genus Agriotes comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 212: A method of controlling Anthonomus grandis comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 213: A method of controlling representatives of the genus Curculio comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 214: A method of controlling Diabrotica balteata comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle

expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 215: A method of controlling representatives of the genus Leptinotarsa comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 216: A method of controlling representatives of the genus Lissorhoptrus comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 217: A method of controlling representatives of the genus Otiorhynchus comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 218: A method of controlling representatives of the genus Aleurothrixus comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 219: A method of controlling representatives of the genus Aleyrodes comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 220: A method of controlling representatives of the genus Aonidiella comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 221: A method of controlling representatives of the family Aphididae comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 222: A method of controlling representatives of the genus Aphis comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 223: A method of controlling Bemisia tabaci comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 224: A method of controlling representatives of the genus Empoasca comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 225: A method of controlling representatives of the genus Mycus comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 226: A method of controlling representatives of the genus Nephotettix comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 227: A method of controlling representatives of the genus Nilaparvata comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 228: A method of controlling representatives of the genus Pseudococcus comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 229: A method of controlling representatives of the genus Psylla comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of

the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 230: A method of controlling representatives of the genus Quadraspidiotus comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 231: A method of controlling representatives of the genus Schizaphis comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 232: A method of controlling representatives of the genus Trialeurodes comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 233: A method of controlling representatives of the genus Lyriomyza comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 234: A method of controlling representatives of the genus Oscinella comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 235: A method of controlling representatives of the genus Phorbia comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 236: A method of controlling representatives of the genus Frankliniella comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 237: A method of controlling representatives of the genus Thrips comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 238: A method of controlling Scirtothrips aurantii comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 239: A method of controlling representatives of the genus Aceria comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 240: A method of controlling representatives of the genus Aculus comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 241: A method of controlling representatives of the genus Brevipalpus comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 242: A method of controlling representatives of the genus Panonychus comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 243: A method of controlling representatives of the genus Phyllocoptruta comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 244: A method of controlling representatives of the genus Tetranychus comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination

of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 245: A method of controlling representatives of the genus Heterodera comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 246: A method of controlling representatives of the genus Meloidogyne comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 247: A method of controlling Mamestra brassica comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Example B1: Action against Anthonomus grandis adults, Spodoptera littoralis or Heliothis virescens

Young transgenic cotton plants which express the δ-endotoxin CryIIIA are sprayed with an aqueous emulsion spray mixture comprising 100, 50, 10, 5, 1 ppm of imidacloprid respectively. After the spray coating has dried on, the cotton plants are populated with 10 adult Anthonomus grandis, 10 Spodoptera littoralis larvae or 10 Heliothis virescens larvae respectively and introduced into a plastic container. Evaluation takes place 3 to 10 days later. The percentage reduction in population, or the percentage reduction in feeding damage (% action), is determined by comparing the number of dead beetles and the feeding damage on the transgenic cotton plants with that of non-transgenic cotton plants which have been treated with an emulsion spray mixture comprising imidacloprid and conventional CryIIIA-toxin at a concentration of in each case 100, 50, 10, 5, 1 ppm respectively.

In this test, the control of the tested insects in the transgenic plant is superior to the control on the non-transgenic plant.

Example B2: Action against Anthonomus grandis adults, Spodoptera littoralis or Heliothis virescens

Young transgenic cotton plants which express the δ-endotoxin CryIIIA are sprayed with an aqueous emulsion spray mixture comprising 100, 50, 10, 5, 1 ppm of thiamethoxam respectively. After the spray coating has dried on, the cotton plants are populated with 10 adult Anthonomus grandis, 10 Spodoptera littoralis larvae or 10 Heliothis virescens larvae respectively and introduced into a plastic container. Evaluation takes place 3 to 10 days later. The percentage reduction in population, or the percentage reduction in feeding damage (% action), is determined by comparing the number of dead beetles and the feeding damage on the transgenic cotton plants with that of non-transgenic cotton plants which have been treated with an emulsion spray mixture comprising thiamethoxam and conventional CryIIIA-toxin at a concentration of in each case 100, 50, 10, 5, 1 ppm respectively.

In this test, the control of the tested insects in the transgenic plant is superior, while it is insufficient in the non-transgenic plant.

Example B3: Action against Anthonomus grandis adults, Spodoptera littoralis or Heliothis virescens

Young transgenic cotton plants which express the δ-endotoxin CryIIIA are sprayed with an aqueous emulsion spray mixture comprising 100, 50, 10, 5, 1 ppm of Ti-435 respectively. After the spray coating has dried on, the cotton plants are populated with 10 adult Anthonomus grandis, 10 Spodoptera littoralis larvae or 10 Heliothis virescens larvae respectively and introduced into a plastic container. Evaluation takes place 3 to 10 days later. The percentage reduction in population, or the percentage reduction in feeding damage (% action), is determined by comparing the number of dead beetles and the feeding damage on the transgenic cotton plants with that of non-transgenic cotton plants which have been treated with an emulsion spray mixture comprising Ti-435 and conventional CryIIIA-toxin at a concentration of in each case 100, 50, 10, 5, 1 ppm respectively.

In this test, the control of the tested insects in the transgenic plant is superior, while it is insufficient in the non-transgenic plant.

Example B4: Action against Anthonomus grandis adults, Spodoptera littoralis or Heliothis virescens

Young transgenic cotton plants which express the δ-endotoxin Cryla(c) are sprayed with an aqueous emulsion spray mixture comprising 100, 50, 10, 5, 1 ppm of Ti-435 respectively. After the spray coating has dried on, the cotton plants are populated with 10 adult Anthonomus grandis, 10 Spodoptera littoralis larvae or 10 Heliothis virescens larvae

respectively and introduced into a plastic container. Evaluation takes place 3 to 10 days later. The percentage reduction in population, or the percentage reduction in feeding damage (% action), is determined by comparing the number of dead beetles and the feeding damage on the transgenic cotton plants with that of non-transgenic cotton plants which have been treated with an emulsion spray mixture comprising Ti-435 and conventional CryIIIA-toxin at a concentration of in each case 100, 50, 10, 5, 1 ppm respectively.

In this test, the control of the tested insects in the transgenic plant is superior, while it is insufficient in the non-transgenic plant.

Example B5: Action against Anthonomus grandis adults, Spodoptera littoralis or Heliothis virescens

Young transgenic cotton plants which express the δ-endotoxin Cryla(c) are sprayed with an aqueous emulsion spray mixture comprising 100, 50, 10, 5, 1 ppm of thiamethoxam respectively. After the spray coating has dried on, the cotton plants are populated with 10 adult Anthonomus grandis, 10 Spodoptera littoralis larvae or 10 Heliothis virescens larvae respectively and introduced into a plastic container. Evaluation takes place 3 to 10 days later. The percentage reduction in population, or the percentage reduction in feeding damage (% action), is determined by comparing the number of dead beetles and the feeding damage on the transgenic cotton plants with that of non-transgenic cotton plants which have been treated with an emulsion spray mixture comprising thiamethoxam and conventional CryIIIA-toxin at a concentration of in each case 100, 50, 10, 5, 1 ppm respectively.

In this test, the control of the tested insects in the transgenic plant is superior, while it is insufficient in the non-transgenic plant.

Example B6: Action against Anthonomus grandis adults, Spodoptera littoralis or Heliothis virescens

Young transgenic cotton plants which express the δ-endotoxin Cryla(c) are sprayed with an aqueous emulsion spray mixture comprising 100, 50, 10, 5, 1 ppm of imidacloprid respectively. After the spray coating has dried on, the cotton plants are populated with 10 adult Anthonomus grandis, 10 Spodoptera littoralis larvae or 10 Heliothis virescens larvae respectively and introduced into a plastic container. Evaluation takes place 3 to 10 days later. The percentage reduction in population, or the percentage reduction in feeding damage (% action), is determined by comparing the number of dead beetles and the feeding damage on the transgenic cotton plants with that of non-transgenic cotton plants

which have been treated with an emulsion spray mixture comprising imidacloprid conventional CryIIIA-toxin at a concentration of in each case 100, 50, 10, 5, 1 ppm respectively.

In this test, the control of the tested insects in the transgenic plant is superior, while it is insufficient in the non-transgenic plant.

Example B7: Action against Ostrinia nubilalis, Spodoptera spp. or Heliothis spp.

A plot (a) planted with maize cv. KnockOut® and an adjacent plot (b) of the same size which is planted with conventional maize, both showing natural infestation with Ostrinia nubilalis, Spodoptera spp. or Heliothis, are sprayed with an aqueous emulsion spray mixture comprising 200, 100, 50, 10, 5, 1ppm of Ti-435. Immediately afterwards, plot (b) is treated with an emulsion spray mixture comprising 200, 100, 50, 10, 5, 1 ppm of the endotoxin expressed by KnockOut®. Evaluation takes place 6 days later. The percentage reduction in population (% action) is determined by comparing the number of dead pests on the plants of plot (a) with that on the plants of plot (b).

Improved control of Ostrinia nubilalis, Spodoptera spp. or Heliothis is observed on the plants of plot (a), while plot (b) shows a control level of not over 60%.

Example B8: Action against Ostrinia nubilalis, Spodoptera spp. or Heliothis spp.

A plot (a) planted with maize cv. KnockOut® and an adjacent plot (b) of the same size which is planted with conventional maize, both showing natural infestation with Ostrinia nubilalis, Spodoptera spp. or Heliothis, are sprayed with an aqueous emulsion spray mixture comprising 200, 100, 50, 10, 5, 1ppm of thiamethoxam. Immediately afterwards, plot (b) is treated with an emulsion spray mixture comprising 200, 100, 50, 10, 5, 1 ppm of the endotoxin expressed by KnockOut®. Evaluation takes place 6 days later. The percentage reduction in population (% action) is determined by comparing the number of dead pests on the plants of plot (a) with that on the plants of plot (b).

Improved control of Ostrinia nubilalis, Spodoptera spp. or Heliothis is observed on the plants of plot (a), while plot (b) shows a control level of not over 60%.

Example B9: Action against Ostrinia nubilalis, Spodoptera spp. or Heliothis spp.

A plot (a) planted with maize cv. KnockOut® and an adjacent plot (b) of the same size which is planted with conventional maize, both showing natural infestation with Ostrinia nubilalis, Spodoptera spp. or Heliothis, are sprayed with an aqueous emulsion spray mixture

comprising 200, 100, 50, 10, 5, 1ppm of imidacloprid. Immediately afterwards, plot (b) is treated with an emulsion spray mixture comprising 200, 100, 50, 10, 5, 1 ppm of the endotoxin expressed by KnockOut®. Evaluation takes place 6 days later. The percentage reduction in population (% action) is determined by comparing the number of dead pests on the plants of plot (a) with that on the plants of plot (b).

Improved control of Ostrinia nubilalis, Spodoptera spp. or Heliothis spp. is observed on the plants of plot (a), while plot (b) shows a control level of not over 60%.

Example B10: Action against Diabrotica balteata

A plot (a) planted with maize seedlings cv. KnockOut® and an adjacent plot (b) of the same size which is planted with conventional maize are sprayed with an aqueous emulsion of a spray mixture comprising 400 ppm thiamethoxam. Immediately afterwards, plot (b) is treated with an emulsion spray mixture comprising 400 ppm of the endotoxin expressed by KnockOut®. After the spray coating has dried on, the seedlings are populated with 10 Diabrotica balteata larvae in the second stage and transferred to a plastic container. The test is evaluated 6 days later. The percentage reduction in population (% action) is determined by comparing the number of dead pests on the plants of plot (a) with that on the plants of plot (b).

Improved control of Diabrotica balteata is observed on the plants of plot (a), while plot (b) shows a control level of not over 60%.

Example B11: Action against Aphis gossypii

Cotton seedlings on a plot (a) expressing the δ-endotoxin CryIIIa on a plot (a) and conventional cotton seedlings on a plot (b) are infected with Aphis gossypi and subsequently sprayed with a spray mixture comprising 400 ppm thiamethoxam. Immediately afterwards, plot (b) is treated with an emulsion spray mixture comprising 400 ppm of the δ-endotoxin CryIIIa. The seedlings of plot (a) and (b) are then incubated at 20°C. The test is evaluated after 3 and 6 days.

The percentage reduction in population (% action) is determined by comparing the number of dead pests on the plants of plot (a) with that on the plants of plot (b). Improved control of Aphis gossypi is observed on the plants of plot (a), while plot (b) shows a control level of not over 60%.

Example B12: Action against Frankliniella occidentalis

Cotton seedlings expressing the δ-endotoxin CryIIIa on a plot (a) and conventional cotton seedlings on a plot (b) are infected with Frankliniella occidentalis and subsequently sprayed with a spray mixture comprising 400 ppm thiamethoxam. Immediately afterwards, plot (b) is treated with an emulsion spray mixture comprising 400 ppm of the δ-endotoxin CryIIIa. The seedlings of plot (a) and (b) are then incubated at 20°C. The test is evaluated after 3 and 6 days.

The percentage reduction in population (% action) is determined by comparing the number of dead pests on the plants of plot (a) with that on the plants of plot (b). Improved control of Frankliniella occidentalis is observed on the plants of plot (a), while plot (b) shows a control level of not over 60%.

Example B13: Action against Aphis gossypii

Cotton seedlings expressing the δ-endotoxin CryIA(c) on a plot (a) and conventional cotton seedlings on a plot (b) are infected with Aphis gossypii and subsequently sprayed with a spray mixture comprising 400 ppm thiamethoxam. Immediately afterwards, plot (b) is treated with an emulsion spray mixture comprising 400 ppm of the δ-endotoxin CryIIIa. The seedlings of plot (a) and (b) are then incubated at 20°C. The test is evaluated after 3 and 6 days.

The percentage reduction in population (% action) is determined by comparing the number of dead pests on the plants of plot (a) with that on the plants of plot (b). Improved control of Aphis gossypii is observed on the plants of plot (a), while plot (b) shows a control level of not over 60%.

Example B14: Action against Frankliniella occidentalis

Cotton seedlings expressing the δ-endotoxin CryIa(c) on a plot (a) and conventional cotton seedlings on a plot (b) are infected with Frankliniella occidentalis and subsequently sprayed with a spray mixture comprising 400 ppm thiamethoxam. Immediately afterwards, plot (b) is treated with an emulsion spray mixture comprising 400 ppm of the δ-endotoxin CryIa(c). The seedlings of plot (a) and (b) are then incubated at 20°C. The test is evaluated after 3 and 6 days.

The percentage reduction in population (% action) is determined by comparing the number of dead pests on the plants of plot (a) with that on the plants of plot (b). Improved control of

Frankliniella occidentalis is observed on the plants of plot (a), while plot (b) shows a control level of not over 60%.

Example B15: Action against Nephotettix cincticeps

Rice plants on a plot (a) expressing the δ-endotoxin CryIA(b) and conventional rice plants on a plot (b) are sprayed with a spray mixture comprising 400 ppm thiamethoxam. Immediately afterwards, plot (b) is treated with an emulsion spray mixture comprising 400 ppm of the δ-endotoxin CryIA(b). After the spray coating has dried on, the plants are infected with Nephotettix cincticeps of the 2nd and 3rd stages. The seedlings of plot (a) and (b) are then incubated at 20°C. The test is evaluated after 21 days.

The percentage reduction in population (% action) is determined by comparing the number of dead pests on the plants of plot (a) with that on the plants of plot (b). Improved control of Nephotettix cincticeps is observed on the plants of plot (a), while plot (b) shows a control level of not over 60%.

Example B16: Action against Nephotettix cincticeps (systemic)

Rice plants expressing the δ-endotoxin Cryla(b) are planted in a in pot (A) and conventional ice plants are planted in a pot (B). Pot (A) is placed in an aqueous emulsion containing 400 ppm thiamethoxam, whereas plot (B) is placed in a pot containing 400 ppm thiamethoxam and 400 ppm of the δ-endotoxin CryI(b). The plants are subsequently infected with Nephotettix cincticeps larvae of the second and third stage. The test is evaluated after 6 days.

The percentage reduction in population (% action) is determined by comparing the number of dead pests on the plants of pot (A) with that on the plants of pot (B). Improved control of Nephotettix cincticeps is observed on the plants of pot (A), while pot (B) shows a control level of not over 60%.

Example B17: Action against Nilaparvata lugens

Rice plants on a plot (a) expressing the δ-endotoxin CryIA(b) and conventional rice plants on a plot (b) are infected with Nilaparvata lugens, subsequently sprayed with a spray mixture comprising 400 ppm thiamethoxam. Immediately afterwards, plot (b) is treated with an emulsion spray mixture comprising 400 ppm of the δ-endotoxin CryIA(b). The seedlings of plot (a) and (b) are then incubated at 20°C. The test is evaluated after 21 days.

The percentage reduction in population (% action) is determined by comparing the number of dead pests on the plants of plot (a) with that on the plants of plot (b). Improved control of Nilaparvata lugens is observed on the plants of plot (a), while plot (b) shows a control level of not over 60%.

Example B18: Action against Nilaparvata lugens (systemic)

Rice plants expressing the δ-endotoxin CrylA(b) are planted in a in pot (A) and conventional rice plants are planted in a pot (B). Pot (A) is placed in an aqueous emulsion containing 400 ppm thiamethoxam, whereas plot (B) is place in a pot copntaining 400 ppm thiamethoxam and 400 ppm of the δ-endotoxin CrylA(b). The plants are subsequently infected with Nilaparvata lugens larvae of the second and third stage. The test is evaluated after 6 days.

The percentage reduction in population (% action) is determined by comparing the number of dead pests on the plants of pot (A) with that on the plants of pot (B). Improved control of Nephotettix cincticeps is observed on the plants of pot (A), while pot (B) shows a control level of not over 60%.

Example B19: Action against Nephotettix cincticeps

Rice plants on a plot (a) expressing the δ-endotoxin CrylA(c) and conventional rice plants on a plot (b) are sprayed with a spray mixture comprising 400 ppm thiamethoxam. Immediately afterwards, plot (b) is treated with an emulsion spray mixture comprising 400 ppm of the δ-endotoxin CrylA(c). After the spray coating has dried on, the plants are infected with Nephotettix cincticeps of the 2nd and 3rd stages. The seedlings of plot (a) and (b) are then incubated at 20°C. The test is evaluated after 21 days.

The percentage reduction in population (% action) is determined by comparing the number of dead pests on the plants of plot (a) with that on the plants of plot (b). Improved control of Nephotettix cincticeps is observed on the plants of plot (a), while plot (b) shows a control level of not over 60%.

Example B20: Action against Nephotettix cincticeps (systemic)

Rice plants expressing the δ-endotoxin Cryla(c) are planted in a in pot (A) and conventional ice plants are planted in a pot (B). Pot (A) is placed in an aqueous emulsion containing 400 ppm thiamethoxam, whereas plot (B) is placed in a pot containing 400 ppm thiamethoxam and 400 ppm of the δ-endotoxin Cryl(c). The plants are subsequently infected with

Nephotettix cincticeps larvae of the second and third stage. The test is evaluated after 6 days.

The percentage reduction in population (% action) is determined by comparing the number of dead pests on the plants of pot (A) with that on the plants of pot (B). Improved control of Nephotettix cincticeps is observed on the plants of pot (A), while pot (B) shows a control level of not over 60%.

Example B21: Action against Nilaparvata lugens

Rice plants on a plot (a) expressing the δ-endotoxin CryIA(c) and conventional rice plants on a plot (b) are infected with Nilaparvata lugens, subsequently sprayed with a spray mixture comprising 400 ppm thiamethoxam. Immediately afterwards, plot (b) is treated with an emulsion spray mixture comprising 400 ppm of the δ-endotoxin CryIA(c). The seedlings of plot (a) and (b) are then incubated at 20°C. The test is evaluated after 21 days.

The percentage reduction in population (% action) is determined by comparing the number of dead pests on the plants of plot (a) with that on the plants of plot (b). Improved control of Nilaparvata lugens is observed on the plants of plot (a), while plot (b) shows a control level of not over 0%.

Example B22: Action against Nilaparvata lugens (systemic)

Rice plants expressing the δ-endotoxin CryIA(c) are planted in a in pot (A) and conventional rice plants are planted in a pot (B). Pot (A) is placed in an aqueous emulsion containing 400 ppm thiamethoxam, whereas plot (B) is place in a pot copntaining 400 ppm thiamethoxam and 400 ppm of the δ-endotoxin CryIA(c). The plants are subsequently infected with Nilaparvata lugens larvae of the second and third stage. The test is evaluated after 6 days.

The percentage reduction in population (% action) is determined by comparing the number of dead pests on the plants of pot (A) with that on the plants of pot (B). Improved control of Nephotettix cincticeps is observed on the plants of pot (A), while pot (B) shows a control level of not over 60 %.

Patent claims:

1. Method of controlling pests in crops of transgenic useful plants, characterized in that a pesticidal composition comprising a nitroimino- or nitroguanidino-compound in free form or in agrochemically useful salt form as active ingredient and at least one auxiliary is applied to the pests or their environment.

2. Method according to claim 1, characterized in that thiamethoxam is employed.

3. Method according to claim 1, characterized in that imidacloprid is employed.

4. Method according to claim 1, characterized in that the transgenic plant is treated.

5. Method according to any one of claims 1 to 4, characterized in that the transgenic crop of useful plants is maize.

6. Method according to any one of claims 1 to 4, characterized in that the transgenic crop of useful plants is soya beans.

7. Method according to claim 4, characterized in that the propagation material of the transgenic useful plant is treated.

INTERNATIONAL SEARCH REPORT

Inte ,onal Application No
PCT/EP 99/00183

A. CLASSIFICATION OF SUBJECT MATTER
IPC 6 A01N51/00 A01N47/40 //(A01N51/00,63:02,63:00),(A01N47/40, 63:02,63:00)

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
IPC 6 A01N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category °	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
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[X] Further documents are listed in the continuation of box C.

[X] Patent family members are listed in annex.

° Special categories of cited documents :

"A" document defining the general state of the art which is not considered to be of particular relevance

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"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.

"&" document member of the same patent family

Date of the actual completion of the international search	Date of mailing of the international search report
14 May 1999	27/05/1999

Name and mailing address of the ISA	Authorized officer
European Patent Office, P.B. 5818 Patentlaan 2 NL - 2280 HV Rijswijk Tel. (+31-70) 340-2040, Tx. 31 651 epo nl, Fax: (+31-70) 340-3016	Lamers, W

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C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

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INTERNATIONAL SEARCH REPORT

Inte onal Application No
PCT/EP 99/00183

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PCT/EP 99/00183

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